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Datasheet for ABIN1981837

## STAT1 ELISA Kit

6 Images

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### Overview

Quantity:	96 tests
Target:	STAT1
Binding Specificity:	pTyr701
Reactivity:	Human, Mouse, Rat
Method Type:	Cell ELISA
Application:	ELISA

### Product Details

Purpose:	Cell-Based Human/Mouse Stat 1 (Tyr701) Phosphorylation ELISA Kit. Suitable for adherent whole cell lines.
Brand:	CellBIND®
Sample Type:	Cell Culture Cells
Analytical Method:	Semi-Quantitative
Detection Method:	Colorimetric
Specificity:	The antibodies provided in this kit recognizes human and mouse Stat 1 phosphorylated at Tyrosine-701 and total Stat 1 for comparison.
Characteristics:	<ul style="list-style-type: none"><li>• Site and signal pathway-specific</li><li>• In vitro detection of adherent cell culture</li><li>• No sample lysis needed</li><li>• Compatible with a standard ELISA plate reader</li><li>• Faster results than with ELISA</li></ul>

## Product Details

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- Adaptable for high-throughput screening and drug discovery

Components:	<ul style="list-style-type: none"><li>• uncoated 96-well Microplate</li><li>• Wash Buffer A</li><li>• Wash Buffer B</li><li>• Fixing Solution</li><li>• Quenching Buffer</li><li>• Blocking Buffer</li><li>• Anti-phospho antibody</li><li>• Anti-pan antibody</li><li>• HRP-Conjugated Secondary Antibody</li><li>• TMB One-Step Substrate</li><li>• Stop Solution</li></ul>
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Material not included:	<ul style="list-style-type: none"><li>• Distilled or deionized water</li><li>• 100 mL and 1 liter graduated cylinders</li><li>• Tubes to prepare sample dilutions</li><li>• Protease and Phosphatase inhibitors</li><li>• Precision pipettes to deliver 2 <math>\mu</math>L to 1 mL volumes</li><li>• Adjustable 1-25 mL pipettes for reagent preparation</li><li>• Benchtop rocker or shaker</li><li>• Microplate reader capable of measuring absorbance at 450 nm</li></ul>
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## Target Details

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Target:	STAT1
Alternative Name:	STAT1 ( <a href="#">STAT1 Products</a> )
Background:	STAT1
Gene ID:	6772
UniProt:	<a href="#">P42224</a>
Pathways:	<a href="#">JAK-STAT Signaling</a> , <a href="#">RTK Signaling</a> , <a href="#">Interferon-gamma Pathway</a> , <a href="#">Response to Growth Hormone Stimulus</a> , <a href="#">Cellular Response to Molecule of Bacterial Origin</a> , <a href="#">Positive Regulation of Endopeptidase Activity</a> , <a href="#">Hepatitis C</a> , <a href="#">CXCR4-mediated Signaling Events</a>

## Application Details

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Sample Volume:	100 $\mu$ L
Plate:	Uncoated

## Application Details

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Protocol:

1. Seed 10,000-30,000 cells into each well and incubate overnight.
2. Apply various treatment, inhibitors or activators according to manufacture's instructions.
3. Add 100 µL of Fixing Solution into each well and incubate for 20 min at RT with shaking.
4. Add 200 µL of prepared 1X Quenching Buffer and incubate 20 min at RT.
5. Add 200 µL of Blocking Solution and incubate for 1 h at 37 °C.
6. Add 50 µL of 1X anti-phospho-protein specific antibody or anti-pan-protein specific antibody to each well and incubate for 2 h at RT.
7. Add 50 µL of prepared 1X HRP-Anti-Rabbit or Mouse IgG and incubate for 1 h at RT.
8. Add 100 µL of TMB One-Step Substrate Reagent to each well.
9. Incubate 30 min at RT.
10. Add 50 µL of Stop Solution to each well.
11. Read at 450 nm immediately.

Reagent Preparation:

NOTE: Thaw all reagents to room temperature immediately before use. If wash buffers contain visible crystals, warm to room temperature and mix gently until dissolved.

NOTE: Briefly centrifuge (~1,000g) ITEMS G, H, and I before opening to ensure maximum recovery.

For more information look at the picture.

Assay Procedure:

NOTE: ALL incubations and wash steps must be performed under gentle rocking or rotation (~1-2 cycles/sec).

