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anti-Fibrinogen antibody





Image



Publications



Go to Product page

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Quantity:	100 μL
Target:	Fibrinogen
Reactivity:	Human, Rat
Host:	Rabbit
Clonality:	Polyclonal
Conjugate:	This Fibrinogen antibody is un-conjugated
Application:	ELISA, Immunohistochemistry (Paraffin-embedded Sections) (IHC (p)), Immunofluorescence (Paraffin-embedded Sections) (IF (p)), Immunohistochemistry (Frozen Sections) (IHC (fro)), Immunofluorescence (Cultured Cells) (IF (cc))

Product Details

Immunogen:	Fibrinogen from human plasma
Isotype:	IgG
Cross-Reactivity:	Human, Rat
Predicted Reactivity:	Mouse
Purification:	Purified by Protein A.

Target Details

Target:	Fibrinogen
Abstract:	Fibrinogen Products

Target Details

Precaution of Use:

Storage:

rarget Details	
Background:	Synonyms: Fib2, Fibrinogen alpha chain, FGA
	Background: Cleaved by the protease thrombin to yield monomers which, together with
	fibrinogen beta (FGB) and fibrinogen gamma (FGG), polymerize to form an insoluble fibrin
	matrix. Fibrin has a major function in hemostasis as one of the primary components of blood
	clots. In addition, functions during the early stages of wound repair to stabilize the lesion and
	guide cell migration during re-epithelialization. Was originally thought to be essential for platelet
	aggregation, based on in vitro studies using anticoagulated blood. However, subsequent
	studies have shown that it is not absolutely required for thrombus formation in vivo. Enhances
	expression of SELP in activated platelets via an ITGB3-dependent pathway. Maternal fibrinogen
	is essential for successful pregnancy. Fibrin deposition is also associated with infection, where
	it protects against IFNG-mediated hemorrhage. May also facilitate the immune response via
	both innate and T-cell mediated pathways.
Gene ID:	2243
UniProt:	P02671
Application Details	
Application Notes:	ELISA 1:500-1000
	IHC-P 1:200-400
	IHC-F 1:100-500
	IF(IHC-P) 1:50-200
	IF(IHC-F) 1:50-200
	IF(ICC) 1:50-200
Restrictions:	For Research Use only
Handling	
Format:	Liquid
Concentration:	1 μg/μL
Buffer:	0.01M TBS(pH 7.4) with 1 % BSA, 0.02 % Proclin300 and 50 % Glycerol.
Preservative:	ProClin
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handled by trained staff only.

4 °C,-20 °C

This product contains ProClin: a POISONOUS AND HAZARDOUS SUBSTANCE, which should be

Handling

Storage Comment:	Shipped at 4°C. Store at -20°C for one year. Avoid repeated freeze/thaw cycles.
Expiry Date:	12 months

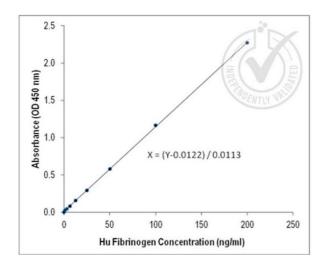
Publications

Product cited in:

Chen, Yang, Liao, Wang, Chen, Sun, Huang: "Effect of the duration of UV irradiation on the anticoagulant properties of titanium dioxide films." in: **ACS applied materials & interfaces**, Vol. 7, Issue 7, pp. 4423-32, (2015) (PubMed).

Chen, Zhao, Chen, Liao, Yang, Sun, Huang: "The effect of full/partial UV-irradiation of TiO2 films on altering the behavior of fibrinogen and platelets." in: **Colloids and surfaces. B, Biointerfaces**, Vol. 122, pp. 709-18, (2014) (PubMed).

Validation report #028531 for ELISA (ELISA)



ELISA

Image 1. Antigen: human liver Primary:Rabbit Anti-human Fibrinogen Polyclonal Antibody, 1:500 Secondary: Goat Anti-Rabbit IgG Antibody HRP Conjugated, at 1:10,000, TMB staining, Read the data in MicroplateReader at 450 nm.





Successfully validated (ELISA (ELISA))

by Alamo Laboratories

Report Number: 028531

Date: Jul 16 2013

1 x PBS per well.

per well and incubated at 37°C for 2 h.

Lot Number:	980680
Method validated:	ELISA (ELISA)
Positive Control:	Liver
Negative Control:	Skeletal muscle
Notes:	Signal was detected in positive control samples but not in negative control samples.
Primary Antibody:	- Antibody: Fibrinogen antibody - Catalog number: ABIN673854 - Batch number: 980680
Controls:	 Positive control: protein extract from human liver (specimen known to contain the target protein) was from Alamo Laboratories, Inc (Cat # 2001-TEL-Hu). Negative control: protein extract from mouse skeletal muscle (specimen known to not contain the target protein) was from Alamo Laboratories, Inc (Cat # 2002-TEM-Mm). Standard curve: serial two-fold dilutions from 200 ng/ml [200, 100, 50, 25, 12.5, 6.25, 3.125, 1.5625, 0.78125, 0] were generated from Fibrinogen peptide stock diluted in 1 x 50 mM Carbonate buffer, pH 9.5. Spike control: standard diluted in protein lysate buffer [25 and 0].
Protocol:	 A 96-well MICROLON® 600 96W High Binding 12 x 8 Clear Strip Microplate was coated with antigen by pipetting 100 μL of sample per well. All samples and standards were assayed in triplicate. The microplate was covered and incubated at 25°C overnight. After overnight coating, plate contents were discarded and wells were washed 3 times with 150 μL of 1 x PBST (PBS, 0.1% Tween-20) per well. Wells were then blocked with 250 μL of 1 x PBS / 0.05% Tween-20 / 1% BSA per well and incubated at 37°C for 2 h. After blocking, plate contents were discarded and wells were washed 3 times with 250 μL of

