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Datasheet for ABIN1981833 JNK ELISA Kit

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Overview

Quantity:	96 tests
Target:	JNK (MAPK8)
Binding Specificity:	pThr183, pTyr185
Reactivity:	Human, Mouse, Rat
Method Type:	Cell ELISA
Application:	ELISA

Product Details

Purpose:	Cell-Based Human/Mouse/Rat JNK (Thr183/Tyr185) Phosphorylation ELISA Kit. Suitable for adherent whole cell lines.
Brand:	CellBIND®
Sample Type:	Cell Culture Cells
Analytical Method:	Semi-Quantitative
Detection Method:	Colorimetric
Specificity:	The antibodies provided in this kit recognizes human, mouse and rat JNK phosphorylated at sites Thr183 and Tyr185 as well as total JNK for comparison.
Characteristics:	 Site and signal pathway-specific In vitro detection of adherent cell culture No sample lysis needed Compatible with a standard ELISA plate reader Faster results than with ELISA

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Product Details

	Adaptable for high-throughput screening and drug discovery
Components:	uncoated 96-well Microplate
	Wash Buffer A
	• Wash Buffer B
	Fixing Solution
	Quenching Buffer
	Blocking Buffer
	Anti-phospho antibody
	Anti-pan antibody
	HRP-Conjugated Secondary Antibody
	TMB One-Step Substrate
	Stop Solution
Material not included:	Distilled or deionized water
	 100 mL and 1 liter graduated cylinders
	Tubes to prepare sample dilutions
	Protease and Phosphatase inhibitors
	 Precision pipettes to deliver 2 µL to 1 mL volumes
	Adjustable 1-25 mL pipettes for reagent preparation
	Benchtop rocker or shaker
	Microplate reader capable of measuring absorbance at 450 nm

Target Details

Target:	JNK (MAPK8)
Alternative Name:	JNK (MAPK8 Products)
Gene ID:	5599
UniProt:	P45983
Pathways:	MAPK Signaling, WNT Signaling, TLR Signaling, Fc-epsilon Receptor Signaling Pathway, Neurotrophin Signaling Pathway, Activation of Innate immune Response, Hepatitis C, Toll-Like
	Receptors Cascades, Signaling of Hepatocyte Growth Factor Receptor, S100 Proteins

Application Details

Sample Volume:	100 µL
Plate:	Uncoated
Protocol:	1. Seed 10,000-30,000 cells into each well and incubate overnight.

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	 Apply various treatment, inhibitors or activators according to manufacture's instructions. Add 100 µL of Fixing Solution into each well and incubate for 20 min at RT with shaking. Add 200 µL of prepared 1X Quenching Buffer and incubate 20 min at RT. Add 200 µL of Blocking Solution and incubate for 1 h at 37 °C. Add 50 µL of 1X anti-phospho-protein specific antibody or anti-pan-protein specific antibody to each well and incubate for 2 h at RT. Add 50 µL of prepared 1X HRP-Anti-Rabbit or Mouse IgG and incubate for 1 h at RT. Add 100 µL of TMB One-Step Substrate Reagent to each well. Incubate 30 min at RT. Add 50 µL of Stop Solution to each well.
Reagent Preparation:	NOTE: Thaw all reagents to room temperature immediately before use. If wash buffers contain visible crystals, warm to room temperature and mix gently until dissolved. NOTE: Briefly centrifuge (~1,000g) ITEMS G, H, and I before opening to ensure maximum
	recovery.
	For more information look at the picture.
Assay Procedure:	NOTE: ALL incubations and wash steps must be performed under gentle rocking or rotation (~1-2 cycles/sec).
	1. Design your experiment. For example, see Figure 2 below.
	OPTIONAL: If seeding HUVECs, HMEC-1 or other loosely attached cells, coat the Uncoated 96-
	Well Microplate (ITEM A) by adding 100 μ L poly-L-Lysine (Recommended Sigma Aldrich) into
	each well and then follow manufacturer's instructions. A pre-coated CellBIND ${ m I\!B}$ microplate or
	other poly-lysine treated tissue culture plate may be used in place of Item A.
	2. Seed 100 μL of 30,000 cells into each well of the Uncoated 96-Well Microplate (ITEM A)
	provided and incubate overnight at 37 $^\circ C$ with 5 $\%$ CO2.
	NOTE: The optimal cell number used will vary on the cell line and the relative amount of protein
	phosphorylation. More or less cells may be used but this must be determined empirically.
	NOTE: The cells can be starved ~4-24 hours (depending on cell line) prior to treatment with
	inhibitors or activators.
	3. Apply various treatments, inhibitors (such as siRNA or chemicals) or activators according to manufacturer's instructions and incubate for the desired time points.
	NOTE: It is recommended to dissolve inhibitors or activators into serum-free cell culture
	medium before treating the cells (unless otherwise stated in the manufacturer's instructions.)
	4. Discard the cell culture medium by flipping the microplate upside down and gently tapping
	the bottom of the microplate over a sink.
	5. Wash by pipetting 200 μL of the prepared 1X Wash Buffer A (ITEM B) into each well. Discard
	the wash buffer (same as step 4) and wash 2 more times for a total of 3 washes using fresh

