

### Datasheet for ABIN1112751

## **CXCL2 ELISA Kit**



#### Overview

Quantity:	96 tests
Target:	CXCL2
Reactivity:	Mouse
Method Type:	Sandwich ELISA
Detection Range:	15.6-1000 pg/mL
Minimum Detection Limit:	15.6 pg/mL
Application:	ELISA

### **Product Details**

Analytical Method:	Quantitative
Detection Method:	Colorimetric
Sensitivity:	< 5 pg/mL
Components:	1. One 96-well plate pre-coated with anti-Mouse MIP-2 antibody 2. Lyophilized Mouse MIP-2 standards: 2 tubes (10ng / tube) 3. Sample / Standard diluent buffer: 30ml 4. Biotin conjugated anti-Mouse MIP-2 antibody (Concentrated): 130 $\mu$ l.
Material not included:	1. 37 °C incubator 2. Microplate reader (wavelength: 450nm) 3. Precise pipette and disposable pipette tips 4. Automated plate washer 5. ELISA shaker 6. 1.5ml of Eppendorf tubes 7. Plate cover 8. Absorbent filter papers 9. Plastic or glass container with volume of above 1L

# Target Details

Target: CXCL2

# Target Details

MIP-2 (CXCL2 Products)
The major intrinsic protein of the ocular lens fiber membrane (MIP) is an abundant protein that appears during differentiation of the ocular lens and has a molecular weight of about 26,000 daltons. The MIP gene is 3.6 kb, contains 4 exons separated by introns ranging in size from 0.4 to 1.6 kb, and is present in single copy in the haploid human genome.  Cellular Response to Molecule of Bacterial Origin
Cellular response to Molecule of Basterial Origin
This kit was based on sandwich enzyme-linked immune-sorbent assay technology. Anti-MIP-2 polyclonal antibody was pre-coated onto 96-well plates. And the biotin conjugated anti-MIP-2 polyclonal antibody was used as detection antibodies. The standards test samples and biotin conjugated detection antibody were added - the wells subsequently and wash with wash buffer. Avidin-Biotin-Peroxidase Complex was added and unbound conjugates were washed away with wash buffer. TMB substrates were used - visualize HRP enzymatic reaction. TMB was catalyzed by HRP - produce a blue color product that changed into yellow after adding acidic stop solution. The density of yellow is proportional - the MIP-2 amount of sample captured in plate. Read the O.D. absorbance at 450 nm in a microplate reader and then the concentration of MIP-2 can be calculated.
Pre-coated
1. Before the experiment, centrifuge each kit component for several minutes to bring down all reagents to the bottom of tubes. 2. It is recommend to measure each standard and sample in duplicate. 3. Do NOT let the plate completely dry at any time! Since the dry condition can inactivate the biological material on the plate. 4. Do not reuse pipette tips and tubes to avoid cross contamination. 5. Do not use the expired components and the components from different batches. 6. To avoid the marginal effect of plate incubation for temperature differences (the marginal wells always get stronger reaction), it is recommend to equilibrate the ABC working solution and TMB substrate for at least 30 min at room temperature (37°C) before adding to wells. The TMB substrate (Kit Component 8) is colorless and transparent before use, if not, please contact us for replacement.
Preparation of sample and reagents 1. Sample Isolate the test samples soon after collecting, then, analyze immediately (within 2 hours). Or aliquot and store at -20 °C for long term. Avoid

### **Application Details**

analyze immediately or aliquot and store at -20  $^{\circ}\text{C}$  .

Serum: Coagulate the serum at room temperature (about 4 hours). Centrifuge at approximately  $2000 \times g$  for 20 min. Analyze the serum immediately or aliquot and store at -20 °C . Note: 1. Coagulate blood samples completely, then, centrifuge, and avoid hemolysis and particle. 2.

NaN3 can not be used as test sample preservative, since it is the inhibitor for HRP.

Restrictions:

For Research Use only

### Handling

Preservative:

Sodium azide, Thimerosal (Merthiolate)