

Datasheet for ABIN625160
CCL2 ELISA Kit



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Overview

Quantity:	96 tests
Target:	CCL2
Reactivity:	Mouse
Method Type:	Sandwich ELISA
Application:	ELISA

Product Details

Purpose:	Mouse MCP-1 (CCL2) ELISA Kit for cell and tissue lysate samples.
Sample Type:	Tissue Lysate, Cell Lysate
Analytical Method:	Quantitative
Detection Method:	Colorimetric
Specificity:	The antibody pair provided in this kit recognizes mouse MCP-1 / CCL2.
Sensitivity:	3 pg/mL
Characteristics:	• Strip plates and additional reagents allow for use in multiple experiments • Quantitative protein detection • Establishes normal range • The best products for confirmation of antibody array data
Components:	• Pre-Coated 96-well Strip Microplate • Wash Buffer • Stop Solution • Assay Diluent(s) • Lyophilized Standard

Product Details

- Biotinylated Detection Antibody
- Streptavidin-Conjugated HRP
- TMB One-Step Substrate

Material not included:	• Distilled or deionized water • Precision pipettes to deliver 2 µL to 1 µL volumes • Adjustable 1-25 µL pipettes for reagent preparation • 100 µL and 1 liter graduated cylinders • Tubes to prepare standard and sample dilutions • Absorbent paper • Microplate reader capable of measuring absorbance at 450nm • Log-log graph paper or computer and software for ELISA data analysis • Cell lysate buffer
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Target Details

Target:	CCL2
Alternative Name:	MCP-1 / CCL2 (CCL2 Products)
Background:	Monocyte chemoattractant protein-1 (MCP-1), also called monocyte chemotactic protein-1, belongs to the family of Chemokines. MCP-1 is expressed by monocytes, vascular endothelial cells, smooth muscle cells, glomerular mesangial cell, and osteoblastic cells. It has been shown to exhibit biological activities other than Chemotaxis. The activity of mouse MCP-1 has been shown to be mediated by the mouse CC chemokine receptor CCR2. The Mouse MCP-1 ELISA (Enzyme-Linked Immunosorbent Assay) kit is an in vitro enzyme-linked immunosorbent assay for the quantitative measurement of mouse MCP-1 cell lysate and tissue lysate. This assay employs an antibody specific for mouse MCP-1 coated on a 96-well plate. Standards and samples are pipetted into the wells and MCP-1 present in a sample is bound to the wells by the immobilized antibody. The wells are washed and biotinylated anti-mouse MCP-1 antibody is added. After washing away unbound biotinylated antibody, HRP-conjugated streptavidin is pipetted to the wells. The wells are again washed, a TMB substrate solution is added to the wells and color develops in proportion to the amount of MCP-1 bound. The Stop Solution changes the color from blue to yellow, and the intensity of the color is measured at 450 nm. Reproducibility: Intra-Assay: CV<10% Inter-Assay: CV<12%.
Gene ID:	20296
UniProt:	P10148
Pathways:	Cellular Response to Molecule of Bacterial Origin, Positive Regulation of Immune Effector

Target Details

Process, ER-Nucleus Signaling, Unfolded Protein Response, The Global Phosphorylation Landscape of SARS-CoV-2 Infection

