antibodies - online.com







Chromogranin A ELISA Kit



Image



Publication



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Quantity:	96 tests
Target:	Chromogranin A (CHGA)
Reactivity:	Mouse
Method Type:	Sandwich ELISA
Detection Range:	78 pg/mL - 5000 pg/mL
Minimum Detection Limit:	78 pg/mL
Application:	ELISA
Product Details	
Purpose:	For the quantitative determination of mouse catestatin concentrations in serum, plasma, tissue
	homogenates, cell lysates.
Sample Type:	Cell Lysate, Plasma, Serum, Tissue Homogenate
Analytical Method:	Quantitative
Detection Method:	Colorimetric
Specificity:	This assay has high sensitivity and excellent specificity for detection of mouse catestatin. No
	significant cross-reactivity or interference between mouse catestatin and analogues was
	observed. Note: Limited by current skills and knowledge, it is impossible for us to complete the
	cross-reactivity detection between mouse catestatin and all the analogues, therefore, cross
	reaction may still exist.
Sensitivity:	19.5 pg/mL

Product Details

Components:

- Assay plate
- Standard
- HRP-avidin (100 x concentrate)
- Biotin-antibody (100 x concentrate)
- · Sample Diluent
- · HRP-avidin Diluent
- · Biotin-antibody Diluent
- Wash Buffer (25 x concentrate)
- · TMB Substrate
- · Stop Solution
- · Adhesive Strip

Target Details

Target:	Chromogranin A (CHGA)
Alternative Name:	catestatin (CHGA Products)
Background:	Catestatin
Pathways:	Negative Regulation of Hormone Secretion, cAMP Metabolic Process, Regulation of G-Protein Coupled Receptor Protein Signaling

Application Details

Application Notes: Optimal working dilution should be determined by the investigator. Sample Volume: 100 μL Assay Time: 1 - 4.5 h Plate: Pre-coated Protocol: 1. Prepare reagents, samples and standards as instructed. 2. Add 100 μL standard or sample to each well. Incubate 2 hours at 37 °C. 3. Remove the liquid of each well, don't wash. 4. Add 100 μL Biotin-antibody (1x) to each well. Incubate 1 hour at 37 °C. 5. Aspirate and wash 3 times. 6. Add 100 μL HRP-avidin (1x) to each well. Incubate 1 hour at 37 °C 7. Aspirate and wash 5 times. 8. Add 90 μL of TMB Substrate to each well. Incubate for 15-30 minutes at 37 °C. Protein the protection of t			
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9. Add 50 μ L Stop Solution to each well. Read at 450 nm within 5 minutes.			
Reagent Preparation: 1. Biotin-antibody (1x) - Centrifuge the vial before opening. Biotin-antibody requires a 1	1. Biotin-antibody (1x) - Centrifuge the vial before opening. Biotin-antibody requires a 100-fold		

- dilution. A suggested 100-fold dilution is 10 μ L of Biotin-antibody + 990 μ L of Biotin-antibody Diluent.
- 2. HRP-avidin (1x) Centrifuge the vial before opening. HRP-avidin requires a 100-fold dilution. A suggested 100-fold dilution is 10 μ L of HRP-avidin + 990 μ L of HRP-avidin Diluent.
- 3. Wash Buffer (1x) If crystals have formed in the concentrate, warm up to room temperature and mix gently until the crystals have completely dissolved. Dilute 20 mL of Wash Buffer Concentrate (25 x) into deionized or distilled water to prepare 500 mL of Wash Buffer (1 x).
- 4. Standard Centrifuge the standard vial at 6000-10000rpm for 30s. Reconstitute the Standard with 1.0 mL of Sample Diluent. Do not substitute other diluents. This reconstitution produces a stock solution of 5000 pg/mL. Mix the standard to ensure complete reconstitution and allow the standard to sit for a minimum of 15 minutes with gentle agitation prior to making dilutions. Pipette 250 μL of Sample Diluent into each tube (S0-S6). Use the stock solution to produce a 2-fold dilution series (below). Mix each tube thoroughly before the next transfer. The undiluted Standard serves as the high standard (5000 pg/mL). Sample Diluent serves as the zero standard (0 pg/mL).