1. Design your experiment. For example, see Figure 2 below.

OPTIONAL: If seeding HUVECs, HMEC-1 or other loosely attached cells, coat the Uncoated 96-Well Microplate (ITEM A) by adding 100 µL poly-L-Lysine (Recommended Sigma Aldrich) into each well and then follow manufacturer's instructions. A pre-coated CellBIND® microplate or other poly-lysine treated tissue culture plate may be used in place of Item A.

2. Seed 100 µL of 30,000 cells into each well of the Uncoated 96-Well Microplate (ITEM A) provided and incubate overnight at 37 °C with 5 % CO<sub>2</sub>.

NOTE: The optimal cell number used will vary on the cell line and the relative amount of protein phosphorylation. More or less cells may be used but this must be determined empirically.

NOTE: The cells can be starved ~4-24 hours (depending on cell line) prior to treatment with inhibitors or activators.

3. Apply various treatments, inhibitors (such as siRNA or chemicals) or activators according to manufacturer's instructions and incubate for the desired time points.

NOTE: It is recommended to dissolve inhibitors or activators into serum-free cell culture medium before treating the cells (unless otherwise stated in the manufacturer's instructions.)

4. Discard the cell culture medium by flipping the microplate upside down and gently tapping the bottom of the microplate over a sink.
5. Wash by pipetting 200 µL of the prepared 1X Wash Buffer A (ITEM B) into each well. Discard

the wash buffer (same as step 4) and wash 2 more times for a total of 3 washes using fresh wash buffer each time. After the final wash, gently blot the microplate onto a paper towel to remove any excess/remaining buffer.

NOTE: To avoid cell loss, do not pipette directly onto the cells. Instead, gently dispense the liquid down the wall of cell culture wells. Also avoid the use of vacuum suction or too forcefully tapping the microplate when discarding any solution.

6. Add 100  $\mu$ L of Fixing Solution (ITEM D) into each well and incubate for 20 minutes at room temperature.

NOTE: The fixing solution is used to permeabilize the cells.

7. Repeat wash step 5.

8. Add 200  $\mu$ L of the prepared 1X Quenching Buffer (ITEM E) into each well and incubate 20 minutes at room temperature.

NOTE: The quenching buffer is used to minimize the background response.

9. Wash 4 times with 1X Wash Buffer A.

10. Add 200  $\mu$ L of the prepared 1X Blocking Buffer (ITEM F) into each well and incubate for 1 hour at 37 °C.

11. Wash 3 times with the prepared 1X Wash Buffer B (ITEM C).

NOTE: If needed, the microplate may be stored at -80 °C for several days after this wash.

12. Add 50  $\mu$ L of the prepared 1X primary antibody (ITEM G or H) into each corresponding well and incubate for 2 hours at room temperature.

13. Wash 4 times with 1X Wash Buffer B.

14. Add 50  $\mu$ L of 1X HRP Conjugated secondary antibody (ITEM I) into each well and incubate for 1 hour at room temperature.

15. Wash 4 times with 1X Wash Buffer B.

16. Add 100  $\mu$ L of the TMB Substrate (ITEM J) into each well and incubate for 30 minutes at room temperature in the dark.

17. Add 50  $\mu$ L of the Stop Solution (ITEM K) into each well. Read at 450 nm immediately.

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Restrictions: For Research Use only

## Handling

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Handling Advice: Avoid repeated freeze-thaw cycles.

Storage: -20 °C

Storage Comment: The entire kit may be stored at -20°C for up to 6 months from the date of shipment. Avoid repeated freeze-thaw cycles.