Application Details

Sample Volume:	100 µL																					
Plate:	Pre-coated																					
Protocol:	1. Prepare all reagents, samples and standards as instructed in the manual. 2. Add 100 µL of standard or sample to each well. 3. Incubate 2.5 h at RT or O/N at 4 °C. 4. Add 100 µL of prepared biotin antibody to each well. 5. Incubate 1 h at RT. 6. Add 100 µL of prepared Streptavidin solution to each well. 7. Incubate 45 min at RT. 8. Add 100 µL of TMB One-Step Substrate Reagent to each well. 9. Incubate 30 min at RT. 10. Add 50 µL of Stop Solution to each well. 11. Read at 450 nm immediately.																					
Reagent Preparation:	1. Bring all reagents and samples to room temperature (18 - 25 °C) before use. 2. Sample dilution: Tissue lysate and cell lysate sample should be diluted at least 5-folds with 1x Sample Diluent Buffer. 3. Sample Diluent Buffer (Item D) and Assay Diluent (Item E) should be diluted 5-folds with deionized or distilled water before use. 4. Preparation of standard: Briefly spin the vial of Item C. Add 400 µL 1x Sample Diluent Buffer (Item D, Sample Diluent Buffer should be diluted 5-fold with deionized or distilled water before use) into Item C vial to prepare a 50 ng/mL standard. Dissolve the powder thoroughly by a gentle mix. Add 40 µL MCP-1 standard from the vial of Item C, into a tube with 960 µL Sample Diluent Buffer to prepare a 2,000 pg/mL stock standard solution. Pipette 400 µL 1x Sample Diluent Buffer into each tube. Use the stock standard solution to produce a dilution series . Mix each tube thoroughly before the next transfer. 1x Sample Diluent Buffer serves as the zero standard (0 pg/mL).<table><tr><td>200 µL</td><td>200 µL</td><td>200 µL</td><td>200µL</td><td>200 µL</td><td>200 µL</td><td>40 µL standard + 960 µL</td></tr><tr><td>2,000</td><td>666.7</td><td>222.2</td><td>74.07</td><td>24.69</td><td>8.23</td><td>2.74</td></tr><tr><td>0 pg/mL</td><td>pg/mL</td><td>pg/mL</td><td>pg/mL</td><td>pg/mL</td><td>pg/mL</td><td>pg/mL</td></tr></table> 5. If the Wash Concentrate (20x) (Item B) contains visible crystals, warm to room temperature and mix gently until dissolved. Dilute 20 ml of Wash Buffer Concentrate into deionized or distilled water to yield 400 ml of 1x Wash Buffer. 6. Briefly spin the Detection Antibody vial (Item F) before use. Add 100 µL of 1x Assay Diuent	200 µL	200 µL	200 µL	200µL	200 µL	200 µL	40 µL standard + 960 µL	2,000	666.7	222.2	74.07	24.69	8.23	2.74	0 pg/mL	pg/mL	pg/mL	pg/mL	pg/mL	pg/mL	pg/mL
200 µL	200 µL	200 µL	200µL	200 µL	200 µL	40 µL standard + 960 µL																
2,000	666.7	222.2	74.07	24.69	8.23	2.74																
0 pg/mL	pg/mL	pg/mL	pg/mL	pg/mL	pg/mL	pg/mL																

into the vial to prepare a detection antibody concentrate. Pipette up and down to mix gently (the concentrate can be stored at 4 °C for 5 days). The detection antibody concentrate should be diluted 80-folds with 1x Assay Diluent and used in step 4 of Part VI Assay Procedure.

7. Briefly spin the HRP-Streptavidin concentrate vial (Item G) before use. HRP-Streptavidin concentrate should be diluted 400-folds with 1x Assay Diluent. For example: Add 30 µL of HRP-Streptavidin concentrate into a tube with 12 ml 1x Assay Diluent to prepare a 400-fold diluted HRP-Streptavidin solution (don't store the diluted solution for next day use). Mix well.

8. Cell lysate buffer should be diluted 2-fold with deionized or distilled water (for cell lysate and tissue lysate).

Assay Procedure:

