antibodies -online.com





Datasheet for ABIN1981832

ERK1/2, JNK, p38 MAPK ELISA Kit





Publications



Go to Product page

()	11	\sim	rv		۱ ۸
	1 \ /	⊢	I \/	╙	1/1

Quantity:	2 x 96 tests	
Target:	ERK1/2, JNK, p38 MAPK	
Binding Specificity:	pThr180, pThr183, pThr185, pThr202, pTyr182, pTyr185, pTyr187, pTyr204	
Reactivity:	Human, Mouse, Rat	
Method Type:	Cell ELISA	
Application:	ELISA	
Product Details		
Purpose:	Cell-Based Human/Mouse/Rat ERK1/2 (Thr202/Tyr204), JNK (Thr183/Tyr185), p38 MAPK	
	(Thr180/Tyr182) Phosphorylation ELISA Kit. Suitable for adherent whole cell lines.	
Brand:	CellBIND®	
Sample Type:	Cell Culture Cells	
Analytical Method:	Semi-Quantitative	
Detection Method:	Colorimetric	
Specificity:	The antibodies provided in this kit recognizes human, mouse and rat Erk1 phosphorylated at	
	sites Thr202/Tyr204, Erk2 phosphorylated at sites Thr185/Tyr187, JNK phosphorylated at sites	
	Thr183/Tyr185 and p38 phosphorylated at sites Thr180/Tyr182. This kit also recognizes total	
	Erk1/2, total JNK and total p38 for comparison.	
Characteristics:	Site and signal pathway-specific	
	In vitro detection of adherent cell culture	
	No sample lysis needed	

Product Details

- · Compatible with a standard ELISA plate reader
- · Faster results than with ELISA
- · Adaptable for high-throughput screening and drug discovery

Components:

- uncoated 96-well Microplate
- · Wash Buffer A
- · Wash Buffer B
- · Fixing Solution
- · Quenching Buffer
- · Blocking Buffer
- · Anti-phospho antibodies
- · Anti-pan antibodies
- · HRP-Conjugated Secondary Antibody
- TMB One-Step Substrate
- · Stop Solution

Material not included:

- · Distilled or deionized water
- 100 mL and 1 liter graduated cylinders
- Tubes to prepare sample dilutions
- · Protease and Phosphatase inhibitors
- Precision pipettes to deliver 2 μL to 1 mL volumes
- Adjustable 1-25 mL pipettes for reagent preparation
- · Benchtop rocker or shaker
- · Microplate reader capable of measuring absorbance at 450 nm

Target Details

Target:	ERK1/2, JNK, p38 MAPK	
UniProt:	P27361, P28482, P45983, Q16539	

Application Details

Plate:	Uncoated
Protocol:	1. Seed 10,000-30,000 cells into each well and incubate overnight.
	2. Apply various treatment, inhibitors or activators according to manufacture's instructions.
	3. Add 100 µL of Fixing Solution into each well and incubate for 20 min at RT with shaking.
	4. Add 200 μL of prepared 1X Quenching Buffer and incubate 20 min at RT.
	5. Add 200 μL of Blocking Solution and incubate for 1 h at 37 °C.
	6. Add 50 μ L of 1X anti-phospho-protein specific antibody or anti-pan-protein specific antibody to each well and incubate for 2 h at RT.
	7. Add 50 μ L of prepared 1X HRP-Anti-Rabbit or Mouse IgG and incubate for 1 h at RT.

- 8. Add 100 µL of TMB One-Step Substrate Reagent to each well.
- 9. Incubate 30 min at RT.
- 10. Add 50 µL of Stop Solution to each well.
- 11. Read at 450 nm immediately.

Reagent Preparation:

NOTE: Thaw all reagents to room temperature immediately before use. If wash buffers contain visible crystals, warm to room temperature and mix gently until dissolved.

NOTE: Briefly centrifuge (~1,000g) ITEMS G, H, and I before opening to ensure maximum recovery.

For more information look at the picture.

Assay Procedure:

NOTE: ALL incubations and wash steps must be performed under gentle rocking or rotation (~1-2 cycles/sec).

1. Design your experiment. For example, see Figure 2 below.

OPTIONAL: If seeding HUVECs, HMEC-1 or other loosely attached cells, coat the Uncoated 96-Well Microplate (ITEM A) by adding 100 μ L poly-L-Lysine (Recommended Sigma Aldrich) into each well and then follow manufacturer's instructions. A pre-coated CellBIND® microplate or other poly-lysine treated tissue culture plate may be used in place of ITEM A.

2. Seed 100 μ L of 10,000 to 30,000 cells into each well of the Uncoated 96- Well Microplate (ITEM A) provided and incubate overnight at 37 °C with 5 % CO2.

NOTE: The optimal cell number used will vary on the cell line and the relative amount of protein phosphorylation. More or less cells may be used but this must be determined empirically.

NOTE: The cells can be starved ~4-24 hours (depending on cell line) prior to treatment with inhibitors or activators.

3. Apply various treatments, inhibitors (such as siRNA or chemicals) or activators according to manufacturer's instructions and incubate for the desired time points.

NOTE: It is recommended to dissolve inhibitors or activators into serum-free cell culture medium before treating the cells (unless otherwise stated in the manufacturer's instructions.)

