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Datasheet for ABIN1000284 NAD/NADH Assay Kit

1 Image

11 Publications



Overview

Quantity:	100 tests
Target:	NAD/NADH
Application:	Biochemical Assay (BCA)

Product Details

Sample Type:	Cell Extracts
Specificity:	0.05 μM
Characteristics:	Sensitive and accurate. Detection limit 0.05 µM, linearity up to 10 µM NAD + /NADH in 96-well plate assay. Convenient. The procedure involves adding a single working reagent, and reading the optical density at time zero and 15 min at room temperature. No 37°C heater is required. High-throughput. Can be readily automated as a high-throughput 96- well plate assay for thousands of samples per day.
Components:	Assay Buffer: 10 mL. Lactate: 1.5 mL. MTT Solution: 1.5 mL. Enzyme A: 120 μL. NAD Standard: 0.5 mL 1 mM. Enzyme B: 120 μL. NAD/NADH Extraction Buffers: each 12 mL.
Material not included:	Pipetting (multi-channel) devices. Clear-bottom 96-well plates (e.g. Corning Costar) and plate reader.

Target Details

Target:	NAD/NADH
Target Type:	Chemical
Background:	Sensitive determination of NAD and NADH by colorimetric (565nm) method.

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Pyridine nucleotides play an important role in metabolism and, thus, there is continual interest in monitoring their concentration levels. Quantitative determination of NAD + /NADH has applications in research pertaining to energy transformation and redox state of cells or tissue. Simple, direct and automation-ready procedures for measuring NAD + /NADH concentration are very desirable. This NAD + /NADH assay kit is based on a lactate dehydrogenase cycling reaction, in which the formed NADH reduces a formazan (MTT) reagent. The intensity of the reduced product color, measured at 565 nm, is proportionate to the NAD + /NADH concentration in the sample. This assay is highly specific for NAD + /NADH and with minimal interference (< 1%) by NADP + /NADPH. Our assay is a convenient method to measure NAD, NADH and their ratio.

Application Details

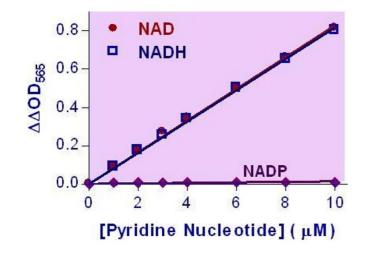
Application Notes:	Direct Assays: NAD + /NADH concentrations and ratios in cell or tissue extracts.
Comment:	1. At these concentrations, the standard curves for NAD and NADH are identical. Since NADH in solution is unstable, we provide only NAD as the standard.
	2. This assay is based on an enzyme-catalyzed kinetic reaction. Addition of Working Reagent
	should be quick and mixing should be brief but thorough. Use of multi-channel pipettor is recommended.
	3. The following substances interfere and should be avoided in sample preparation. EDTA (>0.5
	mM), ascorbic acid, SDS (>0.2%), sodium azide, NP-40 (>1%) and Tween-20 (>1%).
Protocol:	Add 80 μL Working Reagent per well quickly. Tap plate to mix briefly and thoroughly.
	Read optical density (OD0) for time zero at 565 nm (520-600nm) and OD15 after a 15-min
	incubation at room temperature.
	Calculation: Subtract OD0 from OD15 for the standard and sample wells. Use the OD values to
	determine sample NAD/NADH concentration from the standard curve. Note: If the sample OD
	values are higher than the OD value for the 10 μ Mstandard, dilute sample in distilled water and
	repeat this assay. Multiply the results by the dilution factor.
Reagent Preparation:	For each well of reaction, prepare Working Reagent by mixing 60 μ L Assay Buffer, 1 μ L Enzyme
	A, 1 μL Enzyme B, 14 μL Lactate and 14 μL MTT. Fresh reconstitution is recommended.
Sample Preparation:	For tissues weigh \sim 20 mg tissue for each sample, wash with cold PBS. For cell samples, wash
	cells with cold PBS and pellet \sim 10 5 cells for each sample. Homogenize samples (either tissue
	or cells) in a1.5 mL Eppindorf tube with either 100 μ L NAD extraction buffer for NAD

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	determination or 100 μ L NADH extraction buffer for NADH determination. Heat extracts at 60°C
	for 5 min and then add 20 μL Assay Buffer and 100 μL of the opposite extraction buffer to
	neutralize the extracts. Briefly vortex and spin the samples down at 14,000 rpm for 5 min. Use
	supernatant for NAD/NADH assays. Determination of both NAD and NADH concentrations
	requires extractions from two separate samples
Restrictions:	For Research Use only
Handling	
Storage:	4 °C
Publications	
Product cited in:	Bai, Cantó, Oudart, Brunyánszki, Cen, Thomas, Yamamoto, Huber, Kiss, Houtkooper,
	Schoonjans, Schreiber, Sauve, Menissier-de Murcia, Auwerx: "PARP-1 inhibition increases
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	Koo, Gong, Kim, Kim, Lee: "Improvement of coenzyme Q(10) production by increasing the
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	, pp. R84, (2009) (PubMed).

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Biochemical Assay

Image 1.

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