# antibodies.com

### Datasheet for ABIN365957 CCL27 ELISA Kit

Validation

Image



#### Overview

Quantity:	96 tests
Target:	CCL27
Reactivity:	Human
Method Type:	Sandwich ELISA
Detection Range:	0.313-20 ng/mL
Minimum Detection Limit:	0.313 ng/mL
Application:	ELISA

#### Product Details

Purpose:	This assay employs the quantitative sandwich enzyme immunoassay technique.
Sample Type:	Plasma, Serum, Tissue Homogenate
Analytical Method:	Quantitative
Detection Method:	Colorimetric
Specificity:	This assay has high sensitivity and excellent specificity for detection of Human CCL27.
Cross-Reactivity (Details):	Limited by current skills and knowledge, it is impossible for us to complete the cross-reactivity detection between the target antigen and all analogues for other species. Therefore, cross reaction may still exist.
Sensitivity:	0.043 ng/mL
Components:	<ul> <li>Assay plate (12 × 8 coated Microwells)</li> <li>Standard (freeze dried)</li> </ul>

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- Biotin-antibody (100 × concentrate)
- HRP-avidin (100 × concentrate)
- Biotin-antibody Diluent
- HRP-avidin Diluent
- Sample Diluent
- Wash Buffer (25 × concentrate)
- TMB Substrate
- Stop Solution
- Adhesive Strip (for 96 wells)
- Instruction manual

#### Target Details

Target:	CCL27
Alternative Name:	C-C motif chemokine 27 (CCL27) (CCL27 Products)
Background:	Synonyms: ALP, CTACK, CTAK, ESKINE, ILC, PESKY, SCYA27, CC chemokine ILC IL-11 Ralpha- locus chemokine cutaneous T-cell attracting chemokine skinkine small inducible cytokine A27 small inducible cytokine subfa
HGNC:	10626
UniProt:	Q9Y4X3

#### Application Details

upplier is only responsible for the kit itself, but not for the samples consumed during the . The user should calculate the possible amount of the samples used in the whole test. It ereserve sufficient samples in advance. Hes to be used within 5 days may be stored at 2-8°C, otherwise samples must be stored of $(\leq 1 \text{ month})$ or -80°C ( $\leq 2 \text{ months}$ ) to avoid loss of bioactivity and contamination.
e reserve sufficient samples in advance. les to be used within 5 days may be stored at 2-8°C, otherwise samples must be stored °C (≤ 1 month) or -80°C (≤ 2 months) to avoid loss of bioactivity and contamination.
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°C ( $\leq$ 1 month) or -80°C ( $\leq$ 2 months) to avoid loss of bioactivity and contamination.
ly hemolyzed samples are not suitable for use in this assay.
samples are not indicated in the manual, a preliminary experiment to determine the
y of the kit is necessary.
e predict the concentration before assaying. If values for these are not within the range
standard curve, users must determine the optimal sample dilutions for their particular
ments.
e or cell extraction samples prepared by chemical lysis buffer may cause unexpected
results due to the impacts of certain chemicals.
to the possibility of mismatching between antigens from another resource and
dies used in this supplier's kits (e.g., antibody targets conformational epitope rather
near epitope), some native or recombinant proteins from other manufacturers may not
ognized by this supplier's products.

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	<ul> <li>Influenced by factors including cell viability, cell number and cell sampling time, samples from cell culture supernatant may not be recognized by the kit.</li> <li>Fresh samples without long time storage are recommended for the test. Otherwise, protein degradation and denaturalization may occur in those samples and finally lead to wrong results.</li> </ul>
Comment:	Detection wavelength: 450 nm
	Information on standard material:
	Depending on the antigen to be detected, standards can be either native or recombinant
	protein. The recombinant proteins are being expressed in CHO cells in most cases. Please
	inquire for more information. The formulation of auxiliary material in the standard is considered
	proprietary information, however it does not contain any poisonous substance. Proclin 300
	(1:3000) is used as preservative.
	Information on reagents:
	In most cases the stop solution provided is 1 N H2SO4. The formulation of wash solution is
	proprietary information. None of the components contain (sodium) azide, thimerosal, 2-
	mercaptoethanol (2-ME) or any other poisonous materials. For the sandwich method kits, the
	sample diluent, antibody diluent, enzyme diluent and standard all contain BSA.
	Information on antibodies:
	The antibodies provided in different kits vary in regards to clonality and host. Some antibodies
	are affinity purified, some are Protein A
Sample Volume:	100 µL
Assay Time:	1 - 4.5 h
Plate:	Pre-coated
Protocol:	Antibody specific for CCL27 has been pre-coated onto a microplate. Standards and samples
	are pipetted into the wells and any CCL27 present is bound by the immobilized antibody. After
	removing any unbound substances, a biotin-conjugated antibody specific for CCL27 is added to
	the wells. After washing, avidin conjugated Horseradish Peroxidase (HRP) is added to the wells
	Following a wash to remove any unbound avidin-enzyme reagent, a substrate solution is addec
	to the wells and color develops in proportion to the amount of CCL27 bound in the initial step.
	The color development is stopped and the intensity of the color is measured.

