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Datasheet for ABIN625124 G-CSF ELISA Kit

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Overview

Quantity:	96 tests
Target:	G-CSF (CSF3)
Reactivity:	Mouse
Method Type:	Sandwich ELISA
Detection Range:	0.5-150 pg/mL
Minimum Detection Limit:	0.5 pg/mL
Application:	ELISA

Product Details

Purpose:	Mouse GCSF ELISA Kit for cell culture supernatants, plasma, and serum samples.
Sample Type:	Serum, Plasma, Cell Culture Supernatant
Analytical Method:	Quantitative
Detection Method:	Colorimetric
Specificity:	This ELISA kit shows no cross-reactivity with any of the cytokines tested: Mouse CD30, L CD30, T CD40, CRG-2, CTACK, CXCL16, Eotaxin , Eotaxin-2, Fas Ligand, Fractalkine, GM-CFS, IFN- gamma, IGFBP-3, IGFBP-5, IGFBP-6, IL-1 alpha, IL-1 beta, IL-2, IL-3, IL-3 Rb, IL-4, IL-5, IL-6, IL-9, IL-10, IL-12 p40/p70, IL-12 p70, IL-13, IL-17, KC, Leptin R, LEPTIN(OB), LIX, L-Selectin, Lymphotactin, MCP-1, MCP-5, M-CSF, MIG, MIP-1 alpha, MIP-1 gamma, MIP-2, MIP-3 beta, MIP- 3 alpha, PF-4, P-Selectin, RANTES, SCF, SDF-1 alpha, TARC, TCA-3, TECK, TIMP-1, TNF-alpha, TNF RI, TNF RII, TPO, VCAM-1, VEGF.
Sensitivity:	< 0.5 pg/mL

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Product Details

Characteristics:	 Strip plates and additional reagents allow for use in multiple experiments Quantitative protein detection Establishes normal range The best products for confirmation of antibody array data
Components:	Pre-Coated 96-well Strip Microplate
	• Wash Buffer
	Stop Solution
	Assay Diluent(s)
	Lyophilized Standard
	Biotinylated Detection Antibody
	Streptavidin-Conjugated HRP
	TMB One-Step Substrate
Material not included:	Distilled or deionized water
	- Precision pipettes to deliver 2 μ L to 1 μ L volumes
	 Adjustable 1-25 µL pipettes for reagent preparation
	 100 µL and 1 liter graduated cylinders
	 Tubes to prepare standard and sample dilutions
	Absorbent paper
	Microplate reader capable of measuring absorbance at 450nm
	Log-log graph paper or computer and software for ELISA data analysis

Target Details

Target:	G-CSF (CSF3)
Alternative Name:	G-CSF/ CSF3 (CSF3 Products)
Background:	G-CSF is an O-glycosylated glycoprotein, which is secreted by monocytes, macrophages and
	neutrophils. G-CSF is also produced by stromal cells, fibroblasts, and endothelial cells. Epithelial
	carcinomas, acute myeloid leukemia cells and various tumor cell lines, also express this factor.
	G-CSF stimulates the proliferation and differentiation of hematopoietic progenitor cells
	committed to the neutrophil/granulocyte lineage. It is a mitogen for some human myeloid
	leukemia cells and also for some carcinoma cell lines. In vitro G-CSF enhances the antibody-
	dependent cellular cytotoxicity of granulocytes against tumor cells. The Mouse G-CSF ELISA
	(Enzyme-Linked Immunosorbent Assay) kit is an in vitro enzyme-linked immunosorbent assay
	for the quantitative measurement of mouse G-CSF in serum, plasma, cell culture supernatants
	and urine. This assay employs an antibody specific for mouse G-CSF coated on a 96-well plate.
	Standards and samples are pipetted into the wells and G-CSF present in a sample is bound to
	the wells by the immobilized antibody. The wells are washed and biotinylated anti-mouse G-

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	CSF antibody is added. After washing away unbound biotinylated antibody, HRP-conjugated
	streptavidin is pipetted to the wells. The wells are again washed, a TMB substrate solution is
	added to the wells and color develops in proportion to the amount of G-CSF bound. The Stop
	Solution changes the color from blue to yellow, and the intensity of the color is measured at
	450 nm. Reproducibility: Intra-Assay: CV<10% Inter-Assay: CV<12%.
Gene ID:	12985
UniProt:	P09920
Pathways:	Cellular Response to Molecule of Bacterial Origin, Regulation of Actin Filament Polymerization

