Datasheet for ABIN625669

Mouse Inflammation Array G1

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Overview

Quantity:	4 samples
Reactivity:	Mouse
Method Type:	Sandwich ELISA
Application:	Antibody Array (AA)

Product Details

Purpose:	G-Series Mouse Inflammation Antibody Array 1 Kit. Detects 40 Mouse Inflammatory Factors. Suitable for all liquid sample types.
Brand:	RayBio®
Sample Type:	Serum, Plasma, Cell Culture Supernatant, Cell Lysate, Tissue Lysate
Analytical Method:	Semi-Quantitative
Detection Method:	Fluorometric
Specificity:	BLC (CXCL13), CD30 Ligand (TNFSF8), Eotaxin-1 (CCL11), Eotaxin-2 (MPIF-2/CCL24), Fas Ligand (TNFSF6), Fractalkine (CX3CL1), GCSF, GM-CSF, IFN-gamma, IL-1 alpha (IL-1 F1), IL-1 beta (IL-1 F2), IL-2, IL-3, IL-4, IL-6, IL-9, IL-10, IL-12 p40/p70, IL-12 p70, IL-13, IL-17A, I-TAC (CXCL11), KC (CXCL1), Leptin, LIX, Lymphotactin (XCL1), MCP-1 (CCL2), M-CSF, MIG (CXCL9), MIP-1 alpha (CCL3), MIP-1 gamma, RANTES (CCL5), SDF-1 alpha (CXCL12 alpha), I-309 (TCA-3/CCL1), TECK (CCL25), TIMP-1, TIMP-2, TNF alpha, TNF RI (TNFRSF1A), TNF RII (TNFRSF1B)
Characteristics:	• High sensitivity and specificity • Low sample volume (10-100 µL per array) • Large dynamic range of detection • Compatible with most sample types

Product Details

	• Test 4 or 8 samples on each slide • Suitable for high-throughput assays
Components:	Cytokine Antibody Array glass slide (4 or 8 arrays per slide) Biotinylated Detection Antibodies Streptavidin-conjugated HiLytePlus™ 555 Fluor Blocking Buffer 20X Wash Buffer I 20X Wash Buffer II 2X Cell Lysis Buffer G-Series Antibody Array accessories Accessories include: 16-well incubation chamber with gasket, protective cover, snap-on sides, adhesive film
Material not included:	Small plastic boxes or containers Pipettors, pipette tips and other common lab consumables Orbital shaker or oscillating rocker Aluminum foil Gene microarray scanner or similar laser fluorescence scanner

Application Details

Application Notes:	Completely cover array area with sample or buffer during incubation. Avoid foaming during incubation steps. Perform all incubation and wash steps under gentle rocking or rotation. Cover the incubation chamber with adhesive film during incubation, particularly when incubation is more than 2 hours or <70 µL of sample or reagent is used. Several incubation steps such as step 6 (blocking), step 7 (sample incubation), step 10 (detection antibody incubation), or step 13 (Cy3 equivalent dyestreptavidin incubation) may be done overnight at 4 °C. Please make sure to cover the incubation chamber tightly to prevent evaporation.
Comment:	The G-Series arrays feature fluorescent signal detection. The antibodies are spotted on glass slide solid supports and require a laser scanner for data collection. All G-Series arrays work on the sandwich ELISA principle, utilizing a matched pair of antibodies: an immobilized capture antibody and a corresponding biotinylated detection antibody.
Sample Volume:	100 µL
Assay Time:	6 h
Plate:	Glass Slide

