

Datasheet for ABIN2344850

CytoSelect™ 96-well Cell Migration Assay (3 µm), Fluorometric[Go to Product page](#)

4 Publications

Overview

Quantity:	96 tests
Reactivity:	Mammalian
Application:	Cellular Assay (CA)

Product Details

Brand:	CytoSelect™
Sample Type:	Serum, Cell Samples
Analytical Method:	Quantitative
Detection Method:	Fluorometric
Characteristics:	<p>CytoSelect™ 96-well Cell Migration Assay Kit utilizes a polycarbonate membrane plate (3 µm pore size) to assay the migratory properties of cells. The kit does not require you to prelabel the cells with Calcein AM or remove non-migratory cells (i.e. cotton swabbing). Any migratory cells are first dissociated from the membrane, then lysed and detected with CyQuant® GR Dye.</p> <p>CytoSelect™ 96-well Cell Migration Assay Kit provides a robust system for the quantitative determination of cell migration. The kit contains sufficient reagents for the evaluation of 96 samples. The 3 µm pore size is optimal for leukocyte cell migration. However, in the case of epithelial or fibroblast chemotaxis, a larger pore size (8 µm) is recommended. The CytoSelect™ Cell Migration Assay Kit contains a polycarbonate membrane chamber (3 µm pore size) in a 96-well plate. The membrane serves as a barrier to discriminate migratory cells from non-migratory cells. Migratory cells are able to extend protrusions towards chemoattractants (via actin cytoskeleton reorganization) and ultimately pass through the pores of the polycarbonate membrane. These migratory cells are then dissociated from the membrane and subsequently detected with CyQuant® GR Dye.</p>

Product Details

Components:	<ol style="list-style-type: none">1. 96-well Cell Migration Plate : One sterile 96-well plate (see Figure 1 for components)2. 96-well Cell Harvesting Tray : One 96-well tray3. Cell Detachment Solution : One 20 mL bottle4. 4X Lysis Buffer : One 10 mL bottle5. CyQuant® GR Dye : One 75 µL tube
Material not included:	<ol style="list-style-type: none">1. Migratory cell lines2. Cell culture medium3. Serum free medium, such as DMEM containing 0.5 % BSA, 2 mM CaCl₂ and 2 mM MgCl₂4. FBS or desired chemoattractant5. Cell culture incubator (37 °C, 5 % CO₂ atmosphere)6. Light microscope7. 96-well plate suitable for a fluorescence plate reader8. Fluorescence plate reader 4 Top Plate Cover Middle Migration Plate Membrane Chamber Bottom Feeder Tray : Components of the 96-well Cell Migration Plate.

Target Details

Background:	<p>Cell migration is a highly integrated, multistep process that orchestrates embryonic morphogenesis, tissue repair and regeneration. It plays a pivotal role in the disease progression of cancer, mental retardation, atherosclerosis, and arthritis. The initial response of a cell to a migration-promoting agent is to polarize and extend protrusions in the direction of the attractant, these protrusions can consist of large, broad lamellipodia or spike-like filopodia. In either case, these protrusions are driven by actin polymerization and can be stabilized by extracellular matrix (ECM) adhesion or cell-cell interactions.</p>
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Application Details

Application Notes:	Optimal working dilution should be determined by the investigator.
Comment:	<ul style="list-style-type: none">• Fully quantify chemotaxis with no manual cell counting• Measure chemotaxis in less than 6 hours with most cell types• Membrane inserts are uncoated to allow use with any chemoattractant
Assay Procedure:	<ol style="list-style-type: none">1. Allow the 96-well Migration Plate to warm up at room temperature for 10 minutes.2. Prepare a cell suspension containing 0.5-5.0 x 10⁶ cells/mL in serum free media. Agents that inhibit or stimulate cell migration can be added directly to the cell suspension. (Note: Overnight starvation may be performed prior to running the assay)3. Under sterile conditions, separate the cover and membrane chamber from the 96-well Migration Plate.4. Add 150 µL of media containing 10 % fetal bovine serum or desired chemoattractant(s) to

Application Details

- the wells of the feeder tray.
- Place the membrane chamber back into the feeder tray (containing chemoattractant solution). Ensure no bubbles are trapped under the membrane. 5
 - Gently mix the cell suspension (without chemoattractant) from step 2 and add 100 μL to the membrane chamber.
 - Finally, cover the plate and transfer to a cell culture incubator for 2-24 hours.
 - Just prior to the end of the incubation, pipette 150 μL of prewarmed Cell Detachment Solution into wells of the clean, 96-Well Cell Harvesting Tray (provided).
 - Carefully remove the 96-well Migration Plate from the incubator. Separate the membrane chamber from the feeder tray. Note: Retain the feeder tray for step
 10. Remove the cells/media from the top side of the membrane chamber by aspirating or inverting. Place the membrane chamber into the Cell Harvesting Tray containing 150 μL of Cell Detachment Solution (step 8). Incubate 30 minutes at 37 $^{\circ}\text{C}$.
 - Completely dislodge the cells from the underside of the membrane by gently tilting the membrane chamber several times in the Cell Detachment Solution.
 - In a clean 96-well plate (not provided), combine 75 μL of media from the feeder tray (step 9) with 75 μL of the detachment solution (step 11).
 - Prepare sufficient 4X Lysis Buffer/CyQuant[®] GR dye solution for all samples by diluting the dye 1:75 in 4X Lysis Buffer (for example, add 5 μL dye to 370 μL of 4X Lysis Buffer).
 - Add 50 μL of 4X Lysis Buffer/CyQuant[®] GR dye solution to each well (already containing 150 μL of Cell Detachment Solution). Incubate 20 minutes at room temperature.
 - Transfer 150 μL of the mixture to a 96-well plate suitable for fluorescence measurement. Read the fluorescence with a fluorescence plate reader at 480 nm/520 nm.

Restrictions: For Research Use only

Handling

Storage: 4 $^{\circ}\text{C}$

Storage Comment: Store all components at 4 $^{\circ}\text{C}$.

Publications

Product cited in: Pengjam, Madhyastha, Madhyastha, Yamaguchi, Nakajima, Maruyama: "Anthraquinone Glycoside Aloin Induces Osteogenic Initiation of MC3T3-E1 Cells: Involvement of MAPK Mediated Wnt and Bmp Signaling." in: **Biomolecules & therapeutics**, Vol. 24, Issue 2, pp. 123-31, (2016) ([PubMed](#)).

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