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Datasheet for ABIN427274 BIRC2 ELISA Kit



Overview

Quantity:	96 tests
Target:	BIRC2
Reactivity:	Mouse
Application:	ELISA

Product Details

Sample Type:	Plasma, Serum
Detection Method:	Colorimetric
Sensitivity:	The sensitivity of this assay, or Lower Limit of Detection (LLD) was defined as the lowest protein concentration that could be differentiated from zero. It was determined the mean O.D. Value of 20 replicates of the zero standard added by their three standard deviations.
Material not included:	 Microplate reader with 450nm filter. Precision single or multi-channel pipettes and disposable tips. Eppendorf Tubes for diluting samples. Deionized or distilled water. Absorbent paper for blotting the microtiter plate. Container for Wash Solution

Target Details

Target:	BIRC2
Alternative Name:	Baculoviral IAP Repeat Containing Protein 2 (BIRC2) (BIRC2 Products)
Pathways:	Apoptosis, Caspase Cascade in Apoptosis, Activation of Innate immune Response, Toll-Like

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Receptors Cascades

Application Details

pur products with other manufacturers.
aterial:
ombinant protein or natural protein, that will depend on the specific
on system is E.coli or yeast or mammal cell. There is 0.05% proclin
servative.

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solution is TBS. The standard diluent contains 0.02 % sodium azide, assay diluent A and assay	
diluent B contain 0.01% sodium azide. Some kits can contain is BSA in them.	

Information on antibodies:

The provided antibodies and their host vary in different kits.

Plate:	Pre-coated
Reagent Preparation:	 Bring all kit components and samples to room temperature (18-25°C) before use. Standard - Reconstitute the Standard with the Standard Diluent, keep at room temperature for 10 min and shake gently (not to foam). Prepare tubes containing Standard Diluent to produce a double dilution series. Assay Diluent A and Assay Diluent B - Dilute 6mL of Assay Diluent A or B Concentrate(2×) with 6mL of deionized or distilled water to prepare 12 mL of Assay Diluent A or B. (In fact, more than 6mL Assay Diluent A and Assay Diluent a more than 6mL Assay Diluent A and Assay Diluent B are contained in the bottles. Therefore, in every test, please precisely pipette required amount of Diluent and make double dilution in a new container. The prepared working dilution cannot be frozen.) Detection Reagent A and Detection Reagent B - Briefly spin or centrifuge the stock Detection A and Detection B before use. Dilute to the working concentration with working Assay Diluent A or B, respectively (1:100). Wash Solution - Dilute 20mL of Wash Solution concentrate (30×) with 580mL of deionized or distilled water to prepare 600 mL of Wash Solution (1×). TMB substrate - Aspirate the needed dosage of the solution with sterilized tips and do not
	 dump the residual solution into the vial again. Note: Making serial dilution in the wells directly is not permitted. Prepare standard within 15 minutes before assay. Please do not dissolve the reagents at
	 37°C directly. Please carefully reconstitute Standards or working Detection Reagent A and B according to the instruction, and avoid foaming and mix gently until the crystals are completely dissolved. To minimize imprecision caused by pipetting, use small volumes and ensure that pipettors are calibrated. It is recommended to suck more than 10µL for once pipetting. The reconstituted Standards, Detection Reagent A and Detection Reagent B can be used on once.
	 If crystals have formed in the Wash Solution concentrate (30×), warm to room temperature.
Sample Collection:	Serum: Allow samples to clot for two hours at room temperature or overnight at 4°C before centrifugation for 20 minutes at approximately 1000 × g. Assay immediately or store samples in aliquot at -20°C or -80°C. Avoid repeated freeze/thaw cycles.
	Plasma: Collect plasma using EDTA or heparin as an anticoagulant. Centrifuge samples for 15 minutes at 1000 × g within 30 minutes of collection. Remove plasma and assay immediately c

store samples in aliquot at -20°C or -80°C. Avoid repeated freeze/thaw cycles.

Biological Fluids: Centrifuge samples for 20 minutes at $1000 \times g$. Remove particulates and assay immediately or store samples in aliquot at -20 °C or -80 °C for later use. Avoid repeated freeze/thaw cycles.