	wash buffer each time. After the final wash, gently blot the microplate onto a paper towel to
	remove any excess/remaining buffer.
	NOTE: To avoid cell loss, do not pipette directly onto the cells. Instead, gently dispense the
	liquid down the wall of cell culture wells. Also avoid the use of vacuum suction or too forcefull
	tapping the microplate when discarding any solution.
	6. Add 100 μL of Fixing Solution (ITEM D) into each well and incubate for 20 minutes at room
	temperature.
	NOTE: The fixing solution is used to permeabilize the cells.
	7. Repeat wash step 5.
	8. Add 200 μL of the prepared 1X Quenching Buffer (ITEM E) into each well and incubate 20
	minutes at room temperature.
	NOTE: The quenching buffer is used to minimize the background response.
	9. Wash 4 times with 1X Wash Buffer A.
	10. Add 200 μ L of the prepared 1X Blocking Buffer (ITEM F) into each well and incubate for 1
	hour at 37 °C.
	11. Wash 3 times with the prepared 1X Wash Buffer B (ITEM C).
	NOTE: If needed, the microplate may be stored at -80 °C for several days after this wash.
	12. Add 50 μ L of the prepared 1X primary antibody (ITEM G or H) into each corresponding wel
	and incubate for 2 hours at room temperature.
	13. Wash 4 times with 1X Wash Buffer B.
	14. Add 50 μ L of the prepared 1X HRP Conjugated secondary antibody (ITEM I) into each well
	and incubate for 1 hour at room temperature.
	15. Wash 4 times with 1X Wash Buffer B.
	16. Add 100 μ L of the TMB Substrate (ITEM J) into each well and incubate for 30 minutes at
	room temperature in the dark.
	17. Add 50 μL of the Stop Solution (ITEM K) into each well. Read at 450 nm immediately.
Restrictions:	For Research Use only
Handling	
Handling Advice:	Avoid repeated freeze-thaw cycles.
Storage:	-20 °C
Storage Comment:	The entire kit may be stored at -20°C for up to 6 months from the date of shipment. Avoid

Expiry Date:

6 months

repeated freeze-thaw cycles.

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Images

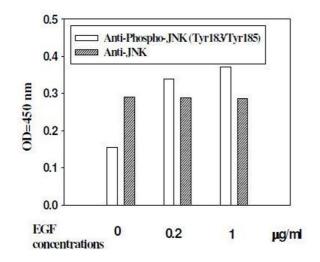
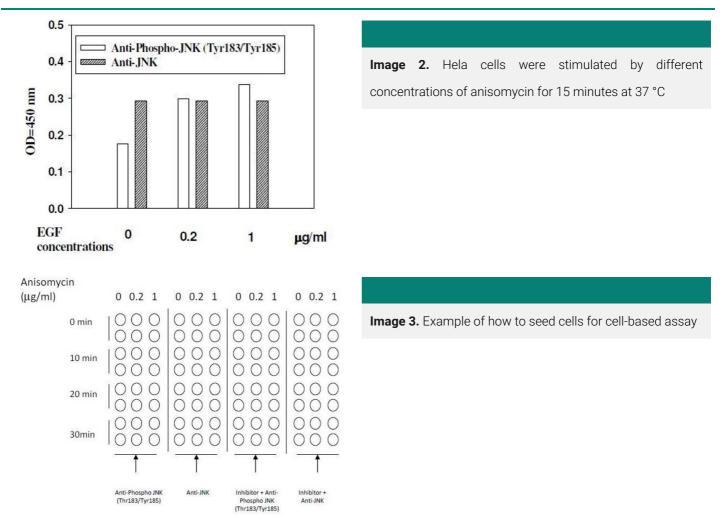


Image 1. Hela cells were stimulated by different concentrations of anisomycin for 1 hour at 37 °C

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Images



Please check the product details page for more images. Overall 6 images are available for ABIN1981833.