

Datasheet for ABIN6972849

anti-TCF7L1 antibody (N-Term)



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Overview

Quantity:	100 µL
Target:	TCF7L1
Binding Specificity:	N-Term
Reactivity:	Human
Host:	Rabbit
Clonality:	Polyclonal
Conjugate:	This TCF7L1 antibody is un-conjugated
Application:	Western Blotting (WB), ChIP DNA-Sequencing (ChIP-seq), Chromatin Immunoprecipitation (ChIP), Cleavage Under Targets and Release Using Nuclease (CUT&RUN)

Product Details

Immunogen:	This TCF7L1 / TCF3 antibody was raised against a peptide from the N-terminus of human TCF7L1 / TCF3.
Isotype:	IgG
Characteristics:	TCF3 / TCF7L1 (T-cell factor 3, TCF7L1) is a member of the TCF/LEF family and a component of the Wnt signaling pathway and a dominant downstream effector in embryonic stem cells (ESCs). TCF3 binds to DNA and serves as both a repressor as well as an activator of transcription. TCF3 brings developmental signals directly to the core regulatory circuitry of ES cells to influence the balance between pluripotency and differentiation. TCF3 transcriptionally represses many genes important for maintaining pluripotency and self-renewal, as well as those involved in lineage commitment and stem cell differentiation. This effect is in part mediated by the corepressors transducin-like enhancer of split 2 (TLE2) and C-terminal Binding

Product Details

Protein (CtBP). TCF7L1 / TCF3 antibody (pAb) was raised in a Rabbit host. It has been validated for use in Chromatin Immunoprecipitation, ChIP-Seq and Western blot, it has been shown to react with Human samples.

Purification: Affinity Purified

Target Details

Target: TCF7L1

Alternative Name: TCF7L1 / TCF3 ([TCF7L1 Products](#))

Molecular Weight: 80 kDa

NCBI Accession: [NP_112573](#)

Pathways: [WNT Signaling](#), [Stem Cell Maintenance](#)

Application Details

Application Notes: The rabbit anti-TC7L1 antibody ABIN6972849 is suitable for use in CUT&RUN, ChIP-seq, ChIP, and Western Blot. Specific conditions for each assay should be optimized by the end user.

General dilution recommendations for different applications are as follows:

ChIP: 10µL per ChIP

ChIP-seq: 10µL per ChIP

WB: 1:500-1:2,000

CUT&RUN: 1:100

Restrictions: For Research Use only

Handling

Buffer: Purified IgG in PBS with 30 % glycerol and 0.035 % sodium azide.

Preservative: Sodium azide

Precaution of Use: This product contains Sodium azide: a POISONOUS AND HAZARDOUS SUBSTANCE which should be handled by trained staff only.

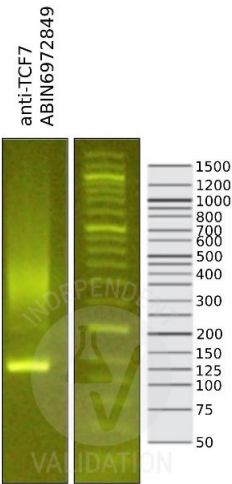
Storage: -20 °C

Storage Comment: Avoid repeated freeze/thaw cycles by aliquoting items into single-use fractions for storage at -20°C for up to 2 years. Keep all reagents on ice when not in storage.

Publications

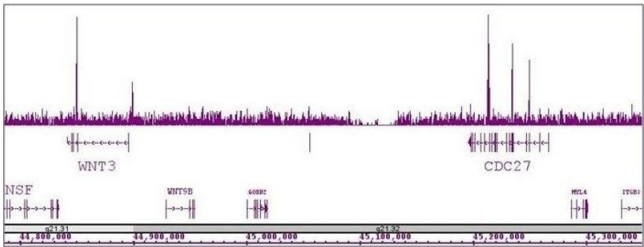
Product cited in: Zambanini, Nordin, Jonasson, Pagella, Cantù: "A new cut&run low volume-urea (LoV-U) protocol optimized for transcriptional co-factors uncovers Wnt/b-catenin tissue-specific genomic targets." in: **Development (Cambridge, England)**, (2022) ([PubMed](#)).