- 4. Discard the cell culture medium by flipping the microplate upside down and gently tapping the bottom of the microplate over a sink.
- 5. Wash by pipetting 200 μ L of the prepared 1X Wash Buffer A (ITEM B) into each well. Discard the wash buffer (same as step 4) and wash 2 more times for a total of 3 washes using fresh wash buffer each time. After the final wash, gently blot the microplate onto a paper towel to remove any excess/remaining buffer.

NOTE: To avoid cell loss, do not pipette directly onto the cells. Instead, gently dispense the liquid down the wall of cell culture wells. Also avoid the use of vacuum suction or too forcefully tapping the microplate when discarding any solution.

 $6. \, \text{Add} \, 100 \, \mu \text{L}$ of Fixing Solution (ITEM D) into each well and incubate for 20 minutes at room temperature.

NOTE: The fixing solution is used to permeabilize the cells.

- 7. Repeat wash step 5.
- 8. Add 200 μ L of prepared 1X Quenching Buffer (ITEM E) into each well and incubate 20 minutes at room temperature.

NOTE: The quenching buffer is used to minimize the background response. 9. Wash 4 times with 1X Wash Buffer A.

- 10. Add 200 μ L of the prepared 1X Blocking Buffer (ITEM F) into each well and incubate for 1 hour at 37 °C.
- 11. Wash 3 times with the prepared 1X Wash Buffer B (ITEM C).

NOTE: If needed, the microplate may be stored at -80 °C for several days after this wash.

- 12. Add 50 μ L of the prepared 1X primary antibody (ITEM G-1, G-2, G-3, H- 1, H-2 or H-3) into each corresponding well and incubate for 2 hours at room temperature.
- 13. Wash 4 times with 1X Wash Buffer B.
- 14. Add 50 μ L of the prepared 1X HRP Conjugated secondary antibody (ITEM I) into each well and incubate for 1 hour at room temperature.
- 15. Repeat step 13.
- 16. Add 100 μ L of the TMB Substrate (ITEM J) into each well and incubate for 30 minutes at room temperature in the dark.
- 17. Add 50 µL of the Stop Solution (ITEM K) into each well. Read at 450 nm immediately.

Restrictions:

For Research Use only

Handling

Handling Advice:	Avoid repeated freeze-thaw cycles.	
Storage:	-20 °C	
Storage Comment:	The entire kit may be stored at -20°C for up to 6 months from the date of shipment. Avoid repeated freeze-thaw cycles.	
Expiry Date:	6 months	

Publications

Product cited in:

Matsui, Yamane, Kobayashi-Hattori, Oishi et al.: "Calcitonin gene-related peptide regulates mitogen-activated protein kinase pathway to decrease transforming growth factor β1-induced

hepatic plasminogen activator inhibitor-1 mRNA expression in HepG2 ..." in: **Bioscience, biotechnology, and biochemistry**, Vol. 78, Issue 5, pp. 787-90, (2015) (PubMed).

Chen, Hsu, Liou, Chen, Chen, Chiu, Yeh et al.: "Baicalein, an active component of Scutellaria baicalensis Georgi, prevents lysophosphatidylcholine-induced cardiac injury by reducing reactive oxygen species production, calcium overload and ..." in: **BMC complementary and alternative medicine**, Vol. 14, pp. 233, (2014) (PubMed).

Bas, Van De Water, Gupta, Dinh, Vu, Martínez-Soriano, Láinez, Marco: "Efficacy of three drugs for protecting against gentamicin-induced hair cell and hearing losses." in: **British journal of pharmacology**, Vol. 166, Issue 6, pp. 1888-904, (2012) (PubMed).

Allen, Wilson, Schneider, Lich, Roberts, Arthur, Woodford, Davis, Uronis, Herfarth, Jobin, Rogers, Ting: "NLRP12 suppresses colon inflammation and tumorigenesis through the negative regulation of noncanonical NF-?B signaling." in: **Immunity**, Vol. 36, Issue 5, pp. 742-54, (2012) (PubMed).

Patel, Jindal, King, Tilles, Yarmush: "The inflammatory response to double stranded DNA in endothelial cells is mediated by NF κ B and TNF α ." in: **PLoS ONE**, Vol. 6, Issue 5, pp. e19910, (2011) (PubMed).

There are more publications referencing this product on: Product page

Images

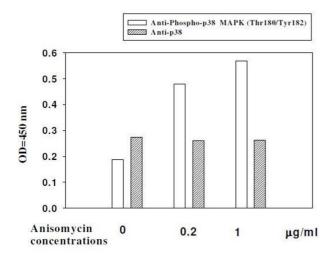


Image 1. Hela cells were stimulated by different concentrations of anisomycin for 1 hour at 37 °C

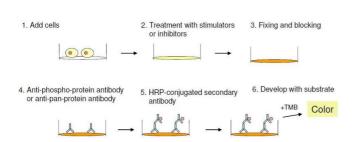


Image 2. Cell-Based protein phosphorylation procedure

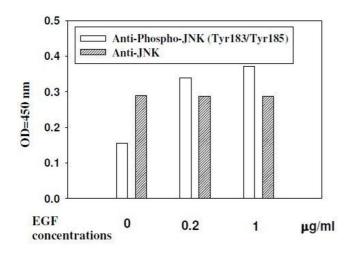


Image 3. Hela cells were stimulated by different concentrations of anisomycin for 1 hour at 37 °C

Please check the product details page for more images. Overall 9 images are available for ABIN1981832.