# .-online.com antibodies

Datasheet for ABIN6957217 ISL1 ELISA Kit

Image



#### Overview

Quantity:	96 tests
Target:	ISL1
Reactivity:	Human
Method Type:	Sandwich ELISA
Detection Range:	0.15 ng/mL - 10 ng/mL
Minimum Detection Limit:	0.15 ng/mL
Application:	ELISA

## Product Details

The kit is a sandwich enzyme immunoassay for in vitro quantitative measurement of ISL1 in human tissue homogenates.
Tissue Homogenate
Quantitative
Colorimetric
This assay has high sensitivity and excellent specificity for detection of ISL LIM Homeobox Protein 1 (ISL1)
0.063 ng/mL
<ul> <li>Pre-coated, ready to use 96-well strip plate, flat buttom</li> <li>Plate sealer for 96 wells</li> <li>Reference Standard</li> </ul>

• Standard Diluent

Order at www.antibodies-online.com | www.antikoerper-online.de | www.anticorps-enligne.fr | www.antibodies-online.cn International: +49 (0)241 95 163 153 | USA & Canada: +1 877 302 8632 | support@antibodies-online.com Page 1/5 | Product datasheet for ABIN6957217 | 09/10/2023 | Copyright antibodies-online. All rights reserved.

- Detection Reagent A
- Detection Reagent B
- Assay Diluent A
- Assay Diluent B
- Reagent Diluent (if Detection Reagent is lyophilized)
- TMB Substrate
- Stop Solution
- Wash Buffer (30 x concentrate)
- Instruction manual

## Target Details

Target:	ISL1
Alternative Name:	ISL LIM Homeobox Protein 1 (ISL1) (ISL1 Products)
Pathways:	Positive Regulation of Peptide Hormone Secretion, Intracellular Steroid Hormone Receptor Signaling Pathway, Peptide Hormone Metabolism, Regulation of Intracellular Steroid Hormone Receptor Signaling, Nuclear Hormone Receptor Binding, Chromatin Binding

## Application Details

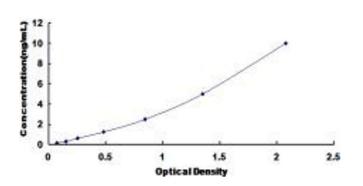
Comment:	Information on standard material:
	The standard might be recombinant protein or natural protein, that will depend on the specific
	kit. Moreover, the expression system is E.coli or yeast or mammal cell. There is 0.05% proclin
	300 in the standard as preservative.
	Information on reagents:
	The stop solution used in the kit is sulfuric acid with concentration of 1 mol/L. And the wash
	solution is TBS. The standard diluent contains 0.02 % sodium azide, assay diluent A and assay
	diluent B contain 0.01% sodium azide. Some kits can contain is BSA in them.
	Information on antibodies:
	The provided antibodies and their host vary in different kits.
Sample Volume:	100 µL
Assay Time:	3 h
Plate:	Pre-coated

Order at www.antibodies-online.com | www.antikoerper-online.de | www.anticorps-enligne.fr | www.antibodies-online.cn International: +49 (0)241 95 163 153 | USA & Canada: +1 877 302 8632 | support@antibodies-online.com Page 2/5 | Product datasheet for ABIN6957217 | 09/10/2023 | Copyright antibodies-online. All rights reserved.