Images



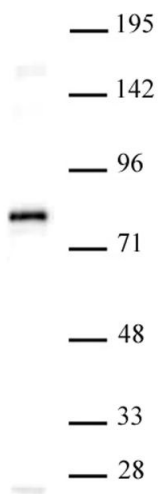
Cleavage Under Targets and Release Using Nuclease

Image 1. Library profiles comparing fragment size distributions on an E-Gel EX 2% agarose gel (Thermo Fisher). Fragments obtained from CUT&RUN using anti-TCF7L1 antibody ABIN6972849 after library preparation, compared to the E-Gel Sizing DNA Ladder (Thermo Fisher). Images provided by Gianluca Zambanini, Anna Nordin and Claudio Cantù, Gene Regulation during Development and Disease, Linköping University (<https://liu.se/en/research/cantu-lab>).



ChIP DNA-Sequencing

Image 2. TCF7L1 / TCF3 antibody (pAb) tested by ChIP-Seq. ChIP was performed using the ChIP-IT High Sensitivity Kit with 30 µg of chromatin from undifferentiated hESC cells and 7 µL of antibody. ChIP DNA was sequenced on the Illumina HiSeq and 26 million sequence tags were mapped to identify TCF7L1 / TCF3 binding sites. The image shows binding across a region of chromosome 17. You can view the complete data set in the UCSC Genome Browser, starting at this specific location, [here](#).



Western Blotting

Image 3. Western blot of TCF7L1 / TCF3 antibody. Nuclear extract of PANC-1 cells (20 µg) probed with TCF7L1 / TCF3 antibody (1:500).

Please check the [product details page](#) for more images. Overall 4 images are available for ABIN6972849.



Successfully validated (Cleavage Under Targets and Release Using Nuclease (CUT&RUN))

by [Gianluca Zambanini](#), [Anna Nordin](#) and [Claudio Cantù](#); Cantù Lab, Gene Regulation during Development and Disease, Linköping University

Report Number: 104351

Date: Feb 28 2022

Target:	TCF7L1
Lot Number:	20011001
Method validated:	Cleavage Under Targets and Release Using Nuclease (CUT&RUN)
Positive Control:	Recombinant anti-H3K27me3 CUT&RUN Positive Control antibody (antibodies-online, ABIN6923144)
Negative Control:	Polyclonal guinea Pig anti-rabbit IgG (antibodies-online, ABIN101961)
Notes:	Passed. ABIN6972849 allows for TCF7L1 targeted digestion using CUT&RUN in human HEK293T cells.
Primary Antibody:	ABIN6972849
Protocol:	- Cell harvest and nuclear extraction - Harvest 250,000 HEK293T cells per antibody to be used at RT stimulated with 10 μM CHIR for 24 h at RT. - Centrifuge cell solution 5 min at 600 x g at RT. - Remove the liquid carefully. - Gently resuspend cells in 1 mL of Nuclear Extraction Buffer (20 mM HEPES-KOH pH 8.2, 20% Glycerol, 0,05% IGEPAL, 0.5 mM Spermidine, 10 mM KCl, Roche Complete Protease Inhibitor EDTA-free). - Move the solution to a 2 mL centrifuge tube. - Pellet the nuclei 800 x g for 5 min. - Repeat the NE wash twice for a total of three washes. - Resuspend the nuclei in 20 μL NE Buffer per sample. - Concanavalin A beads preparation - Prepare one 2 mL microcentrifuge tube. - Gently resuspend the magnetic Concanavalin A Beads (antibodies-online, ABIN6952467). - Pipette 20 μL Con A Beads slurry for each sample into the 2 mL microcentrifuge tube. - Place the tube on a magnet stand until the fluid is clear. Remove the liquid carefully. - Remove the microcentrifuge tube from the magnetic stand. - Pipette 1 mL Binding Buffer (20 mM HEPES pH 7.5, 10 mM KCl, 1 mM CaCl_2, 1 mM MnCl_2) into the tube and resuspend ConA beads by gentle pipetting. - Spin down the liquid from the lid with a quick pulse in a table-top centrifuge.

