

Datasheet for ABIN2862593

Oxytocin ELISA Kit



3

5 x 96 tests

Publications



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Quantity:

Target: Oxytocin (OXT) Reactivity: Various Species Method Type: Sandwich ELISA Application: ELISA Product Details		
Method Type: Sandwich ELISA Application: ELISA		
Application: ELISA		
Product Details		
Purpose: The DetectX® Oxytocin Immunoassay kit is designed to quantitatively measure	e Oxytocin	
present in serum, plasma, clarified milk and tissue culture media samples.		
Brand: DetectX®		
Sample Type: Serum, Plasma, Saliva, Milk, Tissue Culture Medium	Serum, Plasma, Saliva, Milk, Tissue Culture Medium	
Analytical Method: Quantitative	Quantitative	
Detection Method: Colorimetric		
Cross-Reactivity (Details): The following cross reactants were tested in the assay and calculated at the 50) % binding point.	
Steroid Cross Reactivity: Oxytocin 100 %, Isotocin 94.3 %, Mesotocin 88.4 %, Ly	's8-Vasopressin	
0.14 %, Arg8-Vasotocin 0.13 %, Arg8-Vasopressin 0.12 %		
Components: Coated Clear 96 Well Plates Clear plastic microtiter plate(s) with break-apart st	rips coated with	
goat anti-rabbit IgG. 1 Or 5 Each		
Oxytocin Standard Oxytocin at 100,000 pg/mL in a special stabilizing solution.	125 Or 625 μL	
Calibrated to the 4th WHO International Standard NIBSC code: 76/575		
DetectX® Oxytocin Antibody A rabbit polyclonal antibody specific for oxytocin.	3 mL Or 13 mL	

DetectX® Oxytocin Conjugate Oxytocin-peroxidase conjugate in a special stabilizing solution. 3 mL Or 13 mL

Assay Buffer Concentrate Assay Buffer, 5X concentrate that should be diluted with deionized or distilled water. 28 mL or 55 mL

Extraction Solution A special extraction solution for treatment of serum and plasma samples to extract oxytocin. 50 mL or 250 mL

Wash Buffer Concentrate A 20X concentrate that should be diluted with deionized or distilled water. 30 mL Or 125 mL

TMB Substrate 11 mL Or 55 mL

Stop Solution A 1M solution of hydrochloric acid. CAUSTIC. 5 mL Or 25 mL

Plate Sealer 1 Or 5 Each

Material not included:

Distilled or deionized water.

A Speedvac/centrifugal concentrator or N2 gas and gas manifold for evaporation.

Repeater pipet, such as an Eppendorf repeater, with disposable tips to accurately dispense 25,

50 and $100~\mu L.$

A microplate shaker.

Colorimetric 96 well microplate reader capable of reading optical density at 450 nm.

Software for converting raw relative optical density readings from the plate reader and carrying out four parameter logistic curve (4PLC) fitting.

Target Details

Target:	Oxytocin (OXT)
Alternative Name:	Oxytocin (OXT Products)
Target Type:	Hormone
Background:	The neuropeptides, oxytocin and vasopressin were isolated and synthesized by Vincent du
	Vigneaud at Cor- nel Medical College in 1953, work for which he received the Nobel Prize in
	Chemistry in 1955. Oxytocin is a neurohypophysial peptide which is produced in the
	paraventricular nuclei of the hypothalamus and stored in the posterior pituitary. The molecule
	consists of nine amino acids linked with a [1-6] disulfide bond and a semi- flexible
	carboxyamidated tail. A hormone once thought to be limited to female smooth muscle
	reproductive physiology and neurotransmitter1,2, recent studies have begun to investigate
	oxytocin's role in various behav- iors, including orgasm, social recognition, pair bonding, anxiety,
	and maternal behaviors3,4 and is important in male reproductive physiology5. Oxytocin and the
	related neurohypophysial peptide, Arg8-Vasopressin, main- tain renal water and sodium

balance6. Oxytocin Highly conserved across species boundaries, oxytocin-like neurohypophysial peptides are substituted primar- ily at residues 4 and/or 8. In the oxytocin-like peptide, mesotocin, a common peptide found in some fishes, reptiles, birds, amphibians, marsupials and non-mammalian tetrapods, the leucine at residue 8 is substituted for isoleucine7. Acting in classical endocrine fashion, Oxytocin elicits regulatory effects by binding specific cell surface receptors which in turn initiate a secondary intracelluar response cascade via a phosphoinositide signaling pathway

Pathways:

Myometrial Relaxation and Contraction, Feeding Behaviour

Application Details

Application Notes: This assay has been validated for serum, EDTA and heparin plasma, clarified milk, and tissue

cul-ture samples.

Samples containing visible particulate matter should be centrifuged before use.

Oxytocin is identical across all species and we expect this kit may measure oxytocin from sources other than human.

Because of the cross reactivity to mesotocin this kit should also be able to measure mesotocin from birds, fish and amphibians.

The end user should evaluate recoveries of oxytocin in other samples being tested.

Plate: Pre-coated

Protocol: An oxytocin standard is provided to generate a standard curve for the assay and all samples

should be read off the standard curve.

