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Datasheet for ABIN625134 IL-10 ELISA Kit

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Overview

Quantity:	96 tests
Target:	IL-10 (IL10)
Reactivity:	Mouse
Method Type:	Sandwich ELISA
Detection Range:	45-5000 pg/mL
Minimum Detection Limit:	45 pg/mL
Application:	ELISA

Product Details

Purpose:	Mouse IL-10 ELISA Kit for cell culture supernatants, plasma, and serum samples.
Sample Type:	Serum, Plasma, Cell Culture Supernatant
Analytical Method:	Quantitative
Detection Method:	Colorimetric
Specificity:	This ELISA kit shows no cross-reactivity with the following cytokines tested: Mouse CD30, L
	CD30, T CD40, CRG-2, CTACK, CXCL16, Eotaxin , Eotaxin-2, Fas Ligand, Fractalkine, GCSF, GM-
	CFS, IFN- gamma, IGFBP-3, IGFBP-5, IGFBP-6, IL-1 alpha, IL-1 beta, IL-2, IL-3, IL-3 Rb, IL-4, IL-5,
	IL-9, IL-10, IL-12 p40/p70, IL-12 p70, IL-13, IL-17, KC, Leptin R, LEPTIN(OB), LIX, L-Selectin,
	Lymphotactin, MCP-1, MCP- 5, M-CSF, MIG, MIP-1 alpha, MIP-1 gamma, MIP-2, MIP-3 beta,
	MIP-3 alpha, PF-4, P-
Sensitivity:	< 45 pg/mL

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Product Details

Characteristics:	 Strip plates and additional reagents allow for use in multiple experiments Quantitative protein detection Establishes normal range The best products for confirmation of antibody array data
Components:	Pre-Coated 96-well Strip Microplate
	• Wash Buffer
	Stop Solution
	Assay Diluent(s)
	Lyophilized Standard
	Biotinylated Detection Antibody
	Streptavidin-Conjugated HRP
	TMB One-Step Substrate
Material not included:	Distilled or deionized water
	- Precision pipettes to deliver 2 μ L to 1 μ L volumes
	 Adjustable 1-25 µL pipettes for reagent preparation
	 100 µL and 1 liter graduated cylinders
	 Tubes to prepare standard and sample dilutions
	Absorbent paper
	Microplate reader capable of measuring absorbance at 450nm
	Log-log graph paper or computer and software for ELISA data analysis

Target Details

Target:	IL-10 (IL10)
Alternative Name:	IL-10 (IL10 Products)
Background:	The Mouse IL-10 (Interleukin-10) ELISA (Enzyme-Linked Immunosorbent Assay) kit is an in vitro enzyme-linked immunosorbent assay for the quantitative measurement of IL-10 in serum, plasma and cell culture supernatants. This assay employs an antibody specific for mouse IL-10 coated on a 96-well plate. Standards and samples are pipetted into the wells and IL-10 present in a sample is bound to the wells by the immobilized antibody. The wells are washed and biotinylated anti-mouse IL-10 antibody is added. After washing away unbound biotinylated antibody, HRP-conjugated streptavidin is pipetted to the wells. The wells are again washed, a TMB substrate solution is added to the wells and color develops in proportion to the amount of IL-10 bound. The Stop Solution changes the color from blue to yellow, and the intensity of the color is measured at 450 nm. Reproducibility: Intra-Assay: CV<10% Inter-Assay: CV<12%.
Gene ID:	16153

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Target Details

UniProt:	P18893
Pathways:	Cellular Response to Molecule of Bacterial Origin, Regulation of Leukocyte Mediated Immunity, Production of Molecular Mediator of Immune Response, Maintenance of Protein Location,
	Cancer Immune Checkpoints