- Place the tubes on a magnet stand until the fluid is clear. Remove the liquid carefully.
- Remove the microcentrifuge tube from the magnetic stand.
- Repeat the wash twice for a total of three washes.
- Gently resuspend the ConA Beads in a volume of Binding Buffer corresponding to the original volume of bead slurry, i.e. 20 µL per sample.
- Nuclei immobilization – binding to Concanavalin A beads
 - Carefully vortex the nuclei suspension and add 20 µL of the Con A beads in Binding Buffer to the cell suspension for each sample.
 - Close tube tightly incubates 10 min at 4 °C.
 - Put the 2 mL tube on the magnet stand and when the liquid is clear remove the supernatant.
 - Resuspend the beads in 1 mL of EDTA Wash buffer (20 mM HEPES pH 7.5, 150 mM NaCl, 0.5 mM Spermidine, Roche Complete Protease Inhibitor EDTA-free, 2mM EDTA).
 - Incubate 5 min at RT.
 - Place the tube on the magnet stand and when the liquid is clear remove the supernatant.
 - Resuspend the beads in 200 µl of Wash Buffer (20 mM HEPES pH 7.5, 150 mM NaCl, 0.5 mM Spermidine, Roche Complete Protease Inhibitor EDTA-free) per sample.
- Primary antibody binding
 - Divide nuclei suspension into separate 200 µL PCR tubes, one for each antibody.
 - Add 2 µL antibody (anti-TCF7L1 antibody ABIN6972849, anti-H3K27me3 antibody positive control ABIN6923144, and guinea pig anti-rabbit IgG negative control antibody ABIN101961) to the respective tube, corresponding to a 1:100 dilution.
 - Incubate at 4 °C ON.
 - Place the tubes on a magnet stand until the fluid is clear. Remove the liquid carefully.
 - Remove the microcentrifuge tubes from the magnetic stand.
 - Wash with 200 µL of Wash Buffer using a multichannel pipette to accelerate the process.
 - Repeat the wash five times for a total of six washes.
- pAG-MNase Binding
 - Prepare a 1.5 mL microcentrifuge tube containing 100 µL of pAG mix per sample (100 µL of wash buffer + 58.5 µg pAG-MNase per sample).
 - Place the PCR tubes with the sample on a magnet stand until the fluid is clear. Remove the liquid carefully.
 - Remove tubes from the magnetic stand.
 - Resuspend the beads in 100 µL of pAG-MNase premix.
 - Incubate 30 min at 4 °C.
 - Place the tubes on a magnet stand until the fluid is clear. Remove the liquid carefully.
 - Remove the microcentrifuge tubes from the magnetic stand.
 - Wash with 200 µL of Wash Buffer using a multichannel pipette to accelerate the process.
 - Repeat the wash five times for a total of six washes.
 - Resuspend in 100 µL of Wash Buffer.
- MNase digestion and release of pAG-MNase-antibody-chromatin complexes
 - Place PCR tubes on ice and allow to chill.
 - Prepare a 1.5 mL microcentrifuge tube with 102 µl of 2 mM CaCl₂ mix per sample (100 µl

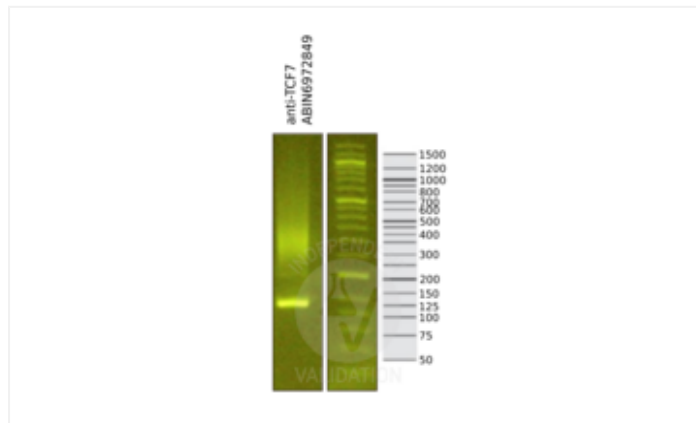
Wash Buffer + 2 µL 100 mM CaCl₂) and let it chill on ice.

