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Datasheet for ABIN6970225 SIRPA ELISA Kit

Image



Overview

Quantity:	96 tests
Target:	SIRPA
Reactivity:	Human
Method Type:	Sandwich ELISA
Detection Range:	0.156 ng/mL - 10 ng/mL
Minimum Detection Limit:	0.156 ng/mL
Application:	ELISA

Product Details

Purpose:	For quantitative detection of SIRPa in serum, plasma, tissue homogenates.
Sample Type:	Plasma, Serum, Tissue Homogenate
Analytical Method:	Quantitative
Detection Method:	Colorimetric
Specificity:	This assay has high sensitivity and excellent specificity for detection of SIRPa. No significant cross-reactivity or interference between SIRPa and analogues was observed. Note: Limited by current skills and knowledge, it is difficult for us to complete the cross-reactivity detection between SIRPa and all the analogues, therefore, cross reaction may still exist.
Sensitivity:	0.094 ng/mL
Components:	Pre-coated, ready to use 96-well strip platePlate sealer for 96 wellsStandard

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	Sample/Standard Dilution Buffer
	Biotin-labeled Antibody (Concentrated)
	Antibody Dilution Buffer
	HRP-Streptavidin Conjugate (SABC)
	SABC Dilution Buffer
	TMB Substrate
	Stop Solution
	Wash Buffer (25 x concentrate)
	Instruction manual
Material not included:	1. Microplate reader (wavelength:450nm)
	2. 37 °C incubator
	3. Automated plate washer
	4. Precision single and multi-channel pipette and disposable tips
	5. Clean tubes and Eppendorf tubes
	6. Deionized or distilled water

Target Details

Target:	SIRPA
Alternative Name:	Signal Regulatory Protein Alpha (SIRPA Products)
Background:	SIRPa, BIT, CD172a, MFR, MYD-1, SHPS1, SIRPA, Bit, BITbrain-immunoglobulin-like molecule with tyrosine-based activation motifs, Brain Ig-like molecule with tyrosine-based activation motifs, CD172 antigen-like family member A, CD172a, CD172a antigen, Inhibitory receptor SHPS-1, Macrophage fusion receptor, MFRtyrosine phosphatase SHP substRate 1, MYD1, MYD-1, MyD-1 antigen, p84, P84, protein tyrosine phosphatase, non-receptor type substRate 1, PTPNS1, SHP substRate 1, SHPS-1, SHPS1CD172A, signal-regulatory protein alpha, Signal- regulatory protein alpha-1, Signal-regulatory protein alpha-2, Signal-regulatory protein alpha-3, SIRPalpha, SIRP-ALPHA-1, Sirp-alpha-1, SIRPalpha2, Sirp-alpha-2, Sirp-alpha-3, SIRPtyrosine- protein phosphatase non-receptor type substRate 1
UniProt:	P78324

Application Details

Sample Volume:	100 µL
Plate:	Pre-coated
Protocol:	1. Wash plate 2 times before adding Standard, Sample (diluted at least 1/2 with Sample Dilution

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Sample Preparation:	 It is recommended to use fresh samples without long storage, otherwise protein degradation and denaturationmay occur in these samples, leading to false results. Samples should therefore be stored for a short periodat 2 - 8 °C or aliquoted at -20 °C (≤1 month) or -80 °C (≤ 3 months). Repeated freeze-thawcycles should be avoided. Prior to assay, the frozen samples should be slowly thawed and centrifuged toremove precipitates.
	2. Dilute the SABC with SABC Dilution Buffer at 1:100 and mix them thoroughly. (i.e. Add 1 μ L of SABC into 99 μ L of SABC Dilution Buffer.)
	(Allow 0.1-0.2 mLmore than the total volume.)
	Prepare it within 30 minutes before experiment. 1. Calculate required total volume of the working solution: 0.1ml/well x quantity of wells.
	Preparation of HRP-Streptavidin Conjugate (SABC) Working Solution: Dranara it within 20 minutes befare experiment
	thoroughly. (i.e. Add 1 μ L Biotin-labeled antibody into 99 μ L Antibody Dilution Buffer.)
	2. Dilute the Biotin-detection antibody with Antibody Dilution Buffer at 1:100 and mix them
	 Calculate required total volume of the working solution: 0.1ml/well x quantity of wells. (Allow 0.1-0.2 mLmore than the total volume.)
	Prepare it within 1 hour before experiment.
	Preparation of Biotin-labeled Antibody Working Solution:
	best to use Standard Solutions within 2 hours.
	tube and mix them thoroughly. Transfer 0.3 mL from 2nd tube to 3rd tube and mix them thoroughly, and so on. Sample Dilution Buffer was used for the blank control. Note: It is
	zero tube) into 1st tube and mix them thoroughly. Transfer 0.3 mL from 1st tube to 2nd
	the Sample Dilution Buffer into each tube. Add 0.3 mLof the above Standard solution (from
	tube concentration higher than the range of the kit,please dilute it and labeled as zero tube. 2. Label 7 EP tubes with 1/2, 1/4, 1/8, 1/16, 1/32, 1/64 and blank respectively. Add 0.3 mLof
	tube at room temperature for 10 minutes and mix them thoroughly. Note: If the standard
	1. Add 1 mL Sample Dilution Buffer into one Standard tube (labeled as zero tube), keep the
	Standards:
	bath Concentrated Wash Buffer into 750 mL Wash Buffer with deionized or distilled water. Put unused solution back at 2-8 °C.
	• Wash Buffer: If crystals have formed in the concentrate, you can warm it with 40 °C water
Reagent Preparation:	Bring all reagents and samples to room temperature for 20 minutes before use.
	γ . Add 50 μ E stop solution. Read at 400 mm initiately and calculation.
	 8. Add 90 μL TMB Substrate Solution. Incubate 10-20 minutes at 37 °C. 9. Add 50 μL Stop Solution. Read at 450nm immediately and calculation.
	7. Aspirate and wash plates 5 times.
	6. Add 100 μL SABC Working Solution into each well and incubate for 30 minutes at 37 °C.
	5. Aspirate and wash plates 3 times.
	 Add 100 μL Biotin-labeled antibody working solution to each well and incubate for 60 minutes at 37 °C.
	3. Aspirate and wash plates 2 times.
	2. Add 100 μL standard or sample to each well and incubate for 90 minutes at 37 °C.
	Buffer) and Control (blank) wells!

