Datasheet for ABIN1563723

LDL ELISA Kit

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Overview

Quantity:	96 tests
Target:	LDL
Reactivity:	Monkey
Method Type:	Sandwich ELISA
Detection Range:	7.813-500 ng/mL
Minimum Detection Limit:	7.813 ng/mL
Application:	ELISA

Product Details

Sample Type:	Plasma, Serum
Analytical Method:	Quantitative
Detection Method:	Colorimetric
Cross-Reactivity (Details):	No significant cross-reactivity or interference between monkey Low Density Lipoprotein and analogues was observed. Note: Limited by current skills and knowledge, it is impossible for us to complete the cross-reactivity detection between Human bFGF and all the analogues, therefore, cross reaction may still exist.
Sensitivity:	4.688 ng/mL
Components:	Micro ELISA Plate, 8 x 12 well strips Reference Standard, 2 vials Reference Standard and Sample Diluent, 20 mL Concentrated Biotinylated Detection Ab, 120 µL

Product Details

	Biotinylated Detection Ab Diluent, 10 mL Concentrated HRP Conjugate, 120 µL HRP Conjugate Diluent, 10 mL Concentrated Wash Buffer (25x), 30 mL Substrate Reagent, 10 mL Stop Solution, 10 mL Plate Sealer, 5 pieces Desiccant, 1 pack Manual Certificate of Analysis
Material not included:	Microplate reader with 450nm wavelength filter High-precision transferpettor, EP tubes and disposable pipette tips 37°C Incubator Deionized or distilled water Absorbent paper Loading slot for Wash Buffer

Target Details

Target:	LDL
Alternative Name:	Low Density Lipoprotein (LDL Products)

Application Details

Application Notes:	**ELISA Plate:** The just opened ELISA Plate may appear water-like substance, which is normal and will not have any impact on the experiment results. **Add Sample:** The interval of sample adding between the first well and the last well should not be too long, otherwise will cause different pre-incubation time, which will significantly affect the experiment's accuracy and repeatability. For each step in the procedure, total dispensing time for addition of reagents or samples to the assay plate should not exceed 10 minutes. Parallel measure ment is recommended. **Incubation:** To prevent evaporation and ensure accurate results, proper adhesion of plate sealers during incubation steps is necessary. Do not allow wells to sit uncovered for extended periods between incubation steps. Do not let the strips dry at any time during the assay. Strict compliance with the given incubation time and temperature. **Washing:** The wash procedure is critical. Insufficient washing will result in poor precision and

falsely elevated absorbance readings. Residual liquid in the reaction wells should be pat dry against absorbent paper in the washing process. But don't put absorbent paper into reaction wells directly. Note that clear the residual liquid and fingerprint in the bottom before measurement, so as not to affect the micro-titer plate reader.

Reagent Preparation: As the volume of Detection Ab and HRP Conjugate is very small, liquid may adhere to the tube wall or tube cap when being transported. You better hand-throw it or centrifugal it for 1 minute at 1000rpm. Please pipette the solution for 4-5 times before pippeting. Please carefully reconstitute Standards, working solutions of Detection Ab and HRP Conjugate according to the instructions. To minimize imprecision caused by pipetting, ensure that pipettors are calibrated. It is recommended to suck more than 10μL for once pipetting. Do not reuse standard solution, working solution of Detection Ab and HRP Conjugate, which have been diluted. If you need to use standard repeatedly, you can divide the standard into small pack according to the amount of each assay, keep them at -20°C to -80°C and avoid repeated freezing and thawing.

Reaction Time Control: Please control reaction time strictly following this product description!

Substrate: Substrate Solution is easily contaminated. Please protect it from light.Stop Solution: As it is an acid solution, please pay attention to the protection of your eyes, hands, face and clothes when using this solution.

Mixing: You'd better use micro-oscillator at the lowest frequency, as sufficient and gentle mixing is particularly important to reaction result. If there is no micro-oscillator available, you can knock the ELISA plate frame gently with your finger before reaction.

