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Datasheet for ABIN1112790 **TGFA ELISA Kit**

Overview

Quantity: 96 tests

Target: TGFA

Reactivity: Human

Method Type: Sandwich ELISA

Detection Range: 15.6-1000 pg/mL

Minimum Detection Limit: 15.6 pg/mL

Application: ELISA

Product Details

Analytical Method: Quantitative

Detection Method: Colorimetric

Sensitivity: < 1 pg/mL

Components: 1. One 96-well plate pre-coated with anti-Human TGFalpha antibody 2. Lyophilized Human TGFalpha standards: 2 tubes (10ng / tube) 3. Sample / Standard diluent buffer: 30ml 4. Biotin conjugated anti-Human TGFalpha antibody (Concentrated): 130 µl.

Material not included: 1. 37 °C incubator 2. Microplate reader (wavelength: 450nm) 3. Precise pipette and disposable pipette tips 4. Automated plate washer 5. ELISA shaker 6. 1.5ml of Eppendorf tubes 7. Plate cover 8. Absorbent filter papers 9. Plastic or glass container with volume of above 1L

Target Details

Target: TGFA

Target Details

Alternative Name: TGFalpha ([TGFA Products](#))

Background: Transforming growth factor alpha (TGF- α) is upregulated in some human cancers. It is produced in macrophages, brain cells, and keratinocytes, and induces epithelial development. It is closely related to EGF, and can also bind to the EGF receptor with similar effects. TGF α stimulates neural cell proliferation in the adult injured brain. TGFA is expressed rhythmically in the suprachiasmatic nucleus, and when infused into the third ventricle it reversibly inhibited locomotor activity and disrupted circadian sleep-wake cycles. TGFA also plays a role in certain paraneoplastic manifestations of melanoma.

Pathways: [NF-kappaB Signaling](#), [RTK Signaling](#), [EGFR Signaling Pathway](#)

Application Details

Comment: This kit was based on sandwich enzyme-linked immune-sorbent assay technology. Anti- TGF α polyclonal antibody was pre-coated onto 96-well plates. And the biotin conjugated anti- TGF α polyclonal antibody was used as detection antibodies. The standards test samples and biotin conjugated detection antibody were added - the wells subsequently and wash with wash buffer. Avidin-Biotin-Peroxidase Complex was added and unbound conjugates were washed away with wash buffer. TMB substrates were used - visualize HRP enzymatic reaction. TMB was catalyzed by HRP - produce a blue color product that changed into yellow after adding acidic stop solution. The density of yellow is proportional - the TGF α amount of sample captured in plate. Read the O.D. absorbance at 450 nm in a microplate reader and then the concentration of TGF α can be calculated.

Plate: Pre-coated

Reagent Preparation: 1. Before the experiment, centrifuge each kit component for several minutes to bring down all reagents to the bottom of tubes. 2. It is recommend to measure each standard and sample in duplicate. 3. Do NOT let the plate completely dry at any time! Since the dry condition can inactivate the biological material on the plate. 4. Do not reuse pipette tips and tubes to avoid cross contamination. 5. Do not use the expired components and the components from different batches. 6. To avoid the marginal effect of plate incubation for temperature differences (the marginal wells always get stronger reaction), it is recommend to equilibrate the ABC working solution and TMB substrate for at least 30 min at room temperature (37°C) before adding to wells. The TMB substrate (Kit Component 8) is colorless and transparent before use, if not, please contact us for replacement.

Sample Preparation: Preparation of sample and reagents 1. Sample Isolate the test samples soon after collecting,

Application Details

then, analyze immediately (within 2 hours). Or aliquot and store at -20 °C for long term. Avoid multiple freeze-thaw cycles.

Tissue lysate, body fluids or cell culture supernatants: Centrifuge to remove precipitate, analyze immediately or aliquot and store at -20 °C .

Serum: Coagulate the serum at room temperature (about 4 hours). Centrifuge at approximately 1000 × g for 10 min. Analyze the serum immediately or aliquot and store at -70 .° C

Plasma: Collect plasma with EDTA as the anticoagulant. Centrifuge for 15 min at 1000 x g within 30 min of collection. For eliminating platelet, suggesting that further centrifugation for 10 min at 2-8°C at 10000 x g. Analyze immediately or aliquot and store frozen at -20 °C. Heparin or citrate can not be used as anticoagulant here. Note: 1. Coagulate blood samples completely, then, centrifuge, and avoid hemolysis and particle. 2. NaN₃ can not be used as test sample preservative, since it is the inhibitor for HRP.

Restrictions: For Research Use only

Handling

Preservative: Sodium azide, Thimerosal (Merthiolate)