Reagent Preparation:

• **Biotin-antibody (1×)** - Centrifuge the vial before opening.

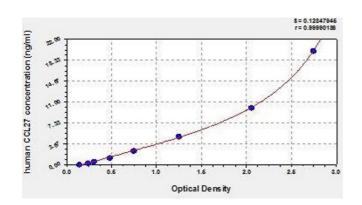
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Biotin-antibody requires a 100-fold dilution. The suggested dilution is 10µL of Biotin-antibody + 990µL of Biotin-antibody Diluent. HRP-avidin (1×) - Centrifuge the vial before opening. HRP-avidin requires a 100-fold dilution. The suggested dilution is 10µL of HRP-avidin + 990µL of HRP-avidin Diluent. Wash Buffer (1x) - If crystals have formed in the concentrate, warm up to room temperature and mix gently until the crystals have completely dissolved. Dilute 20mL of Wash Buffer Concentrate (25x) into deionized or distilled water to prepare 500mL of Wash Buffer (1x). Standard - Centrifuge the standard vial at 6000-10000rpm for 30s. Reconstitute the Standard with 1ml of Sample Diluent. Do not substitute other diluents. This reconstitution produces a stock solution of 200pg/mL. Mix the standard to ensure complete reconstitution and allow the standard to sit for a minimum of 15 minutes with gentle agitation prior to making dilutions. Pipette 250µL of Sample Diluent into each tube. Use the stock solution to produce a 2-fold dilution series. Mix each tube thoroughly before the next transfer. The undiluted Standard serves as the high standard (200pg/mL). Sample Diluent serves as the zero standard (0ng/mL). Note: Kindly use graduated containers to prepare the reagent. Please don't prepare the reagent directly in the Diluent vials provided in the kit. Bring all reagents to room temperature (18-25°C) before use for 30 min. • Prepare fresh standard for each assay. Use within 4 hours and discard after use. Making serial dilution in the wells directly is not permitted. Please carefully reconstitute Standards according to the instruction. Avoid foaming and mix gently until the crystals have completely dissolved. To minimize imprecision caused by pipetting, use small volumes and ensure that pipettors are calibrated. It is recommended to suck more than 10µL when pipetting. It is recommended to use distilled water to prepare reagents and samples. Using contaminated water or container for reagent preparation will influence detection result. Assay Precision: Intra-assay precision (precision within an assay): Three samples of known concentration were tested twenty times on one plate to assess precision. Inter-assay precision (precision between assays): Three samples of known concentration were tested in twenty assays to assess precision. Intra-assay: CV% less than 8% Inter-assay: CV% less than 10% **Restrictions:** For Research Use only Handling Precaution of Use: The Stop Solution provided with this kit is an acid solution. Wear eye, hand, face and clothing

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	protection when using this material.
Handling Advice:	<ul> <li>The kit should not be used beyond the expiration date on the kit label.</li> <li>Do not mix or substitute reagents with those from other lots or sources.</li> <li>If samples generate values higher than the highest standard, dilute the samples with Sample Diluent and repeat the assay.</li> <li>Any variation in Sample Diluent, operator, pipetting technique, washing technique, incubation time/temperature and kit age can cause variation in binding.</li> <li>This assay is designed to eliminate interference by soluble receptors, binding proteins and other factors present in biological samples. Until all factors have been tested in the Immunoassay, the possibility of interference cannot be excluded.</li> </ul>
Storage:	4 °C/-20 °C
Storage Comment:	For unopened kit: All the reagents should be kept according to the labels on vials.
Expiry Date:	6 months