Application Details

Application Notes:	Recommended Dilution for serum and plasma samples20 - 100 fold
Sample Volume:	100 μL
Plate:	Pre-coated
Protocol:	1. Prepare all reagents, samples and standards as instructed in the manual.
	2. Add 100 μ L of standard or sample to each well.
	3. Incubate 2.5 h at RT or O/N at 4 °C.
	4. Add 100 μ L of prepared biotin antibody to each well.
	5. Incubate 1 h at RT.
	6. Add 100 μ L of prepared Streptavidin solution to each well.
	7. Incubate 45 min at RT.
	8. Add 100 µL of TMB One-Step Substrate Reagent to each well.
	9. Incubate 30 min at RT.
	10. Add 50 μL of Stop Solution to each well.
	11. Read at 450 nm immediately.
Reagent Preparation:	1. Bring all reagents and samples to room temperature (18 - 25 °C) before use.
	2. Sample dilution: If your samples need to be diluted, Assay Diluent A (Item D) should be used
	for dilution of serum/plasma samples. 1x Assay Diluent B (Item E) should be used for dilution o
	culture supernatants. Suggested dilution for normal serum/plasma: 20-100 fold*. * Please note
	that levels of the target protein may vary between different specimens. Optimal dilution factors
	for each sample must be determined by the investigator.
	3. Assay Diluent B should be diluted 5-fold with deionized or distilled water.
	4. Preparation of standard: Briefly spin the vial of Item C and then add 400 μ L Assay Diluent A
	(for serum/plasma samples) or 1x Assay Diluent B (for cell culture medium) into Item C vial to
	prepare a 50 ng/mL standard. Dissolve the powder thoroughly by a gentle mix. Add 2 μL G-CSF

	standard from the vial of Item C, into a tube with 664.7 μ L Assay Diluent A or 1x Assay Diluent B
	to prepare a 150 pg/mL stock standard solution. Pipette 400 μ L Assay Diluent A or 1x Assay
	Diluent B into each tube. Use the stock standard solution to produce a dilution series . Mix each
	tube thoroughly before the next transfer. Assay Diluent A or 1x Assay Diluent B serves as the
	zero standard (0 pg/mL). 200 μL 200myl 200 μL 200 μL 2 μL standard +664.7myl 200 μL 150
	50 16.67 5.56 1.85 0.62 0 pg/mL pg/mL pg/mL pg/mL pg/mL pg/mL pg/mL
	5. If the Wash Concentrate (20x) (Item B) contains visible crystals, warm to room temperature
	and mix gently until dissolved. Dilute 20 ml of Wash Buffer Concentrate into deionized or
	distilled water to yield 400 ml of 1x Wash Buffer.
	6. Briefly spin the Detection Antibody vial (Item F) before use. Add 100 μ L of 1x Assay Diluent B
	into the vial to prepare a detection antibody concentrate. Pipette up and down to mix gently (the
	concentrate can be stored at 4 $^\circ$ C for 5 days). The detection antibody concentrate should be
	diluted 65-fold with 1x Assay Diluent B and used in step 4 of Part VI Assay Procedure.
	7. Briefly spin the HRP-Streptavidin concentrate vial (Item G) and pipette up and down to mix
	gently before use. HRP-Streptavidin concentrate should be diluted 80-fold with 1x Assay Diluent
	B. For example: Briefly spin the vial (Item G) and pipette up and down to mix gently . Add 150 μL
	of HRP-Streptavidin concentrate into a tube with 12 ml 1x Assay Diluent B to prepare a final 80
	fold diluted HRP-Streptavidin solution. Mix well.
Assay Procedure:	1. Bring all reagents and samples to room temperature (18 - 25 °C) before use. It is
	recommended that all standards and samples be run at least in duplicate.
	2. Add 100 μL of each standard (see Reagent Preparation step 2) and sample into appropriate
	wells. Cover well and incubate for 2.5 hours at room temperature or over night at 4 °C with
	gentle shaking.
	3. Discard the solution and wash 4 times with 1x Wash Solution. Wash by filling each well with
	Wash Buffer (300 myl) using a multi-channel Pipette or autowasher. Complete removal of liquid
	at each step is essential to good performance. After the last wash, remove any remaining Wash
	at each step is essential to good performance. After the last wash, remove any remaining Wash Buffer by aspirating or decanting. Invert the plate and blot it against clean paper towels.
	Buffer by aspirating or decanting. Invert the plate and blot it against clean paper towels.
	Buffer by aspirating or decanting. Invert the plate and blot it against clean paper towels.4. Add 100 μL of 1x prepared biotinylated antibody (Reagent Preparation step 6) to each well.
	 Buffer by aspirating or decanting. Invert the plate and blot it against clean paper towels. 4. Add 100 μL of 1x prepared biotinylated antibody (Reagent Preparation step 6) to each well. Incubate for 1 hour at room temperature with gentle shaking.
	 Buffer by aspirating or decanting. Invert the plate and blot it against clean paper towels. 4. Add 100 µL of 1x prepared biotinylated antibody (Reagent Preparation step 6) to each well. Incubate for 1 hour at room temperature with gentle shaking. 5. Discard the solution. Repeat the wash as in step
	 Buffer by aspirating or decanting. Invert the plate and blot it against clean paper towels. 4. Add 100 µL of 1x prepared biotinylated antibody (Reagent Preparation step 6) to each well. Incubate for 1 hour at room temperature with gentle shaking. 5. Discard the solution. Repeat the wash as in step 6. Add 100 µL of prepared Streptavidin solution (see Reagent Preparation step 7) to each well.
	 Buffer by aspirating or decanting. Invert the plate and blot it against clean paper towels. 4. Add 100 µL of 1x prepared biotinylated antibody (Reagent Preparation step 6) to each well. Incubate for 1 hour at room temperature with gentle shaking. 5. Discard the solution. Repeat the wash as in step 6. Add 100 µL of prepared Streptavidin solution (see Reagent Preparation step 7) to each well. Incubate for 45 minutes at room temperature with gentle shaking.