- Always in ice, place the samples on the magnetic rack and when the liquid is clear remove the supernatant.
- Resuspend the samples in 100 µL of the 2 mM CaCl₂ mix and incubate in ice for exactly 30 min.
- Place the sample on the magnet stand and when the liquid is clear remove the supernatant.
- Resuspend the sample in 50 µL of 1x Urea STOP Buffer (8.5 M Urea, 100 mM NaCl, 2 mM EGTA, 2 mM EDTA, 0,5% IGEPAL).
- Incubate the samples 1h at 4°C.
- Transfer the supernatant containing the pAG-MNase-bound digested chromatin fragments to fresh 200 µL PCR tubes.
- DNA Clean up
 - Take the Mag-Bind® TotalPure NGS beads (Omega Bio-Tek, M1378-01) from the storage and wait until they are at RT.
 - Add 2x volume of beads to each sample (e.g. 100 µL of beads for 50 µL of sample).
 - Incubate the beads and the sample for 15 min at RT.
 - During incubation prepare fresh EtOH 80%.
 - Place the PCR tubes on a magnet stand and when the liquid is clear remove the supernatant.
 - Add 200 µL of fresh 80% EtOH to the sample without disturbing the beads (Important!!! Do NOT resuspend the beads or remove the tubes from the magnet stand or the sample will be lost).
 - Incubate 30 sec at RT.
 - Remove the EtOH from the sample.
 - Repeat the wash with 80% EtOH.
 - Resuspend the beads in 25 µL of 10 mM Tris.
 - Incubate the sample for 2 min at RT.
 - Repeat the 2x beads clean up as described before (this time with 50 µL of beads for each sample).
 - Resuspend the beads + DNA in 20 µL of 10 mM Tris.
- Library preparation and sequencing
 - Prepare Libraries using KAPA HyperPrep Kit using KAPA Dual-Indexed adapters according to protocol.
 - Sequence samples on an Illumina NextSeq 500 sequencer, using a NextSeq 500/550 High Output Kit v2.5 (75 Cycles), 36 bp PE.
- Peak calling
 - Trim reads using using bbTools bbdup (BBMap - Bushnell B. - sourceforge.net/projects/bbmap/) to remove adapters, artifacts and repeat sequences.
 - Map aligned reads to the hg38 human genome using bowtie with options -m 1 -v 0 -l 0 -X 500.
 - Use SAMtools to convert SAM files to BAM files and remove duplicates.
 - Use BEDtools genomecov to produce Bedgraph files.

- Call peaks using SEACR with a 0.001 threshold and the option norm stringent.

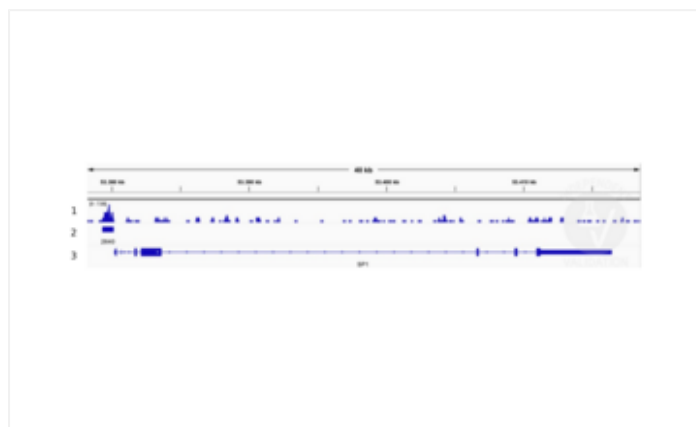
Experimental Notes: Results are published in Zambanini, G. et al. A New CUT&RUN Low Volume-Urea (LoV-U) protocol uncovers Wnt/ β -catenin tissue-specific genomic targets. bioRxiv (2022).
<https://doi.org/10.1101/2022.07.06.498999>

Images for Validation report #104351



Validation image no. 1 for anti-Transcription Factor 7-Like 1 (T-Cell Specific, HMG-Box) (TCF7L1) (N-Term) antibody (ABIN6972849)

Library profiles comparing fragment size distributions on an E-Gel EX 2% agarose gel (Thermo Fisher). Fragments obtained from CUT&RUN using anti-TCF7L1 antibody ABIN6972849 after library preparation, compared to the E-Gel Sizing DNA Ladder (Thermo Fisher).



Validation image no. 2 for anti-Transcription Factor 7-Like 1 (T-Cell Specific, HMG-Box) (TCF7L1) (N-Term) antibody (ABIN6972849)

1. Alignment tracks from CUT&RUN targeting TCF7L1 in HEK293T cells using anti-TCF7L1 antibody ABIN6972849.
2. Peaks called by SEACR from CUT&RUN data using anti-TCF7L1 antibody ABIN6972849.
3. RefSeq Genes.