Images



ELISA

Image 1. Typical standard curve

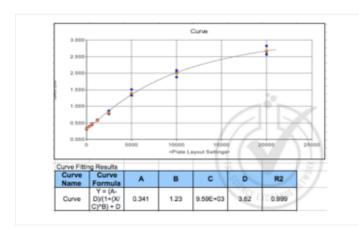
	Successfully validated (ELISA (ELISA))
	by CGIBD Advanced Analytics Core
	Report Number: 029788
ENTIN NO	Date: Aug 12 2014
REPRODUCIBILITY INITIATIVE NO.: 829788 DATE: 08/12/14	
Lot Number:	X19184064
Method validated:	ELISA (ELISA)
Positive Control:	Human serum - expression is ~650 pg/mL
Negative Control:	Goat serum (non-reactive species)
Notes:	Target protein was detected in the positive control sample and not in the negative control
	sample as expected.
Controls:	• Positive control: Human serum (Sigma Aldrich, Cat# H6914-20ML, Lot# SLBK2170V)
	Negative control: Goat serum (Sigma Aldrich, Cat# G9023-10ML, Lot# SLBH2670V)
Protocol:	<ul> <li>1. All reagents were brought up to room temperature for 30 minutes prior to use. The 1x Wash Buffer was prepared by adding 20 mL of 25x Wash Buffer Concentrate to 480 mL of distilled/deionized water and mixing thoroughly.</li> <li>2. The vial of Standard was reconstituted with 1 mL of Sample Diluent, mixed, and allowed to sit for 15 minutes with gentle agitation.</li> <li>3. The standard curve was prepared by creating a 2-fold dilution series of seven standards (including the original undiluted vial) using Sample Diluent. Sample Diluent alone served as the 0 pg/mL standard.</li> <li>4. The assay plate was removed from the foil pouch and 100 µL of each standard and sample were added to the appropriate wells, in triplicate. The plate was covered with the adhesive strip provided and incubated for 2 hours at 37 °C.</li> <li>5. Approximately 10 minutes before the incubation ended, a 1x Biotin-antibody Diluent.</li> <li>6. The liquid from each well was removed.</li> <li>7. 100 µL of 1x Biotin-antibody solution was added to each well, and the plate was covered with a new adhesive strip, and incubated for 1 hour at 37 °C.</li> <li>8. Approximately 10 minutes before the incubation ended, a 1x HRP-avidin solution was prepared by diluting 60 µL of 100x HRP-avidin into 5940 µL of HRP-avidin Diluent.</li> <li>9. Each well was aspirated and washed, repeating the process two times for a total of three washes. Each well was washed by filling each well with 1x Wash Buffer and letting it stand for 2 minutes. After the last wash, remaining Wash Buffer was removed and the plate was inverted and blotted against clean, absorbent paper towels.</li> <li>10. 100 µL of 1x HRP-avidin solution was added to each well, the plate was covered with a</li> </ul>

new adhesive strip, and incubated for 1 hour at 37 °C.

- 11. The aspiration/wash procedure from Step 9 was repeated for an additional 5 washes.
- 12. 90 µL of TMB Substrate was added to each well. The plate was protected from light and incubated for 15-30 minutes at 37 °C, with periodic checking to prevent overdevelopment.
- 13. 50 µL of Stop Solution was added to each well and mixed thoroughly. The optical density (OD) of each well was measured within 5 minutes using a microplate reader set to 450 nm.
- 14. A standard curve was generated by plotting the OD value for each standard on the y-axis against the concentration on the x-axis. A line of best fit through the points on the graph was used to generate an equation to calculate CLA concentrations of the samples based on their average OD values.

Experimental Notes: Well B1 was excluded from this dataset due to possible contamination. Other than that, there were no experimental challenges noted.

#### Images for Validation report #029788



## Validation image no. 1 for Chemokine (C-C Motif) Ligand 27 (CCL27) ELISA Kit (ABIN365957)

Figure 1: CLA standard curve graph and equation.