# Application Details

<ol> <li>Standard - Reconstitute the Standard with 1.0 mL of Standard Diluent, keep for 10 minutes room temperature, shake gently (not to foam). The concentration of the standard in the sto solution is 10 ng/mL. Prepare 7 tubes containing 0.5 mL Standard Diluent and produce a double dilution series. Mix each tube thoroughly before the next transfer. Set up 7 points of diluted standard such as 10 ng/mL, 5 ng/mL, 1.25 ng/mL, 1.25 ng/mL, 0.625 ng/mL, 0.312 ng/mL, 0.156 ng/mL, and the last microcentrifuge tube with Standard Diluent is the blank as 0 ng/mL.</li> <li>Detection Reagent A and Detection Reagent B - Briefly spin or centrifuge the stock Detection A and Detection B before use. Dilute to the working concentration with Assay Diluent A and respectively (1:100).</li> <li>Wash Solution - Dilute 20 mL of Wash Solution concentrate (30x) with 580 mL of deionized or distilled water to prepare 600 mL of Wash Solution (1x).</li> <li>TMB substrate - Aspirate the needed dosage of the solution with sterilized tips and do not dump the residual solution into the vial again.</li> <li>Note:</li> <li>Making serial dilution in the wells directly is not permitted.</li> <li>Prepare standard within 15 minutes before assay. Please do not dissolve the reagents at 37 °C directly.</li> <li>Please carefully reconstitute Standards or working Detection Reagent A and B according to the instruction, and avoid foaming and mix gently until the crystals are completely dissolved.</li> <li>The reconstituted Standards, Detection Reagent A and Detection Reagent B can be used or once.</li> <li>If crystals have formed in the Wash Solution concentrate (30x), warm to room temperature and mix gently until the crystals are completely dissolved.</li> </ol>	Protocol:	1. Prepare all reagents, samples and standards,
<ul> <li>4. Aspirate and wash 3 times,</li> <li>5. Add 100µL prepared Detection Reagent B. Incubate 30 minutes at 37 °C,</li> <li>6. Aspirate and wash 5 times,</li> <li>7. Add 90µL Substrate Solution, Incubate 10·20 minutes at 37 °C,</li> <li>8. Add 50µL Stop Solution. Read at 450nm immediately.</li> </ul> Reagent Preparation: <ol> <li>1. Bring all kit components and samples to room temperature (18-25 °C) before use.</li> <li>2. Standard - Reconstitute the Standard with 1.0 mL of Standard Diluent, keep for 10 minutes room temperature, shake gently (not to foam). The concentration of the standard in the sto solution is 10 ng/mL. Prepare 7 tubes containing 0.5 mL. Standard Diluent and produce a double dilution series. Mk each tube thoroughly before the next transfer. Set up 7 points of diluted standard such as 10 ng/mL, 2.5 ng/mL, 1.25 ng/mL, 0.625 ng/mL, 0.312 ng/mL, 0.156 ng/mL, and the last microcentrifuge tube with Standard Diluent is the blank as 0 ng/mL.</li> <li>3. Detection Reagent A and Detection Reagent B - Briefly spin or centrifuge the stock Detection A and Detection Before use. Dilute to the working concentration with Assay Diluent A and respectively (1:100).</li> <li>4. Wash Solution - Dilute 20 mL of Wash Solution concentrate (30x) with 580 mL of deionized or distilled water to prepare 600 mL of Wash Solution (1x).</li> <li>5. TMB substrate - Aspirate the needed dosage of the solution with sterilized tips and do not dump the residual solution into the vial again.</li> <li>Note:</li> <li>1. Making serial dilution in the wells directly is not permitted.</li> <li>2. Prepare standard within 15 minutes before assay. Please do not dissolve the reagents at 37 °C directly.</li> <li>3. Please carefully reconstitute Standards or working Detection Reagent A and B according to the instruction, and avoid foaming and mix gentyl until the crystals are completely dissolved.</li> <li>a. Contaminated water or container for reagent A and Detection Rea</li></ol>		2. Add 100 $\mu$ L standard or sample to each well. Incubate 1 hours at 37 °C,
S. Add 100µL prepared Detection Reagent B. Incubate 30 minutes at 37 °C,         Add 90µL Substrate Solution. Incubate 10-20 minutes at 37 °C,         Reagent Preparation:         1. Bring all kit components and samples to room temperature (18-25 °C) before use.         2. Standard - Reconstitute the Standard with 1.0 mL of Standard Diluent, keep for 10 minutes irroom temperature, shake gently (not to fram). The concentration of the standard in the sto solution is 10 ng/mL. Prepare 7 tubes containing 0.5 mL Standard Diluent and produce a double dilution series. Mix each tube thoroughly before the next transfer. Set up 7 points of diluted standard such as 10 ng/mL, 2.5 ng/mL, 1.25 ng/mL, 0.625 ng/mL, 0.312 ng/mL, 0.156 ng/mL, and the last microcentrifuge tube with Standard Diluent is the blank as 0 ng/mL.         3. Detection Reagent A and Detection Reagent B - Briefly spin or centrifuge the stock Detection A and Detection B before use. Dilute to the working concentration with Assay Diluent A and respectively (1100).         4. Wash Solution - Dilute 20 mL of Wash Solution concentrate (30x) with 580 mL of deionized or distilled water to prepare 600 mL of Wash Solution (1x).         5. TMB substrate - Aspirate the needed dosage of the solution with sterilized tips and do not dump the residual solution into the vial again.         Note:       1. Making serial cillution in the wells directly is not permitted.         2. Prepare standard within 15 minutes before assay. Please do not dissolve the reagents at 37 °C directly.         3. Please carefully reconstitute Standards or working Detection Reagent A and B according to the instruction, and avoid foaming and mix gently untit the crystals are completely dissolved.		3. Aspirate and add 100 $\mu$ L prepared Detection Reagent A. Incubate 1 hour at 37 °C,
<ul> <li>6. Aspirate and wash 5 times,</li> <li>7. Add 90µL Substrate Solution. Incubate 10-20 minutes at 37 °C,</li> <li>8. Add 50µL Stop Solution. Read at 450nm immediately.</li> <li>Reagent Preparation:</li> <li>1. Bring all kit components and samples to room temperature (18-25 °C) before use.</li> <li>2. Standard - Reconstitute the Standard with 1.0 mL of Standard Diluent, keep for 10 minutes room temperature, shake gently (not to foam). The concentration of the standard in the sto solution is 10 ng/mL. Prepare 7 tubes containing 0.5 mL. Standard Diluent and produce a double dilution series. Mix each tube thoroughly before the next transfer. Set up 7 points of diluted standard such as 10 ng/mL, 5 ng/mL, 2.5 ng/mL, 1.25 ng/mL, 0.625 ng/mL, 0.312 ng/mL, 0.156 ng/mL.</li> <li>3. Detection Reagent A and Detection Reagent B - Briefly spin or centrifuge the stock Detection A and Detection B before use. Dilute to the working concentration with Assay Diluent A and respectively (1:100).</li> <li>4. Wash Solution - Dilute 20 mL of Wash Solution concentrate (30%) with 580 mL of deionized or distiled water to prepare 600 mL of Wash Solution with sterilized tips and do not dump the residual solution into the vial again.</li> <li>Note:</li> <li>1. Making serial dilution in the wells directly is not permitted.</li> <li>2. Prepare standard within 15 minutes before assay. Please do not dissolve the reagents at 37 °C directly.</li> <li>3. Please carefully reconstitute Standards or working Detection Reagent A and B according to the instruction, and avoid foaming and mix gently until the crystals are completely dissolved.</li> <li>4. The reconstituted Standards, Detection Reagent A and Detection Reagent B can be used or once.</li> <li>5. If crystals have formed in the Wash Solution concentrate (30x), warm to room temperature and mix gently until the crystals are completely dissolved.</li> <li>6. Contaminated water or container for reagent preparation will influence the detection result.</li> <li>Sample Preparation:</li> <li>t</li></ul>		4. Aspirate and wash 3 times,
