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Datasheet for ABIN2344858 CytoSelect[™] 24-well Cell Migration Assay (12 μm), Fluorometric

Publication



Overview

| Quantity: | 12 tests |
|--------------|---------------------|
| Reactivity: | Mammalian |
| Application: | Cellular Assay (CA) |

Product Details

| Serum, Cell Samples |
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| |
| Quantitative |
| Fluorometric |
| CytoSelect™ Cell Migration Assay Kit utilizes polycarbonate membrane inserts (8 µm pore size) |
| to assay the migratory properties of cells. The kit does not require you to prelabel the cells with |
| Calcein AM or remove non-migratory cells (i.e. cotton swabbing). Any migratory cells are first |
| dissociated from the membrane, then lysed and detected by the patented CyQuant ${ m I\!B}$ GR Dye |
| (Invitrogen). CytoSelect™ Cell Migration Assay Kit provides a robust system for the quantitative |
| determination of cell migration. The kit contains sufficient reagents for the evaluation of 12 |
| samples. The 12 μm pore size is optimal for studying slow moving cells or cells with large size |
| such as primary astrocytes. The CytoSelect™ Cell Migration Assay Kit contains polycarbonate |
| membrane inserts (12 μ m pore size) in a 24-well plate. The membrane serves as a barrier to |
| discriminate migratory cells from non- migratory cells. Migratory cells are able to extend |
| protrusions towards chemoattractants (via actin cytoskeleton reorganization) and ultimately |
| pass through the pores of the polycarbonate membrane. These migratory cells are then |
| dissociated from the membrane and subsequently detected by the patented CyQuant® GR Dye |
| (Invitrogen). |
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Product Details

| Components: | 1. 24-well Migration Plate : One 24-well plate containing 12 cell culture inserts (12 μm pore size) 2. Cell Detachment Solution : One 5 mL bottle |
|------------------------|--|
| | 3. 4X Lysis Buffer : One 5 mL bottle |
| | 4. CyQuant® GR Dye : One 25 μL tube |
| | 5. Forceps: One each |
| Material not included: | 1. Migratory cell lines |
| | 2. Cell culture medium |
| | 3. Serum free medium, such as DMEM containing 0.5 % BSA, 2 mM CaCl2 and 2 mM MgCl2 |
| | 4. Cell culture incubator (37 °C, 5 % CO2 atmosphere) |
| | 5. Light microscope |
| | 6. 96-well plate suitable for a fluorescence plate reader |
| | 7. Fluorescence plate reader |

Target Details

| Background: | Cell migration is a highly integrated, multistep process that orchestrates embryonic |
|-------------|--|
| | morphogenesis, tissue repair and regeneration. It plays a pivotal role in the disease progression |
| | of cancer, mental retardation, atherosclerosis, and arthritis. The initial response of a cell to a |
| | migration-promoting agent is to polarize and extend protrusions in the direction of the |
| | attractant, these protrusions can consist of large, broad lamellipodia or spike-like filopodia. In |
| | either case, these protrusions are driven by actin polymerization and can be stabilized by |
| | extracellular matrix (ECM) adhesion or cell-cell interactions (via transmembrane receptors). |

Application Details

| Application Notes: | Optimal working dilution should be determined by the investigator. |
|--------------------|--|
| Comment: | Fully quantify chemotaxis with no manual cell counting |
| | Measure chemotaxis in less than 6 hours with most cell types |
| | Membrane inserts are uncoated to allow use with any chemoattractant |
| Assay Procedure: | 1. Under sterile conditions, allow the 24-well migration plate to warm up at room temperature for 10 minutes. |
| | 2. Prepare a cell suspension containing 0.5-1.0 x 106 cells/mL in serum free media. Agents that inhibit or stimulate cell migration can be added directly to the cell suspension. (Note: Overnight starvation may be performed prior to running the assay) |
| | 3. Add 500 µL of media containing 10 % fetal bovine serum or desired chemoattractant(s) to the lower well of the migration plate. |
| | 4. Add 300 μ L of the cell suspension solution to the inside of each insert. |
| | 5. Incubate for 2-24 hours in a cell culture incubator. |

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| | 6. Carefully aspirate the media from the inside of the insert. Transfer the insert to a clean well containing 225 μL of Cell Detachment Solution. Incubate 30 minutes at 37 °C. 7. Completely dislodge the cells from the underside of the membrane by gently tilting the insert several times in the detachment solution. Remove and discard the insert. 8. Prepare sufficient 4X Lysis Buffer/CyQuant® GR dye solution for all samples by diluting the dye 1:75 in 4X Lysis Buffer (for example, add 5 μL dye to 370 μL of 4X Lysis Buffer). 4 9. Add 75 μL of 4X Lysis Buffer/CyQuant® GR dye solution to each well containing cells and 225 μL of Cell Detachment Solution. Incubate 20 minutes at room temperature. 10. Transfer 200 μL of the mixture to a 96-well plate suitable for fluorescence measurement. Read fluorescence with a fluorescence plate reader at 480 nm/520 nm. |
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| Restrictions: | For Research Use only |
| Handling | |
| Storage: | 4 °C |
| Storage Comment: | Store all components at 4°C. |
| Publications | |
| Product cited in: | Dubuisson, Day, Dhurandhar: "Accurate identification of neutralizing antibodies to adenovirus |
| | Ad36, -a putative contributor of obesity in humans." in: Journal of diabetes and its |
| | complications, Vol. 29, Issue 1, pp. 83-7, (2014) (PubMed). |