Application Details

Application Notes:	Recommended Dilution for serum and plasma samples2 fold
Sample Volume:	100 µL
Plate:	Pre-coated
Protocol:	1. Prepare all reagents, samples and standards as instructed in the manual.
	2. Add 100 μ L of standard or sample to each well.
	3. Incubate 2.5 h at RT or O/N at 4 °C.
	4. Add 100 μ L of prepared biotin antibody to each well.
	5. Incubate 1 h at RT.
	6. Add 100 μ L of prepared Streptavidin solution to each well.
	7. Incubate 45 min at RT.
	8. Add 100 μ L of TMB One-Step Substrate Reagent to each well.
	9. Incubate 30 min at RT.
	10. Add 50 μL of Stop Solution to each well.
	11. Read at 450 nm immediately.
Reagent Preparation:	1. Bring all reagents and samples to room temperature (18 - 25 °C) before use.
	2. Sample dilution: 1x Assay Diluent D (Item K) should be used for dilution of serum/plasma
	samples. 1x Assay Diluent B (Item E) should be used for dilution of cell culture supernates.
	Suggested dilution for normal serum/plasma: 2 fold*. * Please note that levels of the target
	protein may vary between different specimens. Optimal dilution factors for each sample must
	be determined by the investigator.
	3. Assay Diluent B and Assay Diluent D should be diluted 5-fold with deionized or distilled water
	before use.
	4. Preparation of standard: Briefly spin the vial of Item C and then add 400 μ L 1x Assay Diluent
	D (for serum/plasma samples) or 1x Assay Diluent B (for cell culture supernates) into Item C
	vial to prepare a 50 ng/mL standard. Dissolve the powder thoroughly by a gentle mix. Add 60 μ L
	IL-10 standard (50 ng/mL) from the vial of Item C, into a tube with 540 μ L 1x Assay Diluent D or
	1x Assay Diluent B to prepare a 5,000 pg/mL standard solution. Pipette 300 μ L 1x Assay Diluent
	D or 1x Assay Diluent B into each tube. Use the 5,000 pg/mL standard solution to produce a
	dilution series . Mix each tube thoroughly before the next transfer. 1x Assay Diluent D or 1x