7. Add 90µL Substrate Solution. Incubate 10-20 minutes at 37 °C,         8. Add 50µL Stop Solution. Read at 450nm immediately.         Reagent Preparation:       1. Bring all kit components and samples to room temperature (18-25 °C) before use.         2. Standard - Reconstitute the Standard with 1.0 rm, of Standard Diluent, keep for 10 minutes room temperature, shake gently (not to foam). The concentration of the standard in the sto solution is 10 ng/mL. Prepare 7 tubes containing 0.5 rm. Standard Diluent and produce a double diluted standard such as 10 ng/mL, 5 ng/mL, 2.5 ng/mL, 0.625 ng/mL, 0.312 ng/mL, 0.156 ng/mL, and the last microcentrifuge tube with Standard Diluent is the blank as 0 ng/mL.         3. Detection Reagent A and Detection Reagent B - Briefly spin or centrifuge the stock Detection A and Detection B before use. Dilute to the working concentration with Assay Diluent A and respectively (1:100).         4. Wash Solution - Dilute 20 mL of Wash Solution (1x).         5. TMB substrate - Aspirate the needed dosage of the solution with sterilized tips and do not dump the residual solution into the vial again.         Note:         1. Making serial dilution in the wells directly is not permitted.         2. Prepare standard within 15 minutes before assay. Please do not dissolve the reagents at 37 °C directly.         3. Please carefully reconstitute Standards or working Detection Reagent A and B according to the instruction, and avoid foaming and mix gently until the crystals are completely dissolved.         4. The reconstituted Standards, Detection Reagent A and Detection Reagent B can be used or once.         5. If crystals have formed in the Wash Solut		5. Add 100µL prepared Detection Reagent B. Incubate 30 minutes at 37 °C,
8. Add 50µL Stop Solution. Read at 450nm immediately:         Reagent Preparation:       1. Bring all kit components and samples to room temperature (18-25 °C) before use.         2. Standard - Reconstitute the Standard with 1.0 mL of Standard Diluent, keep for 10 minutes room temperature, shake gently (not to foam). The concentration of the standard in the sto solution is 10 ng/mL, Prepare 7 tubes containing 0.5 mL Standard Diluent and produce a double dilution series. Mix each tube thoroughly before the next transfer. Set up 7 points of diluted standard such as 10 ng/mL, 5 ng/mL, 1.25 ng/mL, 0.625 ng/mL, 0.312 ng/mL, 0.156 ng/mL, and the last microcentrifuge tube with Standard Diluent is the blank as 0 ng/mL.         3. Detection Reagent A and Detection Reagent B - Briefly spin or centrifuge the stock Detection A and Detection Before use. Dilute to the working concentration with Assay Diluent A and respectively (11:00).         4. Wash Solution - Dilute 20 mL of Wash Solution concentrate (30x) with 580 mL of deionized or distilled water to prepare 600 mL of Wash Solution (1x).         5. TMB substrate - Aspirate the needed dosage of the solution with sterilized tips and do not dump the residual solution into the vial again.         Note:       1. Making serial dilution in the wells directly is not permitted.         2. Prepare standard within 15 minutes before assay. Please do not dissolve the reagents at 37°C directly.         3. Please carefully reconstitute Standards or working Detection Reagent A and B according to the instruction, and avoid foaming and mix gently until the crystals are completely dissolved.         6. Contaminate water or container for reagent A and Detection Reagent B can be used or once.      <		6. Aspirate and wash 5 times,
<ul> <li>Reagent Preparation:</li> <li>1. Bring all kit components and samples to room temperature (18-25 °C) before use.</li> <li>2. Standard - Reconstitute the Standard with 1.0 mL of Standard Diluent, keep for 10 minutes room temperature, shake gently (not to foam). The concentration of the standard in the sto solution is 10 ng/mL, Prepare 7 tubes containing 0.5 mL Standard Diluent and produce a double dilution series. Nix each tube thoroughly before the next transfer. Set up 7 points of diluted standard such as 10 ng/mL, 5 ng/mL, 1.25 ng/mL, 1.25 ng/mL, 0.625 ng/mL, 0.312 ng/mL, 0.156 ng/mL, and the last microcentrifuge tube with Standard Diluent is the blank as 0 ng/mL.</li> <li>3. Detection Reagent A and Detection Reagent B - Briefly spin or centrifuge the stock Detection A and Detection Before use. Dilute to the working concentration with Assay Diluent A and respectively (1:100).</li> <li>4. Wash Solution - Dilute 20 mL of Wash Solution concentrate (30x) with 580 mL of deionized or distilled water to prepare 600 mL of Wash Solution (1x).</li> <li>5. TMB substrate - Aspirate the needed dosage of the solution with sterilized tips and do not dump the residual solution in the wells directly is not permitted.</li> <li>2. Prepare standard within 15 minutes before assay. Please do not dissolve the reagents at 37 °C directly.</li> <li>3. Please carefully reconstitute Standards or working Detection Reagent A and B according to the instruction, and avoid foaming and mix gently until the crystals are completely dissolved.</li> <li>4. The reconstituted Standards, Detection Reagent A and Detection Reagent B can be used or once.</li> <li>5. If crystals have formed in the Wash Solution concentrate (30x), warm to room temperature and mix gently until the crystals are completely dissolved.</li> <li>6. Contaminated water or container for reagent preparation will influence the detection result.</li> </ul>		7. Add 90µL Substrate Solution. Incubate 10-20 minutes at 37 °C,
<ul> <li>2. Standard - Reconstitute the Standard with 1.0 mL of Standard Diluent, keep for 10 minutes room temperature, shake gently (not to foam). The concentration of the standard in the sto solution is 10 ng/mL. Prepare 7 tubes containing 0.5 mL Standard Diluent and produce a double dilution series. Mix each tube thoroughly before the next transfer. Set up 7 points of diluted standard such as 10 ng/mL, 5 ng/mL, 2.5 ng/mL, 1.25 ng/mL, 0.625 ng/mL, 0.312 ng/mL, 0.156 ng/mL, and the last microcentrifuge tube with Standard Diluent is the blank as 0 ng/mL.</li> <li>3. Detection Reagent A and Detection Reagent B - Briefly spin or centrifuge the stock Detection A and Detection Before use. Dilute to the working concentration with Assay Diluent A and respectively (1:100).</li> <li>4. Wash Solution - Dilute 20 mL of Wash Solution concentrate (30x) with 580 mL of deionized or distilled water to prepare 600 mL of Wash Solution (1x).</li> <li>5. TMB substrate - Aspirate the needed dosage of the solution with sterilized tips and do not dump the residual solution into the vial again.</li> <li>Note:         <ol> <li>Making serial dilution in the wells directly is not permitted.</li> <li>Prepare standard within 15 minutes before assay. Please do not dissolve the reagents at 37 °C directly.</li> <li>Please carefully reconstitute Standards or working Detection Reagent A and B according to the instruction, and avoid foaming and mix gently until the crystals are completely dissolve. To minimize imprecision caused by pipetting, use small volumes and ensure that pipettors are calibrated. It is recommended to suck more than 10µL for once pipetting.</li> <li>If crystals have formed in the Wash Solution concentrate (30x), warm to room temperature and mix gently until the crystals are completely dissolved.</li> <li>Contaminated water or container for reagent preparation will influence the detection result.</li> </ol> </li> <li>Sample Prepar</li></ul>		8. Add 50µL Stop Solution. Read at 450nm immediately.