Security: Please wear lab coats and latex gloves for protection. Especially detecting samples of blood or other body fluid, please perform following the national security columns of biological laboratories.

Do not use component from different batches of kit(washing buffer and stop solution can be an exception)

To avoid cross-contamination, change pipette tips between adding of each standard level, between sample adding, and between reagent adding. Also, use separate reservoirs for each reagent. Otherwise, the results will be inaccurate!

Comment:

Information on standard material:

The formulation of the standard is 0.01 M PBS. The standard contains additives (1 % BSA).

Information on reagents:

Reagents include 1 M SO_2. Azide, thimerosal, 2-mercaptoethanol (2-ME) or any other poisonous materials are not used.

Application Details

	Information on antibodies: The provided antibodies and their host vary in different kits. All antibodies are affinity purified
Sample Volume:	100 µL
Plate:	Pre-coated
Reagent Preparation:	Bring all reagents to room temperature (18-25°C) before use. **Wash Buffer:** Dilute 30 mL of Concentrated Wash Buffer into 750 mL of Wash Buffer with deionized or distilled water. Put unused solution back at 4°C. If crystals have formed in the concentrate, you can warm it with 40°C water bath (Heating temperature should not exceed 50°C) and mix it gently until the crystals have completely dissolved. The solution should be cooled to room temperature before use. **Standard:** Prepare standard within 15 minutes before use. Reconstitute the Standard with 1.0 mL of Sample Diluent, let it stand for 10 minutes until it dissolved fully. This reconstitution produces a stock solution. Then make serial dilutions as needed (Making serial dilution in the wells directly is not permitted). The Sample Diluent serves as the zero (0). **Biotinylated Detection Ab:** Calculate the required amount before experiment (100µL /well). In actual preparation, you should prepare 100-200µL more. Centrifuge the stock tube before use, dilute the concentrated Biotinylated Detection Ab to the working concentration using Diluent for Biotinylated Detection Ab (1:100). **Concentrated HRP Conjugate:** Calculate the required amount before experiment (100µL /well). In actual preparation you should prepare 100-200µL more. Dilute the Concentrated HRP Conjugate to the working concentration using Diluent for Concentrated HRP Conjugate (1:100). **Substrate Reagent:** As it is sensitive to light and contaminants, so you shouldn't open the vial until you need it! The needed dosage of the reagent can be aspirated with sterilized tips and the unused residual reagent shouldn't be dumped back into the vial again. **Note:** please don't prepare the reagent directly in the Diluent vials provided in the kit. Contaminated water or container for reagent preparation will influence the result.
Sample Collection:	Samples should be clear and transparent and be centrifuged to remove suspended solids. **Serum:** Allow samples to clot for 2 hours at room temperature or overnight at 4°C before centrifugation for 15 minutes at 1000×g. Collect the supernatant and carry out the assay immediately. Blood collection tubes should be disposable, non-pyrogenic, and non-endotoxin. **Plasma:** Collect plasma using EDTA or heparin as an anticoagulant. Centrifuge samples for 15 minutes at 1000×g at 2 - 8°C within 30 minutes of collection. Collect the supernatant and carry out the assay immediately. Hemolysis samples are not suitable for ELISA assay!