	Assay Diluent B serves as the zero standard (0 pg/mL). 5,000pg/mL standard point may be
	saturated in Assay Diluent B, we recommended starting from 2,500 pg/mL. 60 μ L standard +
	540 μL 300 μL 300 μL 300 μL 300 μL 300 μL 300 μL 300myl 5,000 2,500 1,250 625.0 312.5 156.3 78.13
	0 pg/mL pg/mL pg/mL pg/mL pg/mL pg/mL pg/mL
	5. If the Wash Concentrate (20x) (Item B) contains visible crystals, warm to room temperature
	and mix gently until dissolved. Dilute 20 ml of Wash Buffer Concentrate into deionized or
	distilled water to yield 400 ml of 1x Wash Buffer.
	6. Briefly spin the Detection Antibody vial (Item F) before use. Add 100 μL of 1x Assay Diluent B
	into the vial to prepare a detection antibody concentrate. Pipette up and down to mix gently (the
	concentrate can be stored at 4 °C for 5 days). The detection antibody concentrate should be
	diluted 80-fold with 1x Assay Diluent B and used in step 4 of Part VI Assay Procedure.
	7. Briefly spin the HRP-Streptavidin concentrate vial (Item G) and pipette up and down to mix
	gently before use. HRP-Streptavidin concentrate should be diluted 200-fold with 1x Assay
	Diluent B. For example: Briefly spin the vial (Item G) and pipette up and down to mix gently . Add
	50 μ L of HRP-Streptavidin concentrate into a tube with 10 ml 1x Assay Diluent B to prepare a
	200-fold diluted HRP- Streptavidin solution (don't store the diluted solution for next day use).
	Mix well.
Assay Procedure:	1. Bring all reagents and samples to room temperature (18 - 25 °C) before use. It is
Abbdy Procedure.	
noody noocdare.	recommended that all standards and samples be run at least in duplicate.
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	recommended that all standards and samples be run at least in duplicate. 2. Add 100 µL of each standard (see Reagent Preparation step 2) and sample into appropriate wells. Cover well and incubate for 2.5 hours at room temperature or over night at 4 °C with gentle shaking. 3. Discard the solution and wash 4 times with 1x Wash Solution. Wash by filling each well with Wash Buffer (300 myl) using a multi-channel pipette or autowasher. Complete removal of liquid at each step is essential to good performance. After the last wash, remove any remaining Wash
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	 recommended that all standards and samples be run at least in duplicate. 2. Add 100 μL of each standard (see Reagent Preparation step 2) and sample into appropriate wells. Cover well and incubate for 2.5 hours at room temperature or over night at 4 °C with gentle shaking. 3. Discard the solution and wash 4 times with 1x Wash Solution. Wash by filling each well with Wash Buffer (300 myl) using a multi-channel pipette or autowasher. Complete removal of liquid at each step is essential to good performance. After the last wash, remove any remaining Wash Buffer by aspirating or decanting. Invert the plate and blot it against clean paper towels. 4. Add 100 μL of 1x prepared biotinylated antibody (Reagent Preparation step 6) to each well. Incubate for 1 hour at room temperature with gentle shaking.
	 recommended that all standards and samples be run at least in duplicate. 2. Add 100 μL of each standard (see Reagent Preparation step 2) and sample into appropriate wells. Cover well and incubate for 2.5 hours at room temperature or over night at 4 °C with gentle shaking. 3. Discard the solution and wash 4 times with 1x Wash Solution. Wash by filling each well with Wash Buffer (300 myl) using a multi-channel pipette or autowasher. Complete removal of liquid at each step is essential to good performance. After the last wash, remove any remaining Wash Buffer by aspirating or decanting. Invert the plate and blot it against clean paper towels. 4. Add 100 μL of 1x prepared biotinylated antibody (Reagent Preparation step 6) to each well. Incubate for 1 hour at room temperature with gentle shaking. 5. Discard the solution. Repeat the wash as in step
	 recommended that all standards and samples be run at least in duplicate. 2. Add 100 μL of each standard (see Reagent Preparation step 2) and sample into appropriate wells. Cover well and incubate for 2.5 hours at room temperature or over night at 4 °C with gentle shaking. 3. Discard the solution and wash 4 times with 1x Wash Solution. Wash by filling each well with Wash Buffer (300 myl) using a multi-channel pipette or autowasher. Complete removal of liquid at each step is essential to good performance. After the last wash, remove any remaining Wash Buffer by aspirating or decanting. Invert the plate and blot it against clean paper towels. 4. Add 100 μL of 1x prepared biotinylated antibody (Reagent Preparation step 6) to each well. Incubate for 1 hour at room temperature with gentle shaking. 5. Discard the solution. Repeat the wash as in step 6. Add 100 μL of prepared Streptavidin solution (see Reagent Preparation step 7) to each well.
	 recommended that all standards and samples be run at least in duplicate. 2. Add 100 μL of each standard (see Reagent Preparation step 2) and sample into appropriate wells. Cover well and incubate for 2.5 hours at room temperature or over night at 4 °C with gentle shaking. 3. Discard the solution and wash 4 times with 1x Wash Solution. Wash by filling each well with Wash Buffer (300 myl) using a multi-channel pipette or autowasher. Complete removal of liquid at each step is essential to good performance. After the last wash, remove any remaining Wash Buffer by aspirating or decanting. Invert the plate and blot it against clean paper towels. 4. Add 100 μL of 1x prepared biotinylated antibody (Reagent Preparation step 6) to each well. Incubate for 1 hour at room temperature with gentle shaking. 5. Discard the solution. Repeat the wash as in step 6. Add 100 μL of prepared Streptavidin solution (see Reagent Preparation step 7) to each well. Incubate for 45 minutes at room temperature with gentle shaking.
	 recommended that all standards and samples be run at least in duplicate. 2. Add 100 μL of each standard (see Reagent Preparation step 2) and sample into appropriate wells. Cover well and incubate for 2.5 hours at room temperature or over night at 4 °C with gentle shaking. 3. Discard the solution and wash 4 times with 1x Wash Solution. Wash by filling each well with Wash Buffer (300 myl) using a multi-channel pipette or autowasher. Complete removal of liquid at each step is essential to good performance. After the last wash, remove any remaining Wash Buffer by aspirating or decanting. Invert the plate and blot it against clean paper towels. 4. Add 100 μL of 1x prepared biotinylated antibody (Reagent Preparation step 6) to each well. Incubate for 1 hour at room temperature with gentle shaking. 5. Discard the solution. Repeat the wash as in step 6. Add 100 μL of prepared Streptavidin solution (see Reagent Preparation step 7) to each well. Incubate for 45 minutes at room temperature with gentle shaking. 7. Discard the solution. Repeat the wash as in step