<ul> <li>room temperature, shake gently (not to foam). The concentration of the standard in the sto solution is 10 ng/mL. Prepare 7 tubes containing 0.5 mL Standard Diluent and produce a double diluted standard such as 10 ng/mL, 5 ng/mL, 2.5 ng/mL, 1.25 ng/mL, 0.625 ng/mL, 0.312 ng/mL, 0.156 ng/mL, and the last microcentrifuge tube with Standard Diluent is the blank as 0 ng/mL.</li> <li>3. Detection Reagent A and Detection Reagent B - Briefly spin or centrifuge the stock Detection A and Detection B before use. Dilute to the working concentration with Assay Diluent A and respectively (1:100).</li> <li>4. Wash Solution - Dilute 20 mL of Wash Solution concentrate (30x) with 580 mL of deionized or distilled water to prepare 600 mL of Wash Solution (1x).</li> <li>5. TMB substrate - Aspirate the needed dosage of the solution with sterilized tips and do not dump the residual solution in the wells directly is not permitted.</li> <li>2. Prepare standard within 15 minutes before assay. Please do not dissolve the reagents at 37 °C directly.</li> <li>3. Please carefully reconstitute Standards or working Detection Reagent A and B according to the instruction, and avoid foaming and mix gently until the crystals are completely dissolve To minimize imprecision caused by pipetting, use small volumes and ensure that pipettors are calibrated. It is recommended to suck more than 10µL for once pipetting.</li> <li>4. The reconstituted Standards, Detection Reagent A and Detection Reagent B can be used or once.</li> <li>5. If crystals have formed in the Wash Solution concentrate (30x), warm to room temperature and mix gently until the crystals are completely dissolved.</li> <li>6. Contaminated water or container for reagent preparation will influence the detection result.</li> <li>Sample Preparation:</li> <li>It is recommended to use fresh samples without long storage, otherwise protein degradatio and denaturationmay occur in these samples, leading to false results. Samples should therefore</li></ul>	Reagent Preparation:	
<ul> <li>double dilution series. Mix each tube thoroughly before the next transfer. Set up 7 points of diluted standard such as 10 ng/mL, 5 ng/mL, 2.5 ng/mL, 1.25 ng/mL, 0.625 ng/mL, 0.312 ng/mL, 0.156 ng/mL, and the last microcentrifuge tube with Standard Diluent is the blank as 0 ng/mL.</li> <li>3. Detection Reagent A and Detection Reagent B - Briefly spin or centrifuge the stock Detection A and Detection B before use. Dilute to the working concentration with Assay Diluent A and respectively (1:100).</li> <li>4. Wash Solution - Dilute 20 mL of Wash Solution concentrate (30x) with 580 mL of deionized or distilled water to prepare 600 mL of Wash Solution (1x).</li> <li>5. TMB substrate - Aspirate the needed dosage of the solution with sterilized tips and do not dump the residual solution into the vial again.</li> <li>Note:         <ol> <li>Making serial dilution in the wells directly is not permitted.</li> <li>Prepare standard within 15 minutes before assay. Please do not dissolve the reagents at 37 °C directly.</li> <li>Please carefully reconstitute Standards or working Detection Reagent A and B according to the instruction, and avoid foaming and mix gently until the crystals are completely dissolve. To minimize imprecision caused by pipetting, use small volumes and ensure that pipettors are calibrated. It is recommended to suck more than 10µL for once pipetting.</li> <li>The reconstituted Standards, Detection Reagent A and Detection Reagent B can be used or once.</li> <li>I crystals have formed in the Wash Solution concentrate (30x), warm to room temperature and mix gently until the crystals are completely dissolved.</li> <li>Contaminated water or container for reagent preparation will influence the detection result.</li> </ol> </li> <li>Sample Preparation:         <ul> <li>It is recommended to use fresh samples without long storage, otherwise protein degradatic and denaturationmay occur in</li></ul></li></ul>		room temperature, shake gently (not to foam). The concentration of the standard in the stoc
<ul> <li>diluted standard such as 10 ng/mL, 5 ng/mL, 1.25 ng/mL, 0.625 ng/mL, 0.312 ng/mL, 0.156 ng/mL, and the last microcentrifuge tube with Standard Diluent is the blank as 0 ng/mL.</li> <li>Detection Reagent A and Detection Reagent B - Briefly spin or centrifuge the stock Detection A and Detection B before use. Dilute to the working concentration with Assay Diluent A and respectively (1:100).</li> <li>Wash Solution - Dilute 20 mL of Wash Solution concentrate (30x) with 580 mL of deionized or distilled water to prepare 600 mL of Wash Solution (1x).</li> <li>TMB substrate - Aspirate the needed dosage of the solution with sterilized tips and do not dump the residual solution into the vial again.</li> <li>Note:         <ol> <li>Making serial dilution in the wells directly is not permitted.</li> <li>Prepare standard within 15 minutes before assay. Please do not dissolve the reagents at 37 °C directly.</li> <li>Please carefully reconstitute Standards or working Detection Reagent A and B according to the instruction, and avoid foaming and mix gently until the crystals are completely dissolved.</li> <li>The reconstituted Standards, Detection Reagent A and Detection resure that pipettors are calibrated. It is recommended to suck more than 10µL for once pipetting.</li> <li>The reconstituted Standards, Detection Reagent A and Decection result.</li> <li>En crystals have formed in the Wash Solution concentrate (30x), warm to room temperature and mix gently until the crystals are completely dissolved.</li> <li>Contaminated water or container for reagent preparation will influence the detection result.</li> </ol> </li> <li>Sample Preparation:         <ul> <li>It is recommended to use fresh samples without long storage, otherwise protein degradatic and denturationmay occur in these samples, leading to false results. Samples should therefore be stored for a short periodat 2 - 8 °C or aliquoted</li></ul></li></ul>		solution is 10 ng/mL. Prepare 7 tubes containing 0.5 mL Standard Diluent and produce a
<ul> <li>0.312 ng/mL, 0.156 ng/mL, and the last microcentrifuge tube with Standard Diluent is the blank as 0 ng/mL.</li> <li>3. Detection Reagent A and Detection Reagent B - Briefly spin or centrifuge the stock Detection A and Detection B before use. Dilute to the working concentration with Assay Diluent A and respectively (1100).</li> <li>4. Wash Solution - Dilute 20 mL of Wash Solution concentrate (30x) with 580 mL of deionized or distilled water to prepare 600 mL of Wash Solution (1x).</li> <li>5. TMB substrate - Aspirate the needed dosage of the solution with sterilized tips and do not dump the residual solution into the vial again.</li> <li>Note:         <ol> <li>Making serial dilution in the wells directly is not permitted.</li> <li>Prepare standard within 15 minutes before assay. Please do not dissolve the reagents at 37 °C directly.</li> <li>Please carefully reconstitute Standards or working Detection Reagent A and B according to the instruction, and avoid foaming and mix gently until the crystals are completely dissolved.</li> <li>The reconstituted Standards, Detection Reagent A and Detection Reagent B can be used or once.</li> <li>If crystals have formed in the Wash Solution concentrate (30x), warm to room temperature and mix gently until the crystals are completely dissolved.</li> <li>Contaminated water or container for reagent preparation will influence the detection result.</li> </ol> </li> <li>Sample Preparation:         <ul> <li>It is recommended to use fresh samples without long storage, otherwise protein degradatic and denaturationmay occur in these samples, leading to false results. Samples should therefore be stored for a short periodat 2 - 8 °C or aliquoted at -20 °C (</li> <li>months). Repeated freeze-thawcycles should be avoided. Prior to assay, the frozen</li> </ul> </li> </ul>		double dilution series. Mix each tube thoroughly before the next transfer. Set up 7 points of