Standards or diluted samples are pipetted into a clear microtiter plate coated with an antibody

to capture rabbit antibodies.

An oxytocin-peroxidase conjugate is added to the standards and samples in the wells.

The binding reaction is initiated by the addition of a polyclonal antibody to oxytocin to each well.

After an overnight incubation at 4 °C the plate is washed and supplied substrate is added.

The substrate reacts with the bound oxytocin-per-oxidase conjugate.

After a 30 minute incubation, the reaction is stopped and the intensity of the generated color is

detected in a microtiter plate reader capable of measuring 450nm wavelength.

The concentration of the oxytocin in the sample is calculated, after making suitable correction

for the dilution of the sample, using software available with most plate readers.

Reagent Preparation: Allow the kit reagents to come to room temperature for 30 minutes.

We recommend that all standards and samples be run in duplicate to allow the end user to

accurately determine oxytocin concentrations.

Ensure that all samples have reached room temperature and have been diluted as appropriate prior to running them in the kit.

Assay Buffer Dilute Assay Buffer Concentrate 1:5 by adding one part of the concentrate to four parts of deion- ized water.

Once diluted this is stable at 4 °C for 3 months.

Wash Buffer Dilute Wash Buffer Concentrate 1:20 by adding one part of the concentrate to nineteen parts of deionized water.

Once diluted this is stable at 4 °C for 3 months.

Standard Preparation Label test tubes as #1 through #8.

Pipet 450 μL of Assay Buffer into tube #1 and 300 μL into the remaining tubes.

The oxytocin stock solution contains an organic solvent.

Prerinse the pipet tip several times to ensure accurate delivery.

Carefully add 50 µL of the oxytocin stock solution to tube #1 and vortex completely.

Take 200 μL of the oxytocin solution in tube #1 and add it to tube #2 and vortex completely.

Repeat the serial dilutions for tubes #3 through #8.

The concentration of oxytocin in tubes 1 through 8 will be 10,000, 4,000, 1,600, 640, 256, 102.4, 40.96 and 16.38 pg/mL.

Use all Standards within 2 hours of preparation.

Sample Preparation:

Serum and Plasma Samples Serum and plasma samples should be extracted with the provided Extraction Solution, or with a solid phase C18 column extraction protocol prior to running in the kit. Protocol Using Extraction Solution: Mix 1 part sample with 1.5 parts of Extraction Solution. Vortex and then nutate at room temperature for 90 minutes. Centrifuge for 20 minutes at 4 °C at 1660 x g. Speedvac supernatant to dryness at 37 °C. Reconstitute sample with 250 μ L of Assay Buffer. Milk Samples Milk samples should be clarified by centrifuging at 10,000 x g for 15 minutes. Pierce the top fatty layer and collect the supernatant liquid. Repeat the centrifugation and collection two more times. The collected supernatant liquid must then be diluted 1:10 with the provided Assay Buffer before using in the assay. The clarified milk sample, i.e., the supernatant liquid, can be stored at -20 °C until needed. Use all samples within 2 hour of preparation.

Assay Procedure:

- 1. Use the plate layout sheet on the back page to aid in proper sample and standard identification. Determine the number of wells to be used and return unused wells to the foil pouch with desiccant. Seal the ziploc plate bag and store at 4°C.
- 2. Pipet 100 μL of samples or standards into wells in the plate.
- 3. Pipet 100 µL of Assay Buffer into wells to act as maximum binding wells (Bo or 0 pg/mL).

- 4. Pipet 125 µL of Assay Buffer into the non-specific binding (NSB) wells.
- 5. Add 25 μ L of the DetectX® Oxytocin Conjugate to each well using a repeater pipet.
- 6. Add 25 μ L of the DetectX® Oxytocin Antibody to each well, except the NSB wells, using a repeater pipet.
- 7. Shake the plate in a plate shaker at room temperature for 15 minutes to ensure adequate mixing of the reagents. Cover the plate with the plate sealer and store at 4 °C for 16-18 hours.
- 8. The following day remove the TMB Substrate from the refrigerator and allow to come to room temperature for at least 30 minutes. Addition of cold Substrate will cause depressed signal.
- 9. Aspirate the plate and wash each well 4 times with 300 μ L wash buffer. Tap the plate dry on clean absorbent towels. 10. Add 100 μ L of the TMB Substrate to each well, using a repeater pipet. 11. Incubate the plate at room temperature for 30 minutes without shaking. 12. Add 50 μ L of the Stop Solution to each well, using a repeater or a multichannel pipet. 13. Read the optical density generated from each well in a plate reader capable of reading at 450 nm. 14. Use the plate reader's built-in 4PLC software capabilities to calculate oxytocin concentration for each sample.

Calculation of Results:

Average the duplicate OD readings for each standard and sample.

Create a standard curve by reducing the data using the 4PLC fitting routine on the plate reader, after subtracting the mean ODs for the NSB.

The sample concentrations obtained, calculated from the %B/B0 curve, should be multiplied by the dilution factor to obtain neat sample values.