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## Handling

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Expiry Date: 6 months

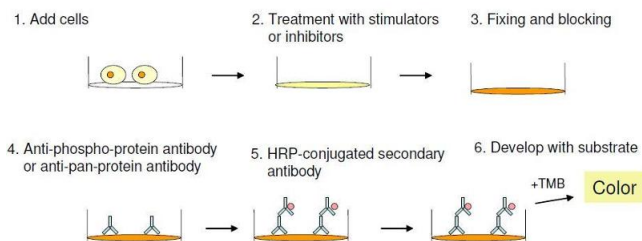
## Publications

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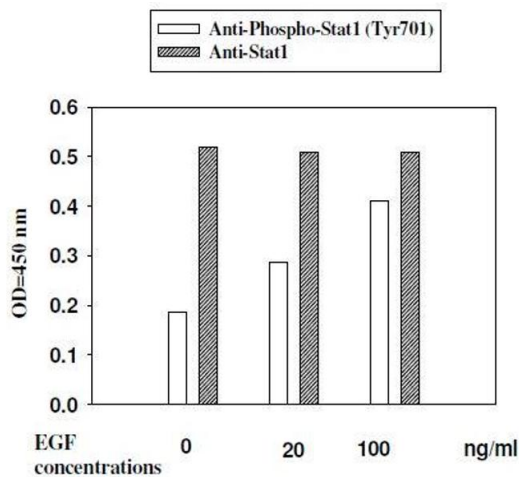
- Product cited in:
- Cesaro-Tadic, Dernick, Juncker, Buurman, Kropshofer, Michel, Fattinger, Delamarche: "High-sensitivity miniaturized immunoassays for tumor necrosis factor alpha using microfluidic systems." in: **Lab on a chip**, Vol. 4, Issue 6, pp. 563-9, (2004) ([PubMed](#)).
- Yan, Qing, Byers, Stadnyk, Al-Hertani, Bortolussi: "Role of MyD88 in diminished tumor necrosis factor alpha production by newborn mononuclear cells in response to lipopolysaccharide." in: **Infection and immunity**, Vol. 72, Issue 3, pp. 1223-9, (2004) ([PubMed](#)).
- Attarbaschi, Willheim, Ramharter, Hofmann, Wahl, Winkler, Graninger, Winkler: "T cell cytokine profile during primary Epstein-Barr virus infection (infectious mononucleosis)." in: **European cytokine network**, Vol. 14, Issue 1, pp. 34-9, (2003) ([PubMed](#)).
- Visser, Graffelman, Blauw, Haspels, Lentjes, de Kloet, Nagelkerken: "LPS-induced IL-10 production in whole blood cultures from chronic fatigue syndrome patients is increased but supersensitive to inhibition by dexamethasone." in: **Journal of neuroimmunology**, Vol. 119, Issue 2, pp. 343-9, (2001) ([PubMed](#)).
- Wahlström, Katchar, Wigzell, Olerup, Eklund, Grunewald: "Analysis of intracellular cytokines in CD4+ and CD8+ lung and blood T cells in sarcoidosis." in: **American journal of respiratory and critical care medicine**, Vol. 163, Issue 1, pp. 115-21, (2001) ([PubMed](#)).

ITEM	COMPONENT	PREPARATION	EXAMPLE
A	Uncoated 96-Well Microplate	No Preparation	N/A
B	20X Wash Buffer A Concentrate	Dilute 20-fold with distilled or deionized water	25 ml of concentrate + 475 ml of water = 500 ml of 1X working solution
C	20X Wash Buffer B Concentrate		
D	Fixing Solution	No Preparation	N/A
E	30X Quenching Buffer Concentrate	Dilute 30-fold with 1X Wash Buffer A	1 ml of concentrate + 29 ml of wash buffer = 30 ml of 1X working solution
F	5X Blocking Buffer Concentrate	Dilute 5-fold with distilled or deionized water	20 ml of concentrate + 80 ml of water = 100 ml of 1X working solution
PRIMARY ANTIBODY	G 1000X Mouse Anti-phospho (Tyr701) STAT1 Concentrate	Dilute 1000-fold with 1X Blocking Buffer	7 µl of concentrate + 6993 µl of 1X Blocking buffer = 7 ml of 1X working solution
	H 1000X Mouse Anti-STAT1 Concentrate		
SECONDARY ANTIBODY	I 1000X HRP Conjugated Anti-Mouse IgG Concentrate		10 µl of concentrate + 9990 µl of 1X Blocking buffer = 10 ml of 1X working solution
J	TMB Substrate	No Preparation	N/A
K	Stop Solution	No Preparation	N/A

**Image 1.** This picture shows the reagent preparation.



**Image 2.** Cell-Based protein phosphorylation procedure



**Image 3.** A431 cells were stimulated by different concentrations of EGF for 30 minutes at 37 °C.

Please check the [product details page](#) for more images. Overall 6 images are available for ABIN1981837.