<ul> <li>blank as 0 ng/mL.</li> <li>3. Detection Reagent A and Detection Reagent B - Briefly spin or centrifuge the stock Detection A and Detection B before use. Dilute to the working concentration with Assay Diluent A and respectively (1:100).</li> <li>4. Wash Solution - Dilute 20 mL of Wash Solution concentrate (30x) with 580 mL of deionized or distilled water to prepare 600 mL of Wash Solution (1x).</li> <li>5. TMB substrate - Aspirate the needed dosage of the solution with sterilized tips and do not dump the residual solution into the vial again.</li> <li>Note: <ol> <li>Making serial dilution in the wells directly is not permitted.</li> <li>Prepare standard within 15 minutes before assay. Please do not dissolve the reagents at 37 °C directly.</li> <li>Please carefully reconstitute Standards or working Detection Reagent A and B according to the instruction, and avoid foaming and mix gently until the crystals are completely dissolved To minimize imprecision caused by pipetting, use small volumes and ensure that pipettors are calibrated. It is recommended to suck more than 10µL for once pipetting.</li> <li>The reconstituted Standards, Detection Reagent A and Detection Reagent B can be used or once.</li> <li>If crystals have formed in the Wash Solution concentrate (30x), warm to room temperature and mix gently until the crystals are completely dissolved.</li> <li>Contaminated water or container for reagent preparation will influence the detection result.</li> </ol> </li> <li>Sample Preparation: <ul> <li>It is recommended to use fresh samples without long storage, otherwise protein degradatid and denaturationmay occur in these samples, leading to false results. Samples should therefore be stored for a short periodat 2 - 8 °C or aliquoted at -20 °C (≤1 month) or -80 °C (≤3 months). Repeated freeze-thawcycles should be avoided. Prior to assay, the frozen</li> </ul></li></ul>		
<ul> <li>A and Detection B before use. Dilute to the working concentration with Assay Diluent A and respectively (1:100).</li> <li>4. Wash Solution - Dilute 20 mL of Wash Solution concentrate (30x) with 580 mL of deionized or distilled water to prepare 600 mL of Wash Solution (1x).</li> <li>5. TMB substrate - Aspirate the needed dosage of the solution with sterilized tips and do not dump the residual solution into the vial again.</li> <li>Note: <ol> <li>Making serial dilution in the wells directly is not permitted.</li> <li>Prepare standard within 15 minutes before assay. Please do not dissolve the reagents at 37 °C directly.</li> <li>Please carefully reconstitute Standards or working Detection Reagent A and B according to the instruction, and avoid foaming and mix gently until the crystals are completely dissolved. To minimize imprecision caused by pipetting, use small volumes and ensure that pipettors are calibrated. It is recommended to suck more than 10µL for once pipetting.</li> <li>The reconstituted Standards, Detection Reagent A and Detection Reagent B can be used or once.</li> <li>If crystals have formed in the Wash Solution concentrate (30x), warm to room temperature and mix gently until the crystals are completely dissolved.</li> <li>Contaminated water or container for reagent preparation will influence the detection result.</li> </ol> </li> </ul>		
<ul> <li>respectively (1:100).</li> <li>4. Wash Solution - Dilute 20 mL of Wash Solution concentrate (30x) with 580 mL of deionized or distilled water to prepare 600 mL of Wash Solution (1x).</li> <li>5. TMB substrate - Aspirate the needed dosage of the solution with sterilized tips and do not dump the residual solution into the vial again.</li> <li>Note: <ol> <li>Making serial dilution in the wells directly is not permitted.</li> <li>Prepare standard within 15 minutes before assay. Please do not dissolve the reagents at 37 °C directly.</li> <li>Please carefully reconstitute Standards or working Detection Reagent A and B according to the instruction, and avoid foaming and mix gently until the crystals are completely dissolved. To minimize imprecision caused by pipetting, use small volumes and ensure that pipettors are calibrated. It is recommended to suck more than 10µL for once pipetting.</li> <li>The reconstituted Standards, Detection Reagent A and Detection Reagent B can be used or once.</li> <li>If crystals have formed in the Wash Solution concentrate (30x), warm to room temperature and mix gently until the crystals are completely dissolved.</li> <li>Contaminated water or container for reagent preparation will influence the detection result.</li> </ol> </li> <li>Sample Preparation: <ul> <li>It is recommended to use fresh samples without long storage, otherwise protein degradatic and denaturationmay occur in these samples, leading to false results. Samples should therefore be stored for a short periodat 2 - 8 °C or aliquoted at -20 °C (≤1 month) or -80 °C (≤3 months). Repeated freeze-thawcycles should be avoided. Prior to assay, the frozen</li> </ul></li></ul>		3. Detection Reagent A and Detection Reagent B - Briefly spin or centrifuge the stock Detectior
<ul> <li>or distilled water to prepare 600 mL of Wash Solution (1x).</li> <li>5. TMB substrate - Aspirate the needed dosage of the solution with sterilized tips and do not dump the residual solution into the vial again.</li> <li>Note: <ol> <li>Making serial dilution in the wells directly is not permitted.</li> <li>Prepare standard within 15 minutes before assay. Please do not dissolve the reagents at 37 °C directly.</li> <li>Please carefully reconstitute Standards or working Detection Reagent A and B according to the instruction, and avoid foaming and mix gently until the crystals are completely dissolved. To minimize imprecision caused by pipetting, use small volumes and ensure that pipettors are calibrated. It is recommended to suck more than 10µL for once pipetting.</li> <li>The reconstituted Standards, Detection Reagent A and Detection Reagent B can be used or once.</li> <li>If crystals have formed in the Wash Solution concentrate (30x), warm to room temperature and mix gently until the crystals are completely dissolved.</li> <li>Contaminated water or container for reagent preparation will influence the detection result.</li> </ol> </li> <li>Sample Preparation: <ul> <li>It is recommended to use fresh samples without long storage, otherwise protein degradatic and denaturationmay occur in these samples, leading to false results. Samples should therefore be stored for a short periodat 2 - 8 °C or aliquoted at -20 °C (≤1 month) or -80 °C (≤3 months). Repeated freeze-thawcycles should be avoided. Prior to assay, the frozen</li> </ul> </li> </ul>		A and Detection B before use. Dilute to the working concentration with Assay Diluent A and respectively (1:100).