Sample Preparation:	Notes:
	• The user should calculate the possible amount of the samples used in the whole test. Please
	reserve sufficient samples in advance.
	 Please predict the concentration before assaying. If values for these are not within the range of the standard curve, users must determine the optimal sample dilutions for their particular experiments.
	 If the samples are not indicated in the manual, a preliminary experiment to determine the validity of the kit is necessary.
	 Tissue or cell extraction samples prepared by chemical lysis buffer may cause unexpected ELISA results due to the impacts from certain chemicals.
	 Due to the possibility of mismatching between antigen from other origin and antibody used i our kits (e.g.,antibody targets conformational epitope rather than linear epitope), some native or recombinant proteins from other manufacturers may not be recognized by our products. Influenced by the factors including cell viability, cell number or sampling time, samples from the recognized by the factors including cell viability.
	 cell culture supernatant may not be detected by the kit. Fresh samples without long time storage is recommended for the test. Otherwise, protein degradation and denaturalization may occur in those samples and finally lead to wrong results.
Assay Procedure:	 1. Determine wells for diluted standard, blank and sample. Prepare wells for standard points 1 well for blank. Add dilutions of standard (read Reagent Preparation), blank and samples into the appropriate wells, respectively. You might now need to incubate at 37 °C. And then add Detection Reagent A to each well. You might need to incubate at 37 °C again. 2. Aspirate the solution and wash with 350 µL of 1X Wash Solution to each well using a squi
	bottle, multi-channel pipette, manifold dispenser or autowasher, and let it sit for 1-2 minutes Remove the remaining liquid from all wells completely by snapping the plate onto absorbent
	paper. Repeat 3 times. After the last wash, remove any remaining Wash Buffer by aspirating or decanting. Invert the plate and blot it against absorbent paper.
	 3. Add 100 µL of Detection Reagent B working solution to each well. Incubate for 30 minutes at 37 °C after covering it with the Plate sealer.
	 4. Repeat the aspiration/wash process for total 5 times as conducted in step 2.
	 5. Add 90 μL of Substrate Solution to each well. Cover with a new Plate sealer. Incubate for 15 - 25 minutes at 37 °C (Don't exceed 30 minutes). Protect from light. The liquid will turn
	blue by the addition of Substrate Solution.
	• 6. Add 50 μ L of Stop Solution to each well. The liquid will turn yellow by the addition of Stop
	solution. Mix the liquid by tapping the side of the plate. If color change does not appear uniform, gently tap the plate to ensure thorough mixing.

 7. Remove any drop of water and fingerprint on the bottom of the plate and confirm there is no bubble on the surface of the liquid. Then, run the microplate reader and conduct measurement at 450 nm immediately.

Note:

- 1. Assay preparation: Keep appropriate numbers of wells for each experiment and remove extra wells from microplate. Rest wells should be resealed and stored at -20 °C.
 2. Samples or reagents addition: Please use the freshly prepared Standard. Please carefully add samples to wells and mix gently to avoid foaming. Do not touch the well wall. For each step in the procedure, total dispensing time for addition of reagents or samples to the assay plate should not exceed 10 minutes. This will ensure equal elapsed time for each pipetting step, without interruption. To avoid cross-contamination, change pipette tips between additions of standards, samples, and reagents. Also, use separated reservoirs for each reagent.
- 3. Incubation: To ensure accurate results, proper adhesion of plate sealers during incubation steps is necessary. Do not allow wells to sit uncovered for extended periods between incubation steps. Once reagents are added to the well strips, DO NOT let the strips DRY at any time during the assay. Incubation time and temperature must be controlled.
- 4. Washing: The wash procedure is critical. Complete removal of liquid at each step is
 essential for good performance. After the last wash, remove any remaining Wash Solution by
 aspirating or decanting and remove any drop of water and fingerprint on the bottom of the
 plate. Insufficient washing will result in poor precision and false elevated absorbance
 reading.
- 5. Controlling of reaction time: Observe the change of color after adding TMB Substrate (e.g. observation once every 10 minutes), if the color is too deep, add Stop Solution in advance to avoid excessively strong reaction which will result in inaccurate absorbance reading.
- 6. TMB Substrate is easily contaminated. Please protect it from light.
- 7. The environment humidity which is less than 60 % might have some effects on the final performance, therefore, a humidifier is recommended to be used at that condition.

Calculation of Results:	Average the duplicate readings for each standard, control, and samples and subtract the
	average zero standard optical density. Construct a standard curve by plotting the mean O.D.
	and concentration for each standard and draw a best fit curve through the points on the graph
	or create a standard curve on log-log graph paper with the target concentration on the y-axis
	and absorbance on the x-axis. Draw the best fit straight line through the standard points and it
	can be determined by regression analysis. Using some plot software, for instance, curve expert
	1.30, is also recommended. If samples have been diluted, the concentration read from the
	standard curve must be multiplied by the dilution factor.
Assay Precision:	Intra-assay Precision (precision within an assay): Three samples with low, medium and high
	levels of the target antigen were tested twenty times on one plate, respectively.

Inter-assay Precision (precision between assays): Three samples with low, medium and high

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Application Details	
	 levels of the target antigen were tested on three different plates, eight replicates in each plate. CV (%) = SD/mean X 100 Intra-assay: CV less than 10 % Inter-assay: CV less than 12 %
Restrictions:	For Research Use only
Handling	
Precaution of Use:	The Stop Solution suggested for use with this kit is an acid solution. Wear eye, hand, face, and clothing protection when using this material.
Handling Advice:	To minimize extra influence on the performance, operation procedures and lab conditions, especially room temperature, air humidity, incubator temperature should be strictly controlled. I is also strongly suggested that the whole assay is performed by the same operator from the beginning to the end.
Storage:	4 °C/-20 °C
Storage Comment:	 For unopened kit: All the reagents should be kept according to the labels on vials. The Standard, Detection Reagent A, Detection Reagent B and the 96-well strip plate should be stored at -20°C upon receipt while the others should be at 4°C. For opened kit: When the kit is opened, the remaining reagents still need to be stored according to the above storage condition. Besides, please return the unused wells to the foil pouch containing the desiccant pack, and reseal along entire edge of zip-seal. Note: It is highly recommended to use the remaining reagents within 1 month provided this is within the expiration date of the kit. For ELISA kit, 1 day storage at 37°C can be considered as 2 months at 4°C, which means 3 days at 37°C equaling 6 months at 4°C.
Expiry Date:	6 months