Datasheet for ABIN285285

anti-C3a antibody

2 Images | 1 Publication

Go to Product page

Overview

Quantity:	500 µL
Target:	C3a
Reactivity:	Human
Host:	Rabbit
Clonality:	Polyclonal
Conjugate:	This C3a antibody is un-conjugated
Application:	Western Blotting (WB), ELISA, Immunodiffusion (ID)

Product Details

Immunogen:	Complement C3a antibody was raised in rabbit using highly purified human complement protein as the immunogen.

Target Details

Target:	C3a
Alternative Name:	Complement C3a (C3a Products)
Pathways:	Complement System

Application Details

Application Notes:	ELISA: 1:500-1:2,000, Immunodiffusion: 1:4, WB: 1:500-1:1,000
Restrictions:	For Research Use only

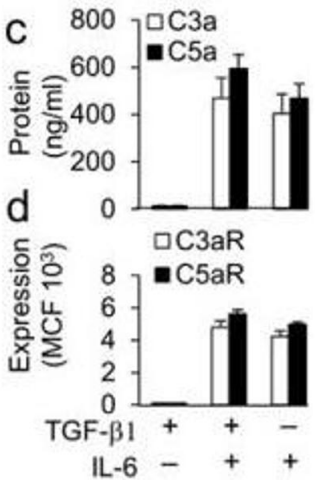
Handling

Format:	Liquid
Concentration:	Lot specific
Buffer:	as a liquid PBS, pH 7.4, with 0.05 % NaN3.
Preservative:	Sodium azide
Precaution of Use:	This product contains sodium azide as preservative. Although the amount of sodium azide is very small appropriate care must be taken when handling this product.
Handling Advice:	Avoid repeated freeze/thaw cycles
Storage:	-20 °C
Storage Comment:	Store at 4 °C for short term storage. Aliquot and store at -20 °C for long term storage.

Publications

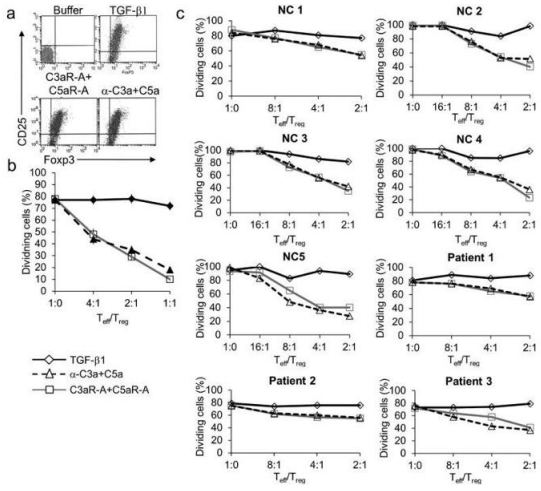
Product cited in:	Strainic, Shevach, An, Lin, Medof: "Absence of signaling into CD4 cells via C3aR and C5aR enables autoinductive TGF-β1 signaling and induction of Foxp3 regulatory T cells." in: **Nature immunology**, Vol. 14, Issue 2, pp. 162-71, (2013) (PubMed).

Images



Flow Cytometry

Image 1. C3a antibody (ABIN285285): Sorted WT Foxp3− CD4+ T cells were incubated for 48 hr with anti-CD3+CD28 beads plus TGF-β1 alone, TGF-β1+IL-6, or IL-6 alone as in (b). Culture supernatants were assayed for (c) C3a and C5a generation by ELISAs, and (d) C3aR and C5aR surface expression by flow cytometry (P<0.01 for TGF-β1 alone vs TGF-β1+IL-6 or IL-6 alone, Mean Fluorescence Intensity (MFI) values; n=5). Source: PMC4144047



Flow Cytometry

Image 2. (a) Flow sorted human CD45RA+CD25−CD4+ T cells (1×106) were incubated for 3 days with soluble anti-CD3 mAb (3 µg/ml), rhIL-2 (5 ng/ml), and 2.5×105 autologous CD11c+ DCs in the absence or presence of 1) TGF-β1 (5 ng/ml), 2) each of C3aR-A/C5aR-A (10 nM), or 3) each of anti-C3a(ABIN285285)/C5a mAbs (10 µg/ml). Percent Foxp3+CD25+ CD4+ T cells then were determined by flow cytometry. (b) Flow sorted CD25+ cells from (a) were incubated for 3 days with in differing Teff/iTreg ratios with 1×106 CFSE labeled autologous naive CD25− cells, anti-CD3 mAb (3 µg/ml), and 1×104 autologous CD11c+ DCs. Percent dividers was determined by CFSE dilution. (c) CD45RA+CD25−CD4+ T cells (1×106) were isolated from 5 healthy controls (NC) and 3 MS patients by flow sorting. The cells were incubated for 3 days with soluble anti-CD3 mAb (3 µg/ml), rhIL-2 (5 ng/ml), and 2.5×105 autologous DCs in the absence or presence of 1) rhTGF-β1 (5 ng/ml), 2) C3aR-A/C5aR-A (10 nM), or 3) anti-C3a (ABIN285285)/C5a mAbs (5 µg/ml). Cells were washed and sorted on CD25. After sorting, CD25+ (Treg) were incubated for 3 days with anti-CD3 mAb, 2.5×105 autologous DCs, and 106 CD25− (Effector) cells prelabeled with CFSE. Percent dividers was determined by CFSE dilution. Source: PMC4144047