<ul> <li>5. TMB substrate - Aspirate the needed dosage of the solution with sterilized tips and do not dump the residual solution into the vial again.</li> <li>Note: <ol> <li>Making serial dilution in the wells directly is not permitted.</li> <li>Prepare standard within 15 minutes before assay. Please do not dissolve the reagents at 37 °C directly.</li> <li>Please carefully reconstitute Standards or working Detection Reagent A and B according to the instruction, and avoid foaming and mix gently until the crystals are completely dissolved. To minimize imprecision caused by pipetting, use small volumes and ensure that pipettors are calibrated. It is recommended to suck more than 10µL for once pipetting.</li> <li>The reconstituted Standards, Detection Reagent A and Detection Reagent B can be used or once.</li> <li>If crystals have formed in the Wash Solution concentrate (30x), warm to room temperature and mix gently until the crystals are completely dissolved.</li> <li>Contaminated water or container for reagent preparation will influence the detection result.</li> </ol> </li> <li>Sample Preparation: <ul> <li>It is recommended to use fresh samples without long storage, otherwise protein degradatic and denaturationmay occur in these samples, leading to false results. Samples should therefore be stored for a short periodat 2 - 8 °C or aliquoted at -20 °C (&lt;1 month) or -80 °C (&lt;3 months). Repeated freeze-thawcycles should be avoided. Prior to assay, the frozen</li> </ul></li></ul>		
dump the residual solution into the vial again.         Note:         1. Making serial dilution in the wells directly is not permitted.         2. Prepare standard within 15 minutes before assay. Please do not dissolve the reagents at 37 °C directly.         3. Please carefully reconstitute Standards or working Detection Reagent A and B according to the instruction, and avoid foaming and mix gently until the crystals are completely dissolved.         7. To minimize imprecision caused by pipetting, use small volumes and ensure that pipettors are calibrated. It is recommended to suck more than 10µL for once pipetting.         4. The reconstituted Standards, Detection Reagent A and Detection Reagent B can be used or once.         5. If crystals have formed in the Wash Solution concentrate (30x), warm to room temperature and mix gently until the crystals are completely dissolved.         6. Contaminated water or container for reagent preparation will influence the detection result.         Sample Preparation: <ul> <li>It is recommended to use fresh samples without long storage, otherwise protein degradatic and denaturationmay occur in these samples, leading to false results. Samples should therefore be stored for a short periodat 2 - 8 °C or aliquoted at -20 °C (s1 month) or -80 °C (s3 months). Repeated freeze-thawcycles should be avoided. Prior to assay, the frozen</li></ul>		or distilled water to prepare 600 mL of Wash Solution (1x).
<ul> <li>Note: <ol> <li>Making serial dilution in the wells directly is not permitted.</li> <li>Prepare standard within 15 minutes before assay. Please do not dissolve the reagents at 37 °C directly.</li> <li>Please carefully reconstitute Standards or working Detection Reagent A and B according to the instruction, and avoid foaming and mix gently until the crystals are completely dissolved. To minimize imprecision caused by pipetting, use small volumes and ensure that pipettors are calibrated. It is recommended to suck more than 10µL for once pipetting.</li> <li>The reconstituted Standards, Detection Reagent A and Detection Reagent B can be used on once.</li> <li>If crystals have formed in the Wash Solution concentrate (30x), warm to room temperature and mix gently until the crystals are completely dissolved.</li> <li>Contaminated water or container for reagent preparation will influence the detection result.</li> </ol> </li> <li>Sample Preparation: <ul> <li>It is recommended to use fresh samples without long storage, otherwise protein degradatic and denaturationmay occur in these samples, leading to false results. Samples should therefore be stored for a short periodat 2 - 8 °C or aliquoted at -20 °C (≤1 month) or -80 °C (≤3 months). Repeated freeze-thawcycles should be avoided. Prior to assay, the frozen</li> </ul></li></ul>		5. TMB substrate - Aspirate the needed dosage of the solution with sterilized tips and do not
<ol> <li>Making serial dilution in the wells directly is not permitted.</li> <li>Prepare standard within 15 minutes before assay. Please do not dissolve the reagents at 37 °C directly.</li> <li>Please carefully reconstitute Standards or working Detection Reagent A and B according to the instruction, and avoid foaming and mix gently until the crystals are completely dissolved To minimize imprecision caused by pipetting, use small volumes and ensure that pipettors are calibrated. It is recommended to suck more than 10µL for once pipetting.</li> <li>The reconstituted Standards, Detection Reagent A and Detection Reagent B can be used on once.</li> <li>If crystals have formed in the Wash Solution concentrate (30x), warm to room temperature and mix gently until the crystals are completely dissolved.</li> <li>Contaminated water or container for reagent preparation will influence the detection result.</li> </ol> Sample Preparation: <ul> <li>It is recommended to use fresh samples without long storage, otherwise protein degradatic and denaturationmay occur in these samples, leading to false results. Samples should therefore be stored for a short periodat 2 - 8 °C or aliquoted at -20 °C (≤1 month) or -80 °C (</li> </ul>		dump the residual solution into the vial again.
<ul> <li>2. Prepare standard within 15 minutes before assay. Please do not dissolve the reagents at 37 °C directly.</li> <li>3. Please carefully reconstitute Standards or working Detection Reagent A and B according to the instruction, and avoid foaming and mix gently until the crystals are completely dissolved. To minimize imprecision caused by pipetting, use small volumes and ensure that pipettors are calibrated. It is recommended to suck more than 10µL for once pipetting.</li> <li>4. The reconstituted Standards, Detection Reagent A and Detection Reagent B can be used or once.</li> <li>5. If crystals have formed in the Wash Solution concentrate (30x), warm to room temperature and mix gently until the crystals are completely dissolved.</li> <li>6. Contaminated water or container for reagent preparation will influence the detection result.</li> </ul> Sample Preparation: <ul> <li>It is recommended to use fresh samples without long storage, otherwise protein degradatio and denaturationmay occur in these samples, leading to false results. Samples should therefore be stored for a short periodat 2 - 8 °C or aliquoted at -20 °C (≤1 month) or -80 °C (≤3 months). Repeated freeze-thawcycles should be avoided. Prior to assay, the frozen</li></ul>		Note:
<ul> <li>37 °C directly.</li> <li>3. Please carefully reconstitute Standards or working Detection Reagent A and B according to the instruction, and avoid foaming and mix gently until the crystals are completely dissolved. To minimize imprecision caused by pipetting, use small volumes and ensure that pipettors are calibrated. It is recommended to suck more than 10µL for once pipetting.</li> <li>4. The reconstituted Standards, Detection Reagent A and Detection Reagent B can be used on once.</li> <li>5. If crystals have formed in the Wash Solution concentrate (30x), warm to room temperature and mix gently until the crystals are completely dissolved.</li> <li>6. Contaminated water or container for reagent preparation will influence the detection result.</li> </ul> Sample Preparation: <ul> <li>It is recommended to use fresh samples without long storage, otherwise protein degradatio and denaturationmay occur in these samples, leading to false results. Samples should therefore be stored for a short periodat 2 - 8 °C or aliquoted at -20 °C (≤1 month) or -80 °C (≤3 months). Repeated freeze-thawcycles should be avoided. Prior to assay, the frozen</li></ul>		1. Making serial dilution in the wells directly is not permitted.
