Datasheet for ABIN1981848

Tyrosine ELISA Kit

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Overview

Quantity:	96 tests
Target:	Tyrosine
Binding Specificity:	phosphorylated
Reactivity:	Human, Mouse, Rat
Method Type:	Cell ELISA
Application:	ELISA

Product Details

Purpose:	Cell-Based Human/Mouse/Rat Tyrosine (Activated) Phosphorylation ELISA Kit. Suitable for adherent whole cell lines.
Brand:	CellBIND®
Sample Type:	Cell Culture Cells
Analytical Method:	Semi-Quantitative
Detection Method:	Colorimetric
Specificity:	The antibodies provided in this kit recognizes human, mouse and rat Phosphotyrosine.
Characteristics:	• Site and signal pathway-specific • In vitro detection of adherent cell culture • No sample lysis needed • Compatible with a standard ELISA plate reader • Faster results than with ELISA • Adaptable for high-throughput screening and drug discovery

Product Details

Components:	• uncoated 96-well Microplate • Wash Buffer A • Wash Buffer B • Fixing Solution • Quenching Buffer • Blocking Buffer • Anti-phospho antibody • HRP-Conjugated Secondary Antibody • TMB One-Step Substrate • Stop Solution
Material not included:	• Distilled or deionized water • 100 mL and 1 liter graduated cylinders • Tubes to prepare sample dilutions • Protease and Phosphatase inhibitors • Precision pipettes to deliver 2 µL to 1 mL volumes • Adjustable 1-25 mL pipettes for reagent preparation • Benchtop rocker or shaker • Microplate reader capable of measuring absorbance at 450 nm

Target Details

Target:	Tyrosine
Background:	Tyrosine

Application Details

Sample Volume:	100 µL
Plate:	Uncoated
Protocol:	1. Seed 10,000-30,000 cells into each well and incubate overnight. 2. Apply various treatment, inhibitors or activators according to manufacture's instructions. 3. Add 100 µL of Fixing Solution into each well and incubate for 20 min at RT with shaking. 4. Add 200 µL of prepared 1X Quenching Buffer and incubate 20 min at RT. 5. Add 200 µL of Blocking Solution and incubate for 1 h at 37 °C. 6. Add 50 µL of 1X anti-phospho-protein specific antibody or anti-pan-protein specific antibody to each well and incubate for 2 h at RT. 7. Add 50 µL of prepared 1X HRP-Anti-Rabbit or Mouse IgG and incubate for 1 h at RT. 8. Add 100 µL of TMB One-Step Substrate Reagent to each well. 9. Incubate 30 min at RT. 10. Add 50 µL of Stop Solution to each well. 11. Read at 450 nm immediately.

Application Details

Reagent Preparation:	NOTE: Thaw all reagents to room temperature immediately before use. If wash buffers contain visible crystals, warm to room temperature and mix gently until dissolved. NOTE: Briefly centrifuge (~1,000g) ITEMS G, H, and I before opening to ensure maximum recovery. For more information look at the picture.
Assay Procedure:	NOTE: ALL incubations and wash steps must be performed under gentle rocking or rotation (~1-2 cycles/sec). 1. Design your experiment. For example, see Figure 2 below. OPTIONAL: If seeding HUVECs, HMEC-1 or other loosely attached cells, coat the Uncoated 96-Well Microplate (ITEM A) by adding 100 µL poly-L-Lysine (Recommended Sigma Aldrich) into each well and then follow manufacturer's instructions. A pre-coated CellBIND® microplate or other poly-lysine treated tissue culture plate may be used in place of Item A. 2. Seed 100 µL of 10,000 to 30,000 cells into each well of the Uncoated 96- Well Microplate (ITEM A) provided and incubate overnight at 37 °C with 5 % CO2. NOTE: The optimal cell number used will vary on the cell line and the relative amount of protein phosphorylation. More or less cells may be used but this must be determined empirically. NOTE: The cells can be starved ~4-24 hours (depending on cell line) prior to treatment with inhibitors or activators. 3. Apply various treatments, inhibitors (such as siRNA or chemicals) or activators according to manufacturer's instructions and incubate for the desired time points. NOTE: It is recommended to dissolve inhibitors or activators into serum-free cell culture medium before treating the cells (unless otherwise stated in the manufacturer's instructions.) 4. Discard the cell culture medium by flipping the microplate upside down and gently tapping the bottom of the microplate over a sink. 5. Wash by pipetting 200 µL of the prepared 1X Wash Buffer A (ITEM B) into each well. Discard the wash buffer (same as step 4) and wash 2 more times for a total of 3 washes using fresh wash buffer each time. After the final wash, gently blot the microplate onto a paper towel to remove any excess/remaining buffer. NOTE: To avoid cell loss, do not pipette directly onto the cells. Instead, gently dispense the liquid down the wall of cell culture wells. Also avoid the use of vacuum suction or too forcefully tapping the microplate when discarding any solution. 6. Add 100 µL of Fixing Solution (ITEM D-1) into each well and incubate for 20 minutes at room temperature. NOTE: The fixing solution is used to permeabilize the cells. 7. Repeat wash step 5.