Application Details

Protocol:	1. Dry the glass slide 2. Block array surface 3. Incubate with Sample 4. Incubate with Biotinylated Detection Antibody Cocktail 5. Incubate with Streptavidin-Conjugated Fluor 6. Disassemble the glass slide 7. Scan with a gene microarray laser scanner 8. Perform densitometry and analysis
Sample Preparation:	Use serum-free conditioned media if possible. If serum-containing conditioned media is required, it is highly recommended that complete medium be used as a control since many types of sera contains cytokines. We recommend the following parameters for your samples: 50 to 100 µl of original or diluted serum, plasma, cell culture media, or other body fluid, or 50-500 µg/ml of protein for cell and tissue lysates. If you experience high background or if the fluorescent signal intensities exceed the detection range, further dilution of your sample is recommended.
Assay Procedure:	Take out the glass slide from the box, and let it equilibrate to room temperature inside the sealed plastic bag for 20-30 minutes. Remove slide from the plastic bag, peel off the cover film, and let it air dry for another 1-2 hours. Blocking & Incubation 1. Add 100 µl Sample Diluent into each well and incubate at room temperature for 30 minutes to block slides. 2. Decant buffer from each well. Add 100 µl of sample to each well. Incubate arrays at room temperature for 1-2 hour. 3. Decant the samples from each well, and wash 5 times (5 min each) with 150 µl of 1X Wash Buffer I at room temperature with gentle shaking. Completely remove wash buffer in each wash step. Dilute 20x Wash Buffer I with H2O. 4. Decant the 1x Wash Buffer I from each well, wash 2 times (5 min each) with 150 µl of 1X Wash Buffer II at room temperature with gentle shaking.Completely remove wash buffer in each wash step. Dilute 20X Wash Buffer II with H2O. Incubation with Biotinylated Antibody Cocktail & Wash 5. Reconstitute the detection antibody by adding 1.4 ml of Sample Diluent to the tube. Spin briefly. 6. Add 80 µl of the detection antibody cocktail to each well. Incubate at room temperature for 1-2 hour.

Application Details

	7. Decant the samples from each well, and wash 5 times (5 mins each) with 150 µl of 1X Wash Buffer I and then 2 times with 150 µl of 1x Wash Buffer II at room temperature with gentle shaking. Completely remove wash buffer in each wash step. Incubation with Cy3 Equivalent Dye-Streptavidin & Wash 8. After briefly spinning down, add 1.4 ml of Sample Diluent to Cy3 equivalent dye-conjugated streptavidin tube. Mix gently. 9. Add 80 µl of Cy3 equivalent dye-conjugated streptavidin to each well. Cover the device with aluminum foil to avoid exposure to light or incubate in dark room. Incubate at room temperature for 1 hour. 10. Decant the samples from each well, and wash 5 times (5 mins each) with 150 µl of 1X Wash Buffer I at room temperature with gentle shaking. Completely remove wash buffer in each wash step. Fluorescence Detection 11. Disassemble the device by pushing clips outward from the slide side. Carefully remove the slide from the gasket. 12. Place the slide in the Slide Washer/Dryer (a 4-slide holder/centrifuge tube), add enough 1x Wash Buffer I (about 30 ml) to cover the whole slide, and then gently shake at room temperature for 15 minutes. Decant Wash Buffer I. Wash with 1x Wash Buffer II (about 30 ml) and gently shake at room temperature for 5 minutes. 13. Remove water droplets completely by gently applying suction with a pipette to remove water droplets. Do not touch the array, only the sides. 14. Imaging: The signals can be visualized through use of a laser scanner equipped with a Cy3 wavelength (green channel) such as Axon GenePix.
Calculation of Results:	Data extraction can be done using the GAL file that is specific for this array along with the microarray analysis software (GenePix, ScanArray Express, ArrayVision, MicroVigene, etc.).
Restrictions:	For Research Use only

Handling

Handling Advice:	Do not touch the surface of the slides, as the microarray slides are very sensitive. Hold the slides by the edges only. Handle all buffers and slides with powder free gloves. Handle glass slide/s in clean environment. The G-Series slides do not have bar codes. To help distinguish one slide from another, transcribe the slide serial number from the slide bag to the back of the slide with a fine point permanent marker. Please write the number on the very bottom edge of the

Handling

	slide, taking care to avoid writing on the array well areas.
Storage:	-20 °C
Storage Comment:	For best results, store the entire kit frozen at -20°C upon arrival. Stored frozen, the kit will be stable for at least 6 months which is the duration of the product warranty period. Once thawed, store array slide(s) and 1X Blocking Buffer at -20°C and all other reagents undiluted at 4°C for no more than 3 months.
Expiry Date:	6 months

Publications

Product cited in:

Biwer, Dsouza, Abidali, Tu, Siniard, DeBoth, Huentelman, Hale: "Time course of cardiac inflammation during nitric oxide synthase inhibition in SHR: impact of prior transient ACE inhibition." in: **Hypertension research : official journal of the Japanese Society of Hypertension**, Vol. 39, Issue 1, pp. 8-18, (2016) (PubMed).

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