<ul> <li>the instruction, and avoid foaming and mix gently until the crystals are completely dissolved. To minimize imprecision caused by pipetting, use small volumes and ensure that pipettors are calibrated. It is recommended to suck more than 10µL for once pipetting.</li> <li>4. The reconstituted Standards, Detection Reagent A and Detection Reagent B can be used on once.</li> <li>5. If crystals have formed in the Wash Solution concentrate (30x), warm to room temperature and mix gently until the crystals are completely dissolved.</li> <li>6. Contaminated water or container for reagent preparation will influence the detection result.</li> </ul> Sample Preparation: <ul> <li>It is recommended to use fresh samples without long storage, otherwise protein degradation and denaturationmay occur in these samples, leading to false results. Samples should therefore be stored for a short periodat 2 - 8 °C or aliquoted at -20 °C (≤1 month) or -80 °C (≤ 3 months). Repeated freeze-thawcycles should be avoided. Prior to assay, the frozen</li></ul>		
<ul> <li>To minimize imprecision caused by pipetting, use small volumes and ensure that pipettors are calibrated. It is recommended to suck more than 10µL for once pipetting.</li> <li>4. The reconstituted Standards, Detection Reagent A and Detection Reagent B can be used on once.</li> <li>5. If crystals have formed in the Wash Solution concentrate (30x), warm to room temperature and mix gently until the crystals are completely dissolved.</li> <li>6. Contaminated water or container for reagent preparation will influence the detection result.</li> <li>Sample Preparation:</li> <li>It is recommended to use fresh samples without long storage, otherwise protein degradatic and denaturationmay occur in these samples, leading to false results. Samples should therefore be stored for a short periodat 2 - 8 °C or aliquoted at -20 °C (≤1 month) or -80 °C (≤3 months). Repeated freeze-thawcycles should be avoided. Prior to assay, the frozen</li> </ul>		3. Please carefully reconstitute Standards or working Detection Reagent A and B according to
<ul> <li>are calibrated. It is recommended to suck more than 10µL for once pipetting.</li> <li>4. The reconstituted Standards, Detection Reagent A and Detection Reagent B can be used on once.</li> <li>5. If crystals have formed in the Wash Solution concentrate (30x), warm to room temperature and mix gently until the crystals are completely dissolved.</li> <li>6. Contaminated water or container for reagent preparation will influence the detection result.</li> </ul> Sample Preparation: <ul> <li>It is recommended to use fresh samples without long storage, otherwise protein degradation and denaturationmay occur in these samples, leading to false results. Samples should therefore be stored for a short periodat 2 - 8 °C or aliquoted at -20 °C (≤1 month) or -80 °C (≤3 months). Repeated freeze-thawcycles should be avoided. Prior to assay, the frozen</li></ul>		the instruction, and avoid foaming and mix gently until the crystals are completely dissolved
<ul> <li>4. The reconstituted Standards, Detection Reagent A and Detection Reagent B can be used on once.</li> <li>5. If crystals have formed in the Wash Solution concentrate (30x), warm to room temperature and mix gently until the crystals are completely dissolved.</li> <li>6. Contaminated water or container for reagent preparation will influence the detection result.</li> <li>Sample Preparation:</li> <li>It is recommended to use fresh samples without long storage, otherwise protein degradation and denaturationmay occur in these samples, leading to false results. Samples should therefore be stored for a short periodat 2 - 8 °C or aliquoted at -20 °C (≤1 month) or -80 °C (≤3 months). Repeated freeze-thawcycles should be avoided. Prior to assay, the frozen</li> </ul>		To minimize imprecision caused by pipetting, use small volumes and ensure that pipettors
<ul> <li>once.</li> <li>5. If crystals have formed in the Wash Solution concentrate (30x), warm to room temperature and mix gently until the crystals are completely dissolved.</li> <li>6. Contaminated water or container for reagent preparation will influence the detection result.</li> <li>Sample Preparation:</li> <li>It is recommended to use fresh samples without long storage, otherwise protein degradation and denaturationmay occur in these samples, leading to false results. Samples should therefore be stored for a short periodat 2 - 8 °C or aliquoted at -20 °C (≤1 month) or -80 °C (≤3 months). Repeated freeze-thawcycles should be avoided. Prior to assay, the frozen</li> </ul>		are calibrated. It is recommended to suck more than $10\mu L$ for once pipetting.
<ul> <li>and mix gently until the crystals are completely dissolved.</li> <li>6. Contaminated water or container for reagent preparation will influence the detection result.</li> <li>Sample Preparation:</li> <li>It is recommended to use fresh samples without long storage, otherwise protein degradation and denaturationmay occur in these samples, leading to false results. Samples should therefore be stored for a short periodat 2 - 8 °C or aliquoted at -20 °C (≤1 month) or -80 °C (≤3 months). Repeated freeze-thawcycles should be avoided. Prior to assay, the frozen</li> </ul>		<ol> <li>The reconstituted Standards, Detection Reagent A and Detection Reagent B can be used on once.</li> </ol>
<ul> <li>6. Contaminated water or container for reagent preparation will influence the detection result.</li> <li>Sample Preparation:         <ul> <li>It is recommended to use fresh samples without long storage, otherwise protein degradation and denaturationmay occur in these samples, leading to false results. Samples should therefore be stored for a short periodat 2 - 8 °C or aliquoted at -20 °C (≤1 month) or -80 °C (≤3 months). Repeated freeze-thawcycles should be avoided. Prior to assay, the frozen</li> </ul> </li> </ul>		5. If crystals have formed in the Wash Solution concentrate (30x), warm to room temperature
<ul> <li>Sample Preparation:         <ul> <li>It is recommended to use fresh samples without long storage, otherwise protein degradation and denaturationmay occur in these samples, leading to false results. Samples should therefore be stored for a short periodat 2 - 8 °C or aliquoted at -20 °C (≤1 month) or -80 °C (so months). Repeated freeze-thawcycles should be avoided. Prior to assay, the frozen</li> </ul> </li> </ul>		and mix gently until the crystals are completely dissolved.
and denaturationmay occur in these samples, leading to false results. Samples should therefore be stored for a short periodat 2 - 8 °C or aliquoted at -20 °C (≤1 month) or -80 °C (: 3 months). Repeated freeze-thawcycles should be avoided. Prior to assay, the frozen		6. Contaminated water or container for reagent preparation will influence the detection result.
therefore be stored for a short periodat 2 - 8 °C or aliquoted at -20 °C (≤1 month) or -80 °C ( 3 months). Repeated freeze-thawcycles should be avoided. Prior to assay, the frozen	Sample Preparation:	It is recommended to use fresh samples without long storage, otherwise protein degradatio
3 months). Repeated freeze-thawcycles should be avoided. Prior to assay, the frozen		
samples should be slowly thawed and centrifuged toremove precipitates.		
		samples should be slowly thawed and centrifuged toremove precipitates.