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	 If the sample type is not specified in the instructions, a preliminary test is necessary to determinecompatibility with the kit. If a lysis buffer is used to prepare tissue homogenates or cell culture supernatant, there is a possibility of causing a deviation due to the introduced chemical substance. The recommended dilution factor is for reference only. Please estimate the concentration of the samples before performing the test. If the values are not in therange of the standard curve, the optimal sample dilution for the particular experiment has to be determined. Samples should then be diluted with PBS (pH =7.0-7.2). Note: The user should estimate the concentration of target protein in the test sample, and select a proper dilution factor to make the diluted target protein concentration fall in the optimal detection range of the kit. Dilute the sample with the provided dilution buffer, and several trials
	may be necessary. The test sample must be well mixed with the dilution buffer. And also
	standard curves and sample should be making in pre-experiment. If samples with very high
	concentrations, dilute samples with PBS first and then dilute the samples with Sample Dilution.
	The matrix components in the sample will affect the test results, which it need to be diluted at
	least 1/2 with Sample Dilution Buffer before testing!
Assay Procedure:	Washing
	Manual: Discard the solution in the plate without touching the side walls. Clap the plate on
	absorbent filter papers or other absorbent material. Fill each well completely with 350 μL wash
	buffer and soak for 1 to 2 minutes, then aspirate contents from the plate, and clap the plate on
	buffer and soak for 1 to 2 minutes, then aspirate contents from the plate, and clap the plate on absorbent filter papers or other absorbent material.
	absorbent filter papers or other absorbent material.
	absorbent filter papers or other absorbent material. Automatic: Aspirate all wells, and then wash plate with 350 µL wash buffer. After the final wash,
	absorbent filter papers or other absorbent material. Automatic: Aspirate all wells, and then wash plate with 350 µL wash buffer. After the final wash, invert plate, and clap the plate on absorbent filter papers or other absorbent material. It is
	absorbent filter papers or other absorbent material. Automatic: Aspirate all wells, and then wash plate with 350 µL wash buffer. After the final wash, invert plate, and clap the plate on absorbent filter papers or other absorbent material. It is recommended that the washer shall be set for soaking 1 minute. (Note: set the height of the
	absorbent filter papers or other absorbent material. Automatic: Aspirate all wells, and then wash plate with 350 µL wash buffer. After the final wash, invert plate, and clap the plate on absorbent filter papers or other absorbent material. It is recommended that the washer shall be set for soaking 1 minute. (Note: set the height of the needles, be sure the fluid can be sipped up completely)
	absorbent filter papers or other absorbent material. Automatic: Aspirate all wells, and then wash plate with 350 µL wash buffer. After the final wash, invert plate, and clap the plate on absorbent filter papers or other absorbent material. It is recommended that the washer shall be set for soaking 1 minute. (Note: set the height of the needles, be sure the fluid can be sipped up completely) When diluting samples and reagents, they must be mixed completely and evenly. Before adding
	absorbent filter papers or other absorbent material. Automatic: Aspirate all wells, and then wash plate with 350 µL wash buffer. After the final wash, invert plate, and clap the plate on absorbent filter papers or other absorbent material. It is recommended that the washer shall be set for soaking 1 minute. (Note: set the height of the needles, be sure the fluid can be sipped up completely) When diluting samples and reagents, they must be mixed completely and evenly. Before adding TMB into wells, equilibrate TMB Substrate for 30 minutes at 37 °C. It is recommended to plot a
	 absorbent filter papers or other absorbent material. Automatic: Aspirate all wells, and then wash plate with 350 µL wash buffer. After the final wash, invert plate, and clap the plate on absorbent filter papers or other absorbent material. It is recommended that the washer shall be set for soaking 1 minute. (Note: set the height of the needles, be sure the fluid can be sipped up completely) When diluting samples and reagents, they must be mixed completely and evenly. Before adding TMB into wells, equilibrate TMB Substrate for 30 minutes at 37 °C. It is recommended to plot a standard curve for each test. 1. Set standard, test samples (diluted at least 1/2 with Sample Dilution Buffer), control (blank) wells on the pre-coated plate respectively, and then, records their positions. It is recommended to measure each standard and sample in duplicate. Wash plate 2 times before adding standard, sample and control (blank) wells! 2. Prepare Standards: Aliquot 100 µL of zero tube, 1sttube, 2ndtube, 3rdtube, 4thtube, 5thtube,
	 absorbent filter papers or other absorbent material. Automatic: Aspirate all wells, and then wash plate with 350 µL wash buffer. After the final wash, invert plate, and clap the plate on absorbent filter papers or other absorbent material. It is recommended that the washer shall be set for soaking 1 minute. (Note: set the height of the needles, be sure the fluid can be sipped up completely) When diluting samples and reagents, they must be mixed completely and evenly. Before adding TMB into wells, equilibrate TMB Substrate for 30 minutes at 37 °C. It is recommended to plot a standard curve for each test. 1. Set standard, test samples (diluted at least 1/2 with Sample Dilution Buffer), control (blank) wells on the pre-coated plate respectively, and then, records their positions. It is recommended to measure each standard and sample in duplicate. Wash plate 2 times before adding standard, sample and control (blank) wells!