ayout							
	7		9	10	11	12	
	STD1	\$TD1	STD1	SPL11	SPL11	SPL11	140
	0	0	0	1	1	1	Constitu
				Human	Human	Human	here
				serum	Herum	BRF/JTT	-
		8702	STC2	90000 90(12	SPL12	SPL12	1-40
		312.5	312.5	2	2	2	Consta
				Human	Human	Human	here
				serum	Herum	B01/JT	-
	STD3	8703	8103	800	84(13	84113	1-40
c	625	625	625	5	5	5	Consta
				Human	Human	Human	-
				serum	Herum	Berum	-
	8104	87D4	8104	SPL14	SPL14	57114	1-40
D	1250	1250	1250	10	10	10	Come Die
				Human	Human	Human	
				Berum	Herum	Berum	-
	8106	5705	8106	54115	5PL15	57115	1000
	2500	2500	2500	20	20	20	Constit
				Human	Human	Human	-
				serum	MPUm.	Minute .	000
	STD6	5706	STD6	5PL16	/ 5PL18	5416	140
	5000	5000	5000	40 /	/40	40	Conto
· ·				Human	/Human	Human	2
				serum	MOUTH .	MATURE .	- T
	5107	5107	5107	5PL17	SPL17	54117	1-40
a	10000	10000	10000	50	50	50	Conto
				Human	Human	Human	lain .
				serum	- seitum	serum/	100
	STDe	STDe	STDe	5PL2	SPL2	5912	140
н	20000	20000	20000	1		1	Louis
				Goat	Goal	Goal	1
				serum.	sofun.	BROWN'	100

### Validation image no. 2 for Chemokine (C-C Motif) Ligand 27 (CCL27) ELISA Kit (ABIN365957)

Figure 2: Plate layout. Standard concentrations are in pg/mL; serum dilution values indicate their fold change from the undiluted stock. Note that well B1 was excluded from this dataset due to possible contamination.

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RAW DW	7	8	9	10		12	
A	0.326	0.349	0.312	0.969	1.032	0.993	RAWOW
В		0.403	0.39	0.729	0.688	0.716	RANOW
С	0.491	0.486	0.439	0.441	0.437	0.374	RAWOW
D	0.591	0.594	0.572	0.267	0.269	0.278	RAN DW
E	0.806	0.876	0.772	0.146	0.152	0.148	BAW DW
F	1.517	1.332	1.377	0.096	0.092	0.096	RANOW
G	1.885	2.085	2.018	0.07	0.085	0.097	RANOW
н	2.65	2.833	2.57	0.025	0.024	0.038	RAW OW

### Validation image no. 3 for Chemokine (C-C Motif) Ligand 27 (CCL27) ELISA Kit (ABIN365957)

Figure 3: Raw OD readings of standards and controls. Note that well B1 was excluded from this dataset due to possible contamination.

Conc							
	7	8	9	10	11	12	
Α	<0.000	73.123	<0.000	3076.408	3284.565	3095.618	Cone
В		389.595	320.865	1880.157	1697.833	1822.256	Cone
С	815.766	792.614	569.938	579.649	560.201	231.943	Conc
D	1267.481	1280.832	1182.759	<0.000	<0.000	<0.000	Conc
E	2225.716	2545.285	2072.498	<0.000	<0.000	<0.000	Conc
F	5981.863	4862.582	5122.738	< 0.000	<0.000	<0.000	Conc
G	8718.388	10627.61	9945.012	< 0.000	<0.000	<0.000	Conc
н	19351.02	>21000.00	17636.95	<0.000	<0.000	<0.000	Cont
						- 2	
onc x Dil					1		
	7	8	9	10	/ 11_	12	
A				3076.408	3284.565	3095,618	Conc k Di
B				3076.408	3284.565 3395.665		
						3644.513	Conc x Di
B				3760.314	3395.665	3644.513 1159.713	Cone + Di Cone + Di
BC				3760.314 2898.244	3395.665 2801.004	3644.513 1159.713 <0.000	Concis De Concis De Concis De
B C D				3760.314 2898.244 <0.000	3395.665 2801.004 <0.000	3644.513 1159.713 <0.000 <0.000	Cone + DA Cone + DA Cone + DA
B C D				3760.314 2898.244 <0.000 <0.000	3395.665 2801.004 <0.000 <0.000	3644.513 1159.713 <0.000 <0.000	Core a Da Core a Da Core a Da Core a Da Core a Da Core a Da

### Validation image no. 4 for Chemokine (C-C Motif) Ligand 27 (CCL27) ELISA Kit (ABIN365957)

Figure 4: CLA concentrations calculated from standard curve formula. Upper panel = uncorrected for dilution; lower panel = corrected for dilution. On average, 3013 pg/mL of CLA was detected in the positive control (human serum) and 0 pg/mL of CLA was detected in the negative control (goat serum).