	<ul> <li>If the sample type is not specified in the instructions, a preliminary test is necessary to determinecompatibility with the kit.</li> </ul>
	• If a lysis buffer is used to prepare tissue homogenates or cell culture supernatant, there is a possibility of causing a deviation due to the introduced chemical substance. The recommended dilution factor is for reference only.
	• Please estimate the concentration of the samples before performing the test. If the values are not in therange of the standard curve, the optimal sample dilution for the particular experiment has to be determined. Samples should then be diluted with PBS (pH =7.0-7.2).
Assay Precision:	Intra-assay Precision (Precision within an assay): 3 samples with low, middle and high level of
	target were tested 20 times on one plate, respectively.
	Inter-assay Precision (Precision between assays): 3 samples with low, middle and high level of
	target were tested on 3 different plates, 8 replicates in each plate.
	CV(%) = SD/meanX100
	Intra-Assay: CV < 10%
	Inter-Assay: CV < 12%
Restrictions:	For Research Use only
Handling	
Precaution of Use:	The Stop Solution suggested for use with this kit is an acid solution. Wear eye, hand, face, and
	clothing protection when using this material.
Storage:	4 °C/-20 °C
Storage Comment:	<ol> <li>For unopened kit: All reagents should be stored according to the labels on the vials. The Standard, Detection Reagent A, Detection Reagent B, and 96-well Strip Plate should be stored at -20 °C upon receipt, while the other reagents should be stored at 4 °C.</li> </ol>
	2. For opened kits: the remaining reagents must be stored according to the above storage conditions. In addition, please return the unused wells to the foil pouch containing the desiccant and seal the foil pouch with the zipper.
Expiry Date:	6 months



### **ELISA**

Image 1. Typical standard curve

Order at www.antibodies-online.com | www.antikoerper-online.de | www.anticorps-enligne.fr | www.antibodies-online.cn International: +49 (0)241 95 163 153 | USA & Canada: +1 877 302 8632 | support@antibodies-online.com Page 5/5 | Product datasheet for ABIN6957217 | 09/10/2023 | Copyright antibodies-online. All rights reserved.