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Application Details

Calculation of Results:	Calculate the mean absorbance for each set of duplicate standards, controls and samples, and
	subtract the average zero standard optical density. Plot the standard curve on log-log graph
	paper or using Sigma plot software, with standard concentration on the x-axis and absorbance
	on the y-axis. Draw the best-fit straight line through the standard points.
	Typical Data: These standard curves are for demonstration only. A standard curve must be run
	with each assay. Assay Diluent D Mouse IL-10 concentration (pg/mL) 10 100 1000 10000 O D
	=4 50 n m 0.01 0.1 1 10 Assay Diluent B Mouse IL-10 concentration (pg/mL) 10 100 1000
	10000 O D =4 50 n m 0.01 0.1 1 10
	<u>Sensitivity:</u> The minimum detectable dose of IL-10 is typically less than 45 pg/mL.
	Recovery: Recovery was determined by spiking various levels of mouse IL-10 into mouse
	serum, plasma and cell culture media. Mean recoveries are as follows: Sample Type Average $\%$
	Recovery Range (%) Serum 85.97 77-93 Plasma 92.20 71-134 Cell culture media 103.7 79-120
	D. REPRODUCIBILITY Intra-Assay: CV<10 % Inter-Assay: CV<12 %
Assay Precision:	Intra-Assay: CV< 10 % Inter-Assay: CV< 12 %
Restrictions:	For Research Use only

Handling

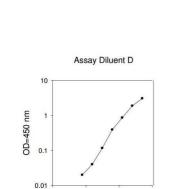
Handling Advice:	Avoid repeated freeze-thaw cycles.
Storage:	-20 °C
Storage Comment:	The entire kit may be stored at -20°C for up to 1 year from the date of shipment. Avoid repeated freeze-thaw cycles. The kit may be stored at 4°C for up to 6 months. For extended storage, it is recommended to store at -80°C.
Expiry Date:	6 months
Publications	
Product cited in:	Bil-Lula, Krzywonos-Zawadzka, Sawicki, Woźniak: "An infection of human adenovirus 31 affects the differentiation of preadipocytes into fat cells, its metabolic profile and fat accumulation." in: Journal of medical virology , Vol. 88, Issue 3, pp. 400-7, (2016) (PubMed).
	Nandi, Bishayi: "Murine macrophage response from peritoneal cavity requires signals mediated by chemokine receptor CCR-2 during Staphylococcus aureus infection." in: Immunologic research , Vol. 64, Issue 1, pp. 213-32, (2016) (PubMed).

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Dey, Majhi, Mahanti, Dey, Bishayi: "In Vitro Anti-inflammatory and Immunomodulatory Effects of Ciprofloxacin or Azithromycin in Staphylococcus aureus-Stimulated Murine Macrophages are Beneficial in the Presence of Cytochalasin D." in: **Inflammation**, Vol. 38, Issue 3, pp. 1050-69, (2015) (PubMed).

Ahmad, Attia, Bakheet, Zoheir, Ansari, Korashy, Abdel-Hamied, Ashour, Abd-Allah et al.: "Naringin attenuates the development of carrageenan-induced acute lung inflammation through inhibition of NF-?b, STAT3 and pro-inflammatory mediators and enhancement of I?B? and anti-inflammatory ..." in: **Inflammation**, Vol. 38, Issue 2, pp. 846-57, (2015) (PubMed).

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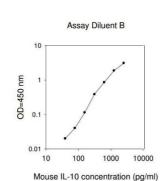
100

1000

Mouse IL-10 concentration (pg/ml)

10

10000



ELISA Image 1.

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