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	 5. Wash: Remove the cover and discard the plate content, and wash plate 2 times with Wash Buffer. Do NOT let the wells dry completely at any time. 6. Biotin-labeled Antibody: Add 100 μL Biotin-labeled antibody working solution into above wells (standard, test sample and blank wells). Add the solution at the bottom of each well without touching the sidewall, cover the plate and incubate at 37 °C for 60 minutes. 7. Wash: Remove the cover, and wash plate 3 times with Wash Buffer, and let the Wash Buffer stay in the wells for 1-2 minutes each time. 8. HRP-Streptavidin Conjugate (SABC): Add 100 μL of SABC Working Solution into each well, cover the plate and incubate at 37 °C for 30 minutes. 9. Wash: Remove the cover and wash plate 5 times with Wash Buffer, and let the wash buffer stay in the wells for 1-2 minutes each time. 10. TMB Substrate: Add 90 μL TMB Substrate into each well, cover the plate and incubate at 37 °C in dark within 10-20 minutes. (Note: The reaction time can be shortened or extended
	 according to the actual color change, but not more than 30 minutes. You can terminate the reaction when apparent gradient appeared in standard wells.) 11. Stop: Add 50 µL Stop Solution into each well. The color will turn yellow immediately. The adding order of Stop Solution should be as the same as the TMB Substrate Solution. 12. OD Measurement: Read the O.D. absorbance at 450nm in Microplate Reader immediately after adding the stop solution.
	Regarding calculation, (the relative 0.D.450) = (the 0.D.450 of each well) - (the 0.D.450 of blank well). The standard curve can be plotted as the relative 0.D.450 of each standard solution (Y) vs. the respective concentration of the standard solution (X). The target concentration of the standard curve. It is recommended to use some professional software to do this calculation, such as Curve Expert 1.3 or 1.4. Note: If the samples measured were diluted, multiply the dilution factor to the concentrations from interpolation to obtain the concentration before dilution.
Assay Precision:	Intra-Assay: CV<8% Inter-Assay: CV<10%
Restrictions:	For Research Use only

Handling

Storage:	4 °C,-20 °C
Storage Comment:	1. For unopened kit: All the reagents should be kept according to the labels on vials. The
	Reference Standard and the 96-well stripe plate should be stored at -20 °C upon receipt while
	the other reagents should be stored at 4 °C.
	2. For used kit: When the kit is used, the remaining reagents need to be stored according to the
	above storage condition. Besides, please return the unused wells to the foil pouch containing
	the desiccant pack, and zip-seal the foil pouch.

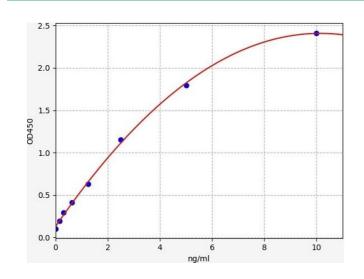
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Handling

Expiry Date:

6 months

Images



ELISA

Image 1. Typical standard curve

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