Datasheet for ABIN5067556

Lipid Droplet Isolation Kit

Go to Product page

1 Image

Overview

Quantity:	50 preparations
Application:	Purification (Purif)

Product Details

Sample Type:	Cell Samples, Tissue Samples
Characteristics:	Lipid Droplet Isolation Kit isolates lipid droplets by simple gradient centrifugation, but circumvents the need for large sample sizes or ultracentrifugation. A lipid droplet source such as tissue or cultured cells is homogenized. A gradient is then created with the homogenate, and the material is centrifuged. The lipid droplets float to the top of the gradient and are recovered by carefully pipetting from the top of the gradient. Each kit provides sufficient reagents to isolate up to 50 preps based on a 50-100 mg tissue or cultured cell sample size.
Components:	1. Reagent A : One 10 mL bottle. 2. 10X Reagent B : One 7 mL bottle. 2

Target Details

Background:	Lipid droplets are organelles that are rich in lipids, contain a lipid rich core, and are surrounded by a phospholipid monolayer as well as outer lipid droplet associated proteins. Lipid droplets are commonly found in adipose tissue of animals, although they are found in all eukaryotes. Lipid droplets function to regulate the hydrolysis and storage of neutral lipids and also serve as storage for cholesterol and acyl- glycerols used to form and maintain cellular membranes. Beyond the function of lipid and cholesterol storage, lipid droplet organelles have been more recently associated with inflammatory responses, obesity, atherosclerosis, and cancer. Lipid droplets have been shown to protect against lipotoxicity in non-adipocytes by storing fatty

Target Details

acids as neutral triacylglycerol. Lipid droplets also aid in protein binding and degradation and have been demonstrated to be used by pathogens such as dengue virus and hepatitis C Virus.

Application Details

Application Notes:	Optimal working dilution should be determined by the investigator.
Comment:	• Isolates lipid droplets from cells or tissues • Uses a simple gradient centrifugation protocol • No ultracentrifugation required
Protocol:	I. Isolation from Cultured Cells 7 1. Trypsinize 1.5-3 x 10 cells (roughly 50-100 mg) and resuspend in 10 mL of growth media in a 15 mL polypropylene tube. 2. Pellet cells at 1000 x g for 5 minutes. 3. Aspirate the media and wash cells with 10 mL of 1X PBS. 4. Pellet cells again at 1000 x g for 5 minutes. 5. Aspirate media and add 1 mL of 1X PBS. 6. Resuspend cells thoroughly and transfer to a 2 mL microcentrifuge tube. 7. Pellet cells again at 1000 x g for 5 minutes. 8. Aspirate 1X PBS and resuspend pellet thoroughly with 200 µL of Reagent A. 9. Incubate on ice for 10 minutes. 10. Add 800 µL of 1X Reagent B and mix well. 11. Incubate on ice for 10 minutes. 12. Homogenize the cells by passing them five times through a one inch 27 gauge needle attached to a 3 mL syringe. 13. Briefly spin the homogenate at 100 x g for 5 seconds 3 14. Carefully layer 600 µL of 1X Reagent B on top of the homogenate by dropwise addition taking care not to disturb the homogenate. 15. Spin the 2 mL microcentrifuge tube in a microcentrifuge for 3 hours at 18000-20000 x g at 4°C. 16. Carefully remove 270 µL (containing the floating lipid droplets) from the top of the tube and transfer to a fresh microcentrifuge tube. 17. Store lipid droplets at -80°C. II. Isolation from Tissue by Dounce Homogenization 1. Weigh out 50-100 mg of tissue and mince into small pieces with a scalpel or scissors. 2. Transfer minced tissue to a glass dounce. 3. Add 200 µL of Reagent A. 4. Incubate on ice for 10 minutes. 5. Add 800 µL of 1X Reagent B. 6. Incubate on ice for 10 minutes. 7. Homogenize the tissue by performing 5 up and down strokes with the loose (A) pestle followed by 5 up and down strokes with the tight (B) pestle. 8. Transfer 1 mL of the homogenate to a 2 mL microcentrifuge tube

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Application Details

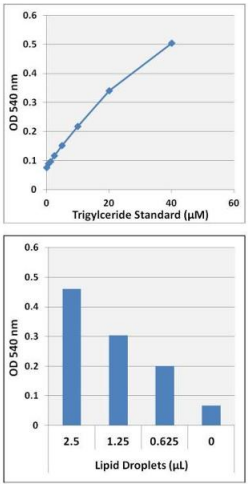
9. Carefully layer 600 µL of 1X Reagent B on top of the homogenate by dropwise addition taking care not to disturb the homogenate.
10. Spin the 2 mL microcentrifuge tube in a microcentrifuge for 3 hours at 18000-20000 xg at 4°C.
11. Carefully remove 270 µL (containing the floating lipid droplets) from the top of the tube and transfer to a fresh microcentrifuge tube.
12. Store lipid droplets at -80°C.

Reagent Preparation:	1X Reagent B: Dilute 10X Reagent B to 1X with deionized water. Stir to homogeneity.
Restrictions:	For Research Use only

Handling

Storage:	4 °C
Storage Comment:	Store the entire kit at 4°C. To avoid possible leakage store bottles upright.

Images



Biochemical Assay

Image 1. Triglyceride Quantification Kit (Colorimetric) (Cat. #STA-396) Performed on Extracted Lipids. (Top) Triglyceride Standard Curve. (Bottom) Lipid droplets isolated from Chicken Liver were tested for the presence of Triglyceride according to the Assay Protocol.