	9. Add 50 μL of Stop Solution (Item I) to each well. Read at 450 nm immediately.
Calculation of Results:	Calculate the mean absorbance for each set of duplicate standards, controls and samples, and
	subtract the average zero standard optical density. Plot the standard curve on log-log graph
	paper or using Sigma plot software, with standard concentration on the x-axis and absorbance
	on the y-axis. Draw the best-fit straight line through the standard points.
	<u>Typical Data</u> : These standard curves are for demonstration only. A standard curve must be run
	with each assay. Assay Diluent A G-CSF concentration (pg/mL) 0.1 1 10 100 1000 O D =4 50 n
	m 0.01 0.1 1 10 Assay Diluent B G-CSF concentration (pg/mL) 0.1 1 10 100 1000 O D =4 50 n m
	0.01 0.1 1 10
	<u>Sensitivity:</u> The minimum detectable dose of G-CSF is typically less than 0.5 pg/mL.
	Recovery: Recovery was determined by spiking various levels of mouse G-CSF into mouse
	serum, plasma and cell culture media. Mean recoveries are as follows: Sample Type Average $\%$
	Recovery Range (%) Serum 95.53 83-104 Plasma 94.28 82-102 Cell culture media 97.52 84-
	104
	Linearity: Sample Type Serum Plasma Cell Culture Media 1:2 Average % of Expected 94 95 93
	Range (%) 83-103 84-103 82-102 1:4 Average % of Expected 97 96 94 Range (%) 85-105 84-
	104 83-102
	Reproducibility: Intra-Assay: CV<10 % Inter-Assay: CV<12 %
Assay Precision:	Intra-Assay: CV< 10 % Inter-Assay: CV< 12 %
Restrictions:	For Research Use only
Handling	
Handling Advice:	Avoid repeated freeze-thaw cycles.
Storage:	-20 °C
Storage Comment:	The entire kit may be stored at -20°C for up to 1 year from the date of shipment. Avoid repeated
	freeze-thaw cycles. The kit may be stored at 4°C for up to 6 months. For extended storage, it is
	recommended to store at -80°C.
Expiry Date:	6 months
Publications	
Product cited in:	Li, Wang, Yi, Jia, Bai, Peng, Yu, Xiong, Xing, Shan, Yang, Dong, Cong: "Succinate ester derivative
	of δ -tocopherol enhances the protective effects against 60Co γ -ray-induced hematopoietic

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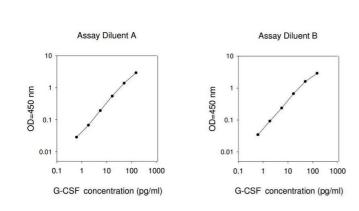
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There are more publications referencing this product on: Product page



Images

ELISA Image 1.

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