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## Datasheet for ABIN6973871 PDGFA ELISA Kit

1 Image

2 Publications



#### Overview

Quantity:	96 tests
Target:	PDGFA
Reactivity:	Rat
Method Type:	Sandwich ELISA
Detection Range:	0.312 pg/mL - 20 pg/mL
Minimum Detection Limit:	0.312 pg/mL
Application:	ELISA

## Product Details

Purpose:	For the quantitative determination of rat platelet-derived growth factor A (PDGF-A) concentrations in serum, plasma.
Sample Type:	Cell Culture Supernatant, Plasma, Serum
Analytical Method:	Quantitative
Detection Method:	Colorimetric
Specificity:	This assay has high sensitivity and excellent specificity for detection of rat PDGF-A. No significant cross-reactivity or interference between rat PDGF-A and analogues was observed. Note: Limited by current skills and knowledge, it is impossible for us to complete the cross-reactivity detection between rat PDGF-A and all the analogues, therefore, cross reaction may still exist.
Sensitivity:	0.078 pg/mL

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### Product Details

#### Components:

- Assay plate
- Standard
- HRP-avidin (100 x concentrate)
- Biotin-antibody (100 x concentrate)
- Sample Diluent
- HRP-avidin Diluent
- Biotin-antibody Diluent
- Wash Buffer (25 x concentrate)
- TMB Substrate
- Stop Solution
- Adhesive Strip

## Target Details

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Target:	PDGFA
Alternative Name:	platelet-derived growth factor alpha polypeptide (PDGFA Products)
Background:	Abbreviation: PDGFA Alias: PDGF-A, PDGF1, PDGF A-chain platelet-derived growth factor alpha platelet-derived growth factor alpha chain platelet-derived growth factor alpha isoform 2 preproprotein
UniProt:	P28576
Pathways:	RTK Signaling, Fc-epsilon Receptor Signaling Pathway, EGFR Signaling Pathway, Neurotrophin Signaling Pathway, Smooth Muscle Cell Migration, Platelet-derived growth Factor Receptor Signaling

#### Application Details

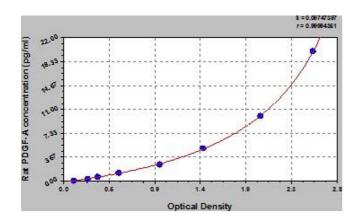
Application Notes:	Optimal working dilution should be determined by the investigator.
Sample Volume:	100 µL
Assay Time:	1 - 4.5 h
Plate:	Pre-coated
Protocol:	<ol> <li>Prepare reagents, samples and standards as instructed.</li> <li>Add 100 μL standard or sample to each well. Incubate 2 hours at 37 °C.</li> <li>Remove the liquid of each well, don't wash.</li> <li>Add 100 μL Biotin-antibody (1x) to each well. Incubate 1 hour at 37 °C.</li> <li>Aspirate and wash 3 times.</li> <li>Add 100 μL HRP-avidin (1x) to each well. Incubate 1 hour at 37 °C</li> </ol>

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	7. Aspirate and wash 5 times.
	8. Add 90 μL of TMB Substrate to each well. Incubate for 15-30 minutes at 37 °C. Protect from light.
	9. Add 50 $\mu L$ Stop Solution to each well. Read at 450 nm within 5 minutes.
Reagent Preparation:	<ol> <li>Biotin-antibody (1x) - Centrifuge the vial before opening. Biotin-antibody requires a 100-fold dilution. A suggested 100-fold dilution is 10 μL of Biotin-antibody + 990 μL of Biotin-antibody Diluent.</li> </ol>
	2. HRP-avidin (1x) - Centrifuge the vial before opening. HRP-avidin requires a 100-fold dilution. A suggested 100-fold dilution is 10 μL of HRP-avidin + 990 μL of HRP-avidin Diluent.
	3. Wash Buffer (1x) - If crystals have formed in the concentrate, warm up to room temperature and mix gently until the crystals have completely dissolved. Dilute 20 mL of Wash Buffer Concentrate (25 x) into deionized or distilled water to prepare 500 mL of Wash Buffer (1 x).
	4. Standard Centrifuge the standard vial at 6000-10000rpm for 30s. Reconstitute the Standard with 1.0 mL of Sample Diluent. Do not substitute other diluents. This reconstitution produces a stock solution of 20 pg/mL. Mix the standard to ensure complete reconstitution and allow the standard to sit for a minimum of 15 minutes with gentle agitation prior to making dilutions. Pipette 250 μL of Sample Diluent into each tube (S0-S6). Use the stock solution to produce a 2-fold dilution series (below). Mix each tube thoroughly before the next transfer. The undiluted Standard serves as the high standard (20 pg/mL). Sample Diluent serves as the zero standard (0 pg/mL).
	Note:
	<ul> <li>Kindly use graduated containers to prepare the reagent. Please don't prepare the reagent directly in the Diluent vials provided in the kit.</li> <li>Bring all reagents to room temperature (18-25 °C) before use for 30 min.</li> <li>Prepare fresh standard for each assay. Use within 4 hours and discard after use.</li> </ul>
	<ul> <li>Making serial dilution in the wells directly is not permitted.</li> <li>Please carefully reconstitute Standards according to the instruction, and avoid foaming and mix gently until the crystals have completely dissolved. To minimize imprecision caused by pipetting, use small volumes and ensure that pipettors are calibrated. It is recommended to suck more than 10 µL for once pipetting.</li> </ul>
	<ul> <li>Distilled water is recommended to be used to make the preparation for reagents.</li> <li>Contaminated water or container for reagent preparation will influence the detection result.</li> </ul>
Sample Preparation:	<ul> <li>It is recommended to use fresh samples without long storage, otherwise protein degradation and denaturationmay occur in these samples, leading to false results. Samples should therefore be stored for a short periodat 2 - 8 °C or aliquoted at -20 °C (≤1 month) or -80 °C (≤ 3 months). Repeated freeze-thawcycles should be avoided. Prior to assay, the frozen samples should be slowly thawed and centrifuged toremove precipitates.</li> <li>If the sample type is not specified in the instructions, a preliminary test is necessary to determinecompatibility with the kit.</li> </ul>
	• If a lysis buffer is used to prepare tissue homogenates or cell culture supernatant, there is a possibility of causing a deviation due to the introduced chemical substance. The

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poine A1 and A2A Decenters
tion date of the kit.
2 - 8°C. Diluent Wash Buffer
n-antibody Diluent Opened kit
- 8° C. If Biotin-antibody don't
l aluminum foil bag, plate and
iration date. May be stored for
ree samples of known
e samples of known
eriments.
The optimal dilution factor
solution to 285 µL of Sample
μL sample to 40 μL of Sample
le Diluent (1:100) before test.
forming the test. If the values e dilution for the particular uted with PBS (pH =7.0-7.2).
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#### **ELISA**

Image 1. Typical Standard Curve

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