Note:

- Kindly use graduated containers to prepare the reagent. Please don't prepare the reagent directly in the Diluent vials provided in the kit.
- Bring all reagents to room temperature (18-25 °C) before use for 30 min.
- Prepare fresh standard for each assay. Use within 4 hours and discard after use.
- · Making serial dilution in the wells directly is not permitted.
- Please carefully reconstitute Standards according to the instruction, and avoid foaming and mix gently until the crystals have completely dissolved. To minimize imprecision caused by pipetting, use small volumes and ensure that pipettors are calibrated. It is recommended to suck more than 10 µL for once pipetting.
- Distilled water is recommended to be used to make the preparation for reagents.
 Contaminated water or container for reagent preparation will influence the detection result.

Sample Preparation:

- It is recommended to use fresh samples without long storage, otherwise protein degradation and denaturationmay occur in these samples, leading to false results. Samples should therefore be stored for a short periodat 2 8 °C or aliquoted at -20 °C (≤1 month) or -80 °C (≤ 3 months). Repeated freeze-thawcycles should be avoided. Prior to assay, the frozen samples should be slowly thawed and centrifuged toremove precipitates.
- If the sample type is not specified in the instructions, a preliminary test is necessary to determine compatibility with the kit.
- If a lysis buffer is used to prepare tissue homogenates or cell culture supernatant, there is a possibility of causing a deviation due to the introduced chemical substance. The recommended dilution factor is for reference only.
- Please estimate the concentration of the samples before performing the test. If the values
 are not in therange of the standard curve, the optimal sample dilution for the particular
 experiment has to be determined. Samples should then be diluted with PBS (pH =7.0-7.2).

Assay Precision:

Intra-assay Precision (Precision within an assay): CV%<8% Three samples of known

Application Details

concentration were tested twenty times on one plate to assess.

Inter-assay Precision (Precision between assays): CV%<10% Three samples of known concentration were tested in twenty assays to assess.

Restrictions:

For Research Use only

Handling

Storage:

4 °C,-20 °C

Storage Comment:

Unopened kit Store at 2 - 8°C. Do not use the kit beyond the expiration date. May be stored for up to 1 month at 2 - 8°C. Coated assay Try to keep it in a sealed aluminum foil bag, plate and avoid the damp. Standard May be stored for up to 1 month at 2 - 8°C. If Biotin-antibody don't make recent use, better keep it store at HRP-avidin -20°C. Biotin-antibody Diluent Opened kit HRP-avidin Diluent Sample May be stored for up to 1 month at 2 - 8°C. Diluent Wash Buffer TMB Substrate Stop Solution *Provided this is within the expiration date of the kit.

Expiry Date:

6 months

Publications

Product cited in:

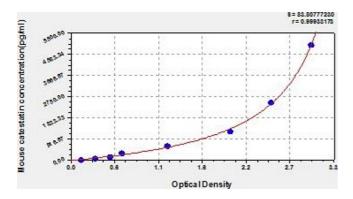
Chen, Li, Wang, Jin, Zheng, Lin, He, Zhang, Ma, Mei, Yu: "MiR-29b-3p promotes chondrocyte apoptosis and facilitates the occurrence and development of osteoarthritis by targeting PGRN." in: **Journal of cellular and molecular medicine**, Vol. 21, Issue 12, pp. 3347-3359, (2018) (PubMed).

Yang, Kang, Xing, Dou, Kang, Li, Quan, Dong: "Effect of microRNA-145 on IL-1β-induced cartilage degradation in human chondrocytes." in: **FEBS letters**, Vol. 588, Issue 14, pp. 2344-52, (2014) (PubMed).

Zheng, Zhong, Qin, Chen, Wu, Zeng, Ye, Li, Yuan, Yao, Chen: "Advanced oxidation protein products induce inflammatory response in fibroblast-like synoviocytes through NADPH oxidase -dependent activation of NF-kB." in: **Cellular physiology and biochemistry : international journal of experimental cellular physiology, biochemistry, and pharmacology**, Vol. 32, Issue 4, pp. 972-85, (2013) (PubMed).

Franchi, Torricelli, Giavaresi, Fini: "Role of moderate exercising on Achilles tendon collagen crimping patterns and proteoglycans." in: **Connective tissue research**, Vol. 54, Issue 4-5, pp. 267-74, (2013) (PubMed).

Images



ELISA

Image 1. Typical Standard Curve