Or use the online tool from www.myassays.com/arbor-assays-oxytocin-eia-kit.assay to calculate the data. *The MyAssays logo is a registered trademark of MyAssays Ltd. typical data Sample Mean OD Net OD % B/B0 Oxytocin Conc. (pg/mL) NSB 0.066 0 - - Standard 1 0.128 0.062 10.7 % 10,000 Standard 2 0.163 0.097 16.7 % 4,000 Standard 3 0.222 0.156 26.9 % 1,600 Standard 4 0.301 0.235 40.6 % 640 Standard 5 0.399 0.333 57.5 % 256 Standard 6 0.489 0.423 73.1 % 102.4 Standard 7 0.567 0.501 86.5 % 40.96 Standard 8 0.624 0.558 96.4 % 16.38 B0 0.645 0.579 100 % 0 Sample 1 0.236 0.170 29.3 % 1,336 Sample 2 0.429 0.363 62.7 % 187.9 Always run your own standard curve for calculation of results.

Do not use this data.

Conversion Factor: 1 ng/mL of oxytocin is equivalent to 0.993 nM.

Calibrated to the 4th WHO International Standard NIBSC code: 76/575

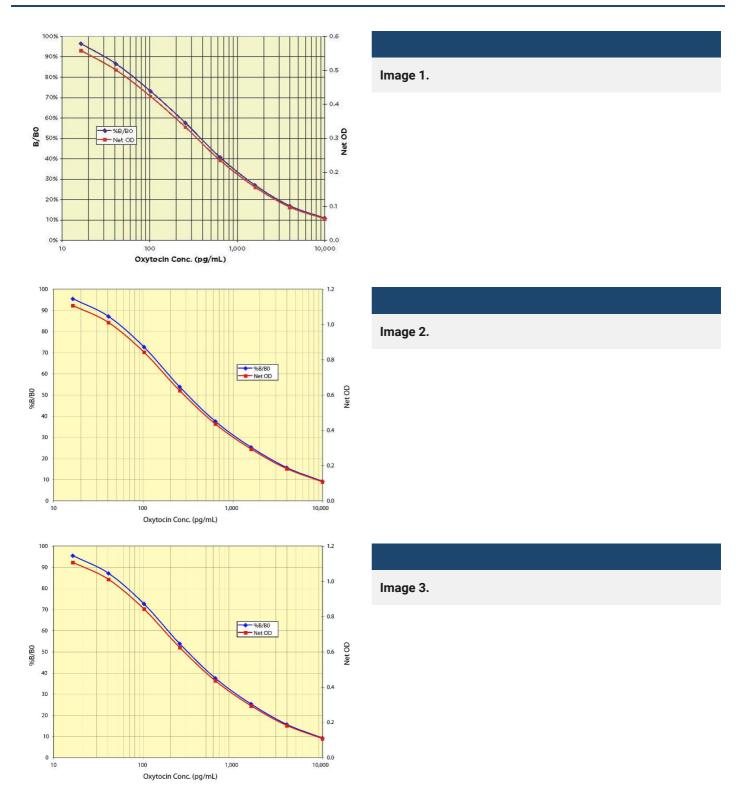
Assay Precision:

Two samples were diluted with Assay Buffer and run in replicates of 20 in an assay. Inter Assay Precision:

Two samples were diluted with Assay Buffer and run in duplicates in 17 assays run over

Application Details

	multiple days by four operators.
Restrictions:	For Research Use only
Handling	
Preservative:	Sodium azide
Precaution of Use:	As with all such products, this kit should only be used by qualified personnel who have had
	labo- ratory safety instruction.
	The complete insert should be read and understood before attempting to use the product.
	The antibody coated plate needs to be stored desiccated.
	The silica gel pack included in the foil ziploc bag will keep the plate dry.
	The silica gel pack will turn from blue to pink if the ziploc has not been closed properly.
	This kit utilizes a peroxidase-based readout system.
	Buffers, including other manufacturers Wash Buffers, containing sodium azide will inhibit color
	production from the enzyme.
	Make sure all buffers used for samples are azide free.
	Ensure that any plate washing system is rinsed well with deionized water prior to using the
	supplied Wash Buffer as prepared on Page 8.
Storage:	4 °C,RT
Storage Comment:	This kit should be stored at 4°C until the expiration date of the kit.
Publications	
Product cited in:	Leeds, Dennis, Lukas, Stoinski, Willis, Schook: "Biologically validating the measurement of
	oxytocin in western lowland gorilla (Gorilla gorilla gorilla) urine and saliva using a commercial
	enzyme immunoassay." in: Primates; journal of primatology , (2018) (PubMed).
	Boose, White, Brand, Meinelt, Snodgrass: "Infant handling in bonobos (Pan paniscus): Exploring
	functional hypotheses and the relationship to oxytocin." in: Physiology & behavior, Vol. 193,
	Issue Pt A, pp. 154-166, (2018) (PubMed).
	Brandtzaeg, Johnsen, Roberg-Larsen, Seip, MacLean, Gesquiere, Leknes, Lundanes, Wilson: "
	Proteomics tools reveal startlingly high amounts of oxytocin in plasma and serum." in:



Please check the product details page for more images. Overall 4 images are available for ABIN2862593.