1. Bring all reagents and samples to room temperature (18 - 25 °C) before use. It is recommended that all standards and samples be run at least in duplicate.
2. Add 100 µL of each standard (see Reagent Preparation step 2) and sample into appropriate wells. Cover well and incubate for 2.5 hours at room temperature or over night at 4 °C with gentle shaking.
3. Discard the solution and wash 4 times with 1x Wash Solution. Wash by filling each well with Wash Buffer (300 µl) using a multi-channel Pipette or autowasher. Complete removal of liquid at each step is essential to good performance. After the last wash, remove any remaining Wash Buffer by aspirating or decanting. Invert the plate and blot it against clean paper towels.
4. Add 100 µL of 1x prepared biotinylated antibody (Reagent Preparation step 6) to each well. Incubate for 1 hour at room temperature with gentle shaking.
5. Discard the solution. Repeat the wash as in step
6. Add 100 µL of prepared Streptavidin solution (see Reagent Preparation step 7) to each well. Incubate for 45 minutes at room temperature with gentle shaking.
7. Discard the solution. Repeat the wash as in step
8. Add 100 µL of TMB One-Step Substrate Reagent (Item H) to each well. Incubate for 30 minutes at room temperature in the dark with gentle shaking.
9. Add 50 µL of Stop Solution (Item I) to each well. Read at 450 nm immediately.

Calculation of Results:

Calculate the mean absorbance for each set of duplicate standards, controls and samples, and subtract the average zero standard optical density. Plot the standard curve on log-log graph paper or using Sigma plot software, with standard concentration on the x-axis and absorbance on the y-axis. Draw the best-fit straight line through the standard points.

Typical Data: These standard curves are for demonstration only. A standard curve must be run with each assay. Assay Diluent Mouse MCP-1 concentration (pg/mL) 1 10 100 1000 10000 O D =4 50 (n m) 0.01 0.1 1 10

Sensitivity: The minimum detectable dose of MCP-1 is typically less than 3 pg/mL.

Application Details

Recovery: Recovery was determined by spiking various levels of mouse MCP-1 into mouse tissue lysate and cell lysate. Mean recoveries are as follows: Sample Type Average % Recovery Range (%) Tissue lysate 85.89 78-101 Cell lysate 87.56 80-102

Linearity: Sample Type Tissue Cell Lysate lysate 1:2 Average % of 87 88 Expected Range (%) 77-99 78-101 1:4 Average % of 89 94 Expected Range (%) 78-103 82-104

Reproducibility: Intra-Assay: CV<10 % Inter-Assay: CV<12 %

Assay Precision: Intra-Assay: CV< 10 % Inter-Assay: CV< 12 %

Restrictions: For Research Use only

Handling

Handling Advice: Avoid repeated freeze-thaw cycles.

Storage: -20 °C

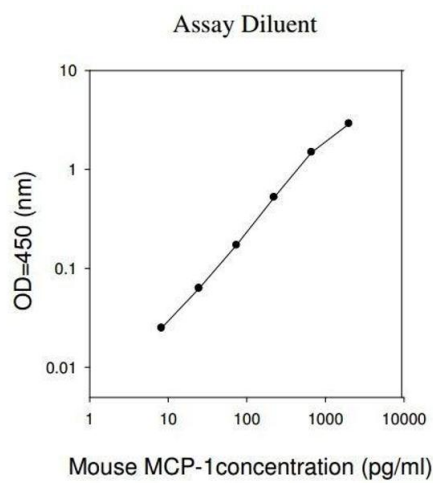
Storage Comment: The entire kit may be stored at -20°C for up to 1 year from the date of shipment. Avoid repeated freeze-thaw cycles. The kit may be stored at 4°C for up to 6 months. For extended storage, it is recommended to store at -80°C.

Expiry Date: 6 months

Publications

Product cited in: Williams, Jiang, Cochran, Dorsam, Graves: "Regulated expression of monocyte chemoattractant protein-1 in normal human osteoblastic cells." in: **The American journal of physiology**, Vol. 263, Issue 1 Pt 1, pp. C194-9, (1992) ([PubMed](#)).

Beall, Mahajan, Kolattukudy: "Conversion of monocyte chemoattractant protein-1 into a neutrophil attractant by substitution of two amino acids." in: **The Journal of biological chemistry**, Vol. 267, Issue 5, pp. 3455-9, (1992) ([PubMed](#)).



ELISA

Image 1.