Application Details

	8. Add 200 µL of the prepared 1X Quenching Buffer (ITEM E) into each well and incubate 20 minutes at room temperature. NOTE: The quenching buffer is used to minimize the background response. 9. Wash 4 times with 1X Wash Buffer A. 10. Add 200 µL of the prepared 1X Blocking Buffer (ITEM F) into each well and incubate for 1 hour at 37 °C. 11. Wash 3 times with the prepared 1X Wash Buffer B (ITEM C). NOTE: If needed, the microplate may be stored at -80 °C for several days after this wash. 12. Add 100 µL of the prepared 1X primary antibody (ITEM G) into each corresponding well and incubate for 1 hour at room temperature. 13. Wash 4 times with 1X Wash Buffer B. 14. Add 100 µL of the TMB Substrate (ITEM J) into each well and incubate for 30 minutes at room temperature in the dark. 15. Add 50 µL of the Stop Solution (ITEM K) into each well. Read at 450 nm immediately.
Restrictions:	For Research Use only

Handling

Handling Advice:	Avoid repeated freeze-thaw cycles.
Storage:	-20 °C
Storage Comment:	The entire kit may be stored at -20°C for up to 6 months from the date of shipment. Avoid repeated freeze-thaw cycles.
Expiry Date:	6 months

Publications

Product cited in:	Banerjee, Lang, Hung, Sengupta, Banerjee, Baksi, Banerjee: "Unfolded protein response is required in nu/nu mice microvasculature for treating breast tumor with tunicamycin." in: **The Journal of biological chemistry**, Vol. 286, Issue 33, pp. 29127-38, (2011) (PubMed). Welters, Oknianska, Erdmann, Ryffel, Morgan: "The protein tyrosine phosphatase-BL, modulates pancreatic beta-cell proliferation by interaction with the Wnt signalling pathway." in: **The Journal of endocrinology**, Vol. 197, Issue 3, pp. 543-52, (2008) (PubMed). Kanangat, Postlethwaite, Hasty, Kang, Smeltzer, Appling, Schaberg et al.: "Induction of multiple

matrix metalloproteinases in human dermal and synovial fibroblasts by Staphylococcus aureus: implications in the pathogenesis of septic arthritis and other soft tissue ..." in: **Arthritis research & therapy**, Vol. 8, Issue 6, pp. R176, (2007) (PubMed).

Maile, Clemmons: "Regulation of insulin-like growth factor I receptor dephosphorylation by SHPS-1 and the tyrosine phosphatase SHP-2." in: **The Journal of biological chemistry**, Vol. 277, Issue 11, pp. 8955-60, (2002) (PubMed).

Gingras, Champagne, Roy, Lavoie: "Cytoplasmic death signal triggered by SRC-mediated phosphorylation of the adenovirus E4orf4 protein." in: **Molecular and cellular biology**, Vol. 22, Issue 1, pp. 41-56, (2001) (PubMed).

Images

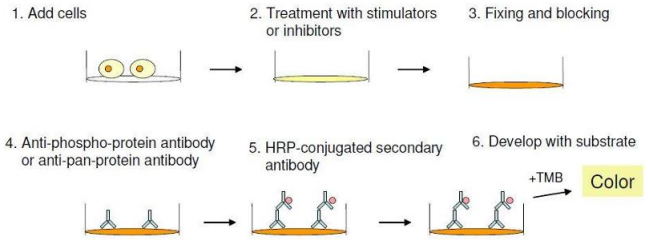

Image 1. Cell-Based protein phosphorylation procedure

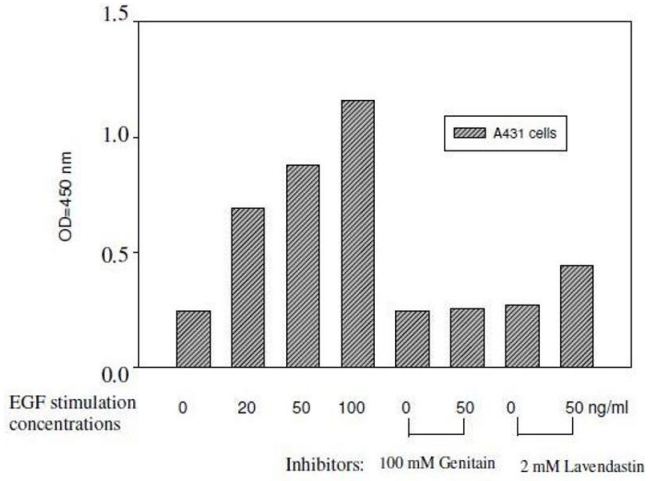

Image 2. A431 cells were treated for 30 min with 50 µL of 100 mM Genistein or 2 mM Lavendustin in appropriate wells at room temperature prior to EGF stimulation. Added 50 µL different concentrations of rhEGF (0, 20, 50 or 100 ng/mL in serum free DMEM) to appropriate wells. Then incubated for 10 min at 37 °C .

Images

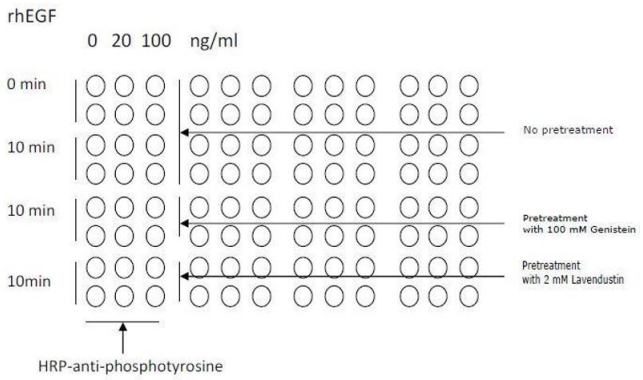

Image 3. Example of how to seed cells for cell-based assay

Please check the product details page for more images. Overall 4 images are available for ABIN1981848.