1% BSA was added per well and incubated at 37°C for 1 h.

• 100 μ L of primary antibody diluted 1:500 in 1 x PBS / 0.05% Tween-20 / 1% BSA was added

• Plate contents were discarded and wells were washed 3 times with 200 µL of 1 x PBST per

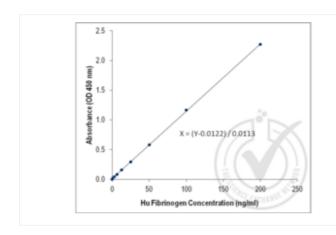
• 100 µL of HRP conjugated secondary antibody diluted 1:10000 in 1 x PBS / 0.05% Tween-20 /

- Plate contents were discarded and wells were washed 3 times with 200 µL of 1 x PBST per
- 100 µL of TMB (3,3', 5,5"-tetramethylbenzidine) substrate was added per well and incubated at 37°C for 10 min.
- 100 µL of STOP solution was added per well.
- The optical density (OD value) of each well was read using a microplate reader set to 450 nm.
- The triplicate readings for each sample were averaged and the average zero standard optical density subtracted. The corrected average-value was tabulated as Average Absorbance. A standard curve was generated by plotting the mean OD value for each standard on the y-axis against the concentration on the x-axis using Excel. A line of best fit through the points on the graph was used to generate the equation X = (Y-0.0122) / 0.0113.
- The equation X = (Y-0.0122) / 0.0113 was used to calculate fibrinogen concentrations of the samples based on their Average Absorbance values.

Experimental Notes:

None

Images for Validation report #028531



Validation image no. 1 for anti-Fibrinogen antibody (ABIN673854)

Figure 1: Graph of corrected-average absorbance (OD 450 nm) readings plotted for standard curve samples.

Туре	Sample ng/ml	Reading- 1	Reading- 2	Reading- 3	Avg Reading	Avg Absorbance	SD	Calculated Conc
Standard Curve	200 ng/mil	2.3472	2.3882	2.3907	2.3754	2.2700	0.0244	199.808
	100 ng/ml	1.2662	1.2688	1.2730	1.2693	1,1640	0.0034	101.932
	50 ng/ml	0.6780	0.7050	0.6680	0.6837	0.5784	0.0191	50.1032
	25 ng/ml	0.3920	0.4180	0.3830	0.3977	0.2924	0.0182	24.7935
	12.5 ng/ml	0.3310	0.2650	0.2020	0.2660	0.1607	0.0645	13:1416
	6.25 ng/ml	0.1848	0.1970	0.1850	0.1889	0.0836	0.0070	6.3215
	3.125 ng/ml	0.1510	0.1490	0.1430	0.1477	0.0424	0.0042	2.6696
	1.5625 ng/ml	0.1350	0.1384	0.1368	0.1367	0.0314	0.0017	1.7021
	0.7813 ng/ml	0.1288	0.1286	0.1298	0:1291	0.0238	0.0006	1.0236
	0 ng/ml	0.1030	0.0970	0.1160	0.1053	0.0000	0.0097	-1.0767
Calles Controls	25 ng/ml	0.4010	0.3980	0.4110	0.4033	0.2980	0.0068	25,299
Spike Controls	0 ng/ml	0.0980	0.1010	0.1223	0.1071	Vo., 0.0018	0.0132	40.9204
Positive Control	Liver Extract	1.9970	1.9240	2.1240	2.0150	1.9097	0.1012	167.920
Negative Control	Muscle	0.1210	0.1060	0.0988	0.1086	0.0033	0.0113	-0.787

Validation image no. 2 for anti-Fibrinogen antibody (ABIN673854)

Table 1: ELISA. Fibrinogen is present in the positive control sample (liver) and absent from the negative control (skeletal muscle) sample. Spike controls indicate no interference in absorbance readings from the protein lysate buffer used to prepare the positive and negative control samples. Absorbance readings (OD 450 nm) are shown for standard curve, spike controls and unknown positive (liver extract) and negative (skeletal muscle extract) control samples.

Value for Average Reading is derived from the average of three readings (OD 450nm). The Average Reading for 0 ng/ml Standard was subtracted from all Average Readings to yield Average Absorbance values for Standards, spike controls and unknown positive (liver extract) and negative (skeletal muscle extract) control samples. Standard deviation is included for all samples. An equation (X = (Y - 0.0122) / 0.0113) was generated from the standard curve and used to calculate fibrinogen concentrations shown in the Table.