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# **Nitric Oxide Assay Kit**



**Publications** 



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Quantity:	100 tests
Target:	Nitric Oxide (NO)
Application:	Biochemical Assay (BCA)

## **Product Details**

Sample Type:	Cell Lysate, Food, Plasma, Serum, Tissue Lysate, Urine
Specificity:	0.6 μΜ
Characteristics:	Sensitive and accurate. Detection range 0.6 - 200 $\mu$ M in 96-well plate. Rapid and reliable. Using an optimized VCI3 reagent, the time required for reduction of NO3 - toNO2 - is 10 min at 60°C. Simple and high-throughput. The procedure involves mixing sample with three reagents, incubation for 10 min at 60°C and reading the optical density. Can be readily automated to measure thousands of samples per day.
Components:	Reagent A: 12 mL. Reagent B: 500 µL. Reagent C: 12 mL. NaOH: 1 mL. ZnSO4: 1 mL. Standard: 1 mL.
Material not included:	Pipetting devices, eppendorf tubes, eppendorf centrifuge, clear, flat bottomed 96 well plates or cuvettes, plate reader or spectrophotometer and heat block or hot water bath (optional).

# **Target Details**

Target:	Nitric Oxide (NO)	
Alternative Name:	Nitric Oxide (NO Products)	
Target Type:	Anorganic	

#### Target Details

Background:

Quantitative determination of nitric oxide by colorimetric (540nm) method.

Procedure: 40 min.

Nitric oxide (NO) is a reactive radical that plays an important role in many key physiological functions. NO, an oxidation product of arginine by nitric oxide synthase, is involved in host defense and development, activation of regulatory proteins and direct covalent interaction with functional biomolecules. Simple, direct and automation-ready procedures for measuring NO are becoming popular in Research and Drug Discovery. Since NO is oxidized to nitrite and nitrate, it is common practice to quantitate total NO2-/NO3- as a measure for NO level. This Nitric Oxide Assay Kit is designed to accurately measure NO production following reduction of nitrate to nitrite using improved Griess method. The procedure is simple and the time required for sample pretreatment and assay is reduced to as short as 30 min.

## **Application Details**

**Application Notes:** 

Direct Assays: NO in plasma, serum, urine, tissue/cells and foods.

Drug Discovery/Pharmacology: effects of drugs on NO metabolism.

Comment:

Antioxidants and nucleophiles (e.g. beta-mercaptoethanol, glutathione, dithiothreitol and cysteine) may interfere with this assay. Avoid using these compounds during sample preparation.

Protocol:

Sample treatment: tissue or cell samples are homogenized in 1 x PBS (pH 7.4). Centrifuge at 10,000g or higher at 4°C. Use supernatant for NO assay. Samples that need deproteination include serum, plasma, whole blood, cell culture media containing FBS, tissue or cell lysates. Urine and saliva do not need deproteination. Deproteination. Mix 150  $\mu$ L sample with 8  $\mu$ L ZnSO4 in1.5-mL tubes. Vortex and then add 8  $\mu$ L NaOH, votex again and centrifuge 10 min at 14,000 rpm. Transfer 100  $\mu$ L of the clear supernatant to a clean tube. Note: If samples need to be deproteinated, 150  $\mu$ L of each standard should be prepared and also treated with ZnSO4 and NaOH to eliminate the need for a dilution factor.

Procedure using 96-well plate:

- 1. Standards. Prepare 500  $\mu$ L 100  $\mu$ M Premix by mixing 50  $\mu$ L 1.0 mM Standard and 450  $\mu$ L distilled water.
- 2. Reaction. Add 100  $\mu$ L of each sample to separate, labeled eppendorf tubes. (We recommend that samples be measured in at least duplicate). Immediately prior to starting the reaction, prepare enough Working Reagent (WR) for all samples and standards by mixing per reaction tube: 100  $\mu$ L Reagent A, 4  $\mu$ L Reagent B and 100  $\mu$ L Reagent C. Add 200  $\mu$ L of the WR to each

sample and standard tube and incubate for 10 min at 60°C. (Alternatively, the reaction can be run at 37°C for 60 min or RT for 150 min.)

3. Measurement. Briefly centrifuge the reaction tubes to pellet any condensation and transfer  $250 \, \mu L$  of each reaction to separate wells in a 96 well plate. Read OD at  $500-570 \, \text{nm}$  (peak  $540 \, \text{nm}$ ).

Procedure using Cuvette: Prepare standards and samples as described for the 96-well procedure except quadruple (4x) the volumes. After the reaction, transfer 1 mL to a cuvette. Measure OD540nm in the cuvette.

Calculation of Results:

Subtract blank OD (Std 4) from the standard OD values and plot the OD against standard concentrations. Determine the slope using linear regression fitting. Conversions: 1 mg/dL NO equals 333  $\mu$ M, 0.001% or 10 ppm.

Restrictions:

For Research Use only

#### Handling

Storage:

4°C

#### **Publications**

Product cited in:

Guo, Li, Ling, Feng, Xia: "Anthocyanin inhibits high glucose-induced hepatic mtGPAT1 activation and prevents fatty acid synthesis through PKCζ." in: **Journal of lipid research**, Vol. 52, Issue 5, pp. 908-22, (2011) (PubMed).

Bytautiene, Tamayo, Kechichian, Drever, Gamble, Hankins, Saade: "Prepregnancy obesity and sFlt1-induced preeclampsia in mice: developmental programming model of metabolic syndrome." in: **American journal of obstetrics and gynecology**, Vol. 204, Issue 5, pp. 398.e1-8, (2011) (PubMed).

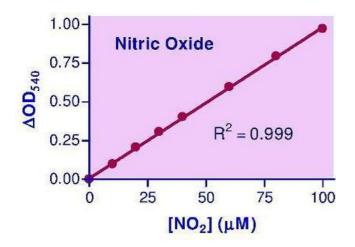
Orban, Palczewska, Palczewski: "Retinyl ester storage particles (retinosomes) from the retinal pigmented epithelium resemble lipid droplets in other tissues." in: **The Journal of biological chemistry**, Vol. 286, Issue 19, pp. 17248-58, (2011) (PubMed).

Uddin, Duy, Cinar, Tesfaye, Tholen, Juengst, Looft, Schellander: "Detection of quantitative trait loci affecting serum cholesterol, LDL, HDL, and triglyceride in pigs." in: **BMC genetics**, Vol. 12, pp. 62, (2011) (PubMed).

Oh, Kim, Jang, Byeon, Ryu, Kim, Ha: "Semipurified fractions from the submerged-culture broth of Agaricus blazei Murill reduce blood glucose levels in streptozotocin-induced diabetic rats." in: **Journal of agricultural and food chemistry**, Vol. 58, Issue 7, pp. 4113-9, (2010) (PubMed).

There are more publications referencing this product on: Product page

# Images



#### **Biochemical Assay**

Image 1.