	Other biological fluids: Centrifuge samples for 20 minutes at 1000×g at 2 - 8°C. Collect the supernatant and carry out the assay immediately. **Tissue homogenates:** You'd better get detailed references from other literatures before assay aiming at different tissue types. For general information, hemolysis blood may affect the result, so you should mince the tissues to small pieces and rinse them in ice-cold PBS (0.02M, pH=7.4) to remove excess blood thoroughly. Tissue pieces should be weighed and then homogenized in PBS (the volume depends on the weight of the tissue) with a glass homogenizer on ice. To further break the cells, you can sonicate the suspension with an ultrasonic cell disrupter or subject it to freeze-thaw cycles. The homogenates are then centrifugated for 5minutes at 5000×g to get the supernate. Note: Samples should be used within 7 days when stored at 2-8°C, otherwise samples must be divided and stored at -20°C (≤ 1month) or -80°C (≤ 6months) to avoid the loss of bioactivity and contamination. Avoid repeated freeze-thaw cycles. Please take the samples to room temperature (18-25°C) without extra heating before performing the assay. Please predict the concentration before assaying. If the sample concentration is not within the range of the standard curve, users must determine the optimal sample dilutions for their particular experiments.
Assay Procedure:	Bring all reagents and samples to room temperature before use. Centrifuge the sample again after thawing before the assay. All the reagents should be mixed thoroughly by gently swirling before pipetting. Avoid foaming. It's recommended that all samples and standards be assayed in duplicate. **1. Add Sample:** Add 100µL of Standard, Blank, or Sample per well. The blank well is added with Reference Standard & Sample diluent. Solutions are added to the bottom of micro ELISA plate well, avoid inside wall touching and foaming as possible. Mix it gently. Cover the plate with sealer we provided. Incubate for 90 minutes at 37°C. **2. Biotinylated Detection Ab:** Remove the liquid of each well, don'tn'tn't wash. Immediately add 100µL of Biotinylated Detection Ab working solution to each well. Cover with the Plate sealer. Gently tap the plate to ensure thorough mixing. Incubate for 1 hour at 37°C. **3. Wash:** Aspirate each well and wash, repeating the process three times. Wash by filling each well with Wash Buffer (approximately 350µL) (a squirt bottle, multi-channel pipette, manifold dispenser or automated washer are needed). Complete removal of liquid at each step is essential. After the last wash, remove remained Wash Buffer by aspirating or decanting. Invert the plate and pat it against thick clean absorbent paper.

Application Details

	4. HRP Conjugate: Add 100µL of HRP Conjugate working solution to each well. Cover with the Plate sealer. Incubate for 30 minutes at 37°C. **5. Wash:** Repeat the wash process for five times as conducted in step 3. **6. Substrate:** Add 90µL of Substrate Solution to each well. Cover with a new Plate sealer. Incubate for about 15 minutes at 37°C. Protect the plate from light. The reaction time can be shortened or extended according to the actual color change, but not more than 30minutes. When apparent gradient appeared in standard wells, user should terminate the reaction. **7. Stop:** Add 50µLof Stop Solution to each well. Then, the color turns to yellow immediately. The order to add stop solution should be the same as the substrate solution. **8. OD Measurement:** Determine the optical density (OD value) of each well at once, using a micro-plate reader set to 450 nm. User should open the micro-plate reader in advance, preheat the instrument, and set the testing parameters. **9.** After experiment, put all the unused reagents back into the refrigerator according to the specified storage temperature respectively until their expiry
Calculation of Results:	Average the duplicate readings for each standard and samples, then subtract the average zero standard optical density. Create a standard curve by plotting the mean OD value for each standard on the y-axisagainst the concentration on the x-axis and draw a best fit curve through the points on the graph. It is recommended to use some professional software to do this calculation, such as curve expert 1.3 or 1.4. In the software interface, a best fitting equation of standard curve will be calculated using OD values and concentrations of standard sample. The software will calculate the concentration of samples after entering the OD value of samples. If samples have been diluted, the concentration calculated from the standard curve must be multiplied by the dilution factor. If the OD of the sample surpasses the upper limit of the standard curve, you should re-test it after appropriate dilution. The actual concentration is the calculated concentration multiplied dilution factor.
Restrictions:	For Research Use only

Handling

Handling Advice:	All the reagents in the kit should be stored according to the labels on vials. Unused wells should be returned to the foil pouch with the desiccant pack and resealed along entire edge of zip-seal. Substrate Reagent shouldn't be kept at -20 °C (Check!). Exposure of reagents to strong light should be avoided in the process of incubation and storage. All the taps of reagents should be tightened to prevent evaporation and microbial contamination. If not to store reagents according to above suggestions, erroneous results may occur.

Handling

Storage:	4 °C