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Datasheet for ABIN7041525

SAA ELISA Kit





Overview

96 tests
SAA
Horse
Sandwich ELISA
2.25 ng/mL - 72 ng/mL
2.25 ng/mL
ELISA
The kit is a sandwich enzyme immunoassay for the in vitro quantitative measurement of SAA in horse serum, plasma.
Plasma, Serum, Urine
Quantitative
Colorimetric
 ELISA Micro Plate antibody coated Detection Antibody (100X) HRP-Streptavidin (100X) Calibrator Diluent Solution Wash Solution Concentrate (20X) Chromogen - Substrate Solution STOP Solution

Product Details

Material not included:

- 1. Microplate reader with 450 ± 10nm filter.
- 2. Precision single or multi-channel pipettes and disposable tips.
- 3. Microcentrifuge tubes for diluting samples.
- 4. Squirt bottle or Microtitre washer
- 5. Deionized or distilled water.
- 6. Container for Wash Solution
- 7. Centrifuge for sample collection
- 8. Anticoagulant for plasma collection
- 9. Incubator capable of maintaining 37 °C.
- 10. Microplate shaker

Target Details

Target: SAA

Alternative Name: SAA (SAA Products)

Application Details

Sample Volume: 100 μL
Assay Time: 1.5 h

Plate: Pre-coated

Protocol:

In this assay the serum amyloid A (SAA) present in samples reacts with the anti-SAA antibodies which have been adsorbed to the surface of polystyrene microtitre wells. After the removal of unbound proteins by washing, the Detection Antibody, biotin conjugated anti-SAA, is added and complexes are formed. Following a wash step, the horseradish peroxidase (HRP) conjugated Streptavidin is added and complexes are formed. After another washing step, the complexes are assayed by the addition of a chromogenic substrate, 3,3',5,5'-tetramethylbenzidine (TMB). The quantity of bound enzyme varies directly with the concentration of SAA in the sample tested; thus, the absorbance, at 450 nm, is a measure of the concentration of SAA in the test sample. The quantity of SAA in the test sample can be interpolated from the standard curve constructed from the standards, and corrected for sample dilution.

Reagent Preparation:

- Bring all reagents to room temperature (16°C to 25°C) before use.
- · Diluent Solution Ready to use as supplied.
- Wash Solution Concentrate The Wash Solution supplied is a 20X Concentrate and must be diluted 1/20 with distilled or deionized water (1 part buffer concentrate, 19 parts dH20).
 Crystal formation in the concentrate may occur when storage temperatures are low.
 Warming of the concentrate to 30-35°Cbefore dilution can dissolve crystals.

- Detection Antibody Calculate the required amount of working conjugate solution for each microtitre plate test strip by adding 10 μ L Detection Antibody to 990 μ L of 1X Diluent for each test strip to be used for testing. Dilute immediately before use and protect from light. Mix uniformly, but gently. Avoid foaming.
- HRP-Streptavidin Calculate the required amount of working conjugate solution for each microtitre plate test strip by adding 10 µL HRP-Streptavidin to 990 µL of 1X Diluent for each test strip to be used for testing. Dilute immediately before use and protect from light. Mix uniformly, but gently. Avoid foaming.
- Pre-coated ELISA Micro Plate Ready to use as supplied. Unseal foil pouch and remove plate
 from pouch. Remove all strips and wells that will not be used in the assay and place back in
 pouch and re-seal along with desiccant.
- Calibrator Prepare according to the lot specific Certificate of Analysis.

Sample Preparation:

- It is recommended to use fresh samples without long storage, otherwise protein degradation and denaturationmay occur in these samples, leading to false results. Samples should therefore be stored for a short periodat 2 8 °C or aliquoted at -20 °C (≤1 month) or -80 °C (≤ 3 months). Repeated freeze-thawcycles should be avoided. Prior to assay, the frozen samples should be slowly thawed and centrifuged toremove precipitates.
- If the sample type is not specified in the instructions, a preliminary test is necessary to determine compatibility with the kit.
- If a lysis buffer is used to prepare tissue homogenates or cell culture supernatant, there is a possibility of causing a deviation due to the introduced chemical substance. The recommended dilution factor is for reference only.
- Please estimate the concentration of the samples before performing the test. If the values are not in therange of the standard curve, the optimal sample dilution for the particular experiment has to be determined. Samples should then be diluted with PBS (pH =7.0-7.2).

Note:

The assay requires that each test sample be diluted before use. All samples should be assayed in duplicate each time the assay is performed. The recommended dilutions are only suggestions. Dilutions should be based on the expected concentration of the unknown sample such that the diluted sample falls within the dynamic range of the standard curve. If unsure of sample level, a serial dilution with one or two representative samples before running the entire plate is highly recommended.

Serum samples – Recommended starting dilution is 1/200. To prepare a 1/200 dilution of a sample, transfer 2 μ L of sample to 398 μ L of 1X diluent. This gives you a 1/200 dilution. Mix thoroughly.

Plasma samples – Recommended starting dilution is 1/200. To prepare a 1/200 dilution of a sample, transfer 2 μ L of sample to 398 μ L of 1X diluent. This gives you a 1/200 dilution. Mix thoroughly.

Assay Procedure:

1. All samples and standards should be assayed in duplicates.

- 2. The Standards and the test sample(s) should be loaded into the ELISA wells as quickly as possible to avoid a shift in OD readings. Using a multichannel pipette would reduce this occurrence. Pipette 100 μ L of Standard 0 (0.0 ng/mL) in duplicate Standard 1 (2.25 ng/mL) in duplicate Standard 2 (4.50 ng/mL) in duplicate Standard 3 (9 ng/mL) in duplicate Standard 4 (18 ng/mL) in duplicate Standard 5 (36 ng/mL) in duplicate Standard 6 (72 ng/mL) in duplicate
- 4. Incubate the micro titer plate while shaking on a microplate shaker at 400rpm at room temperature for sixty (60 \pm 2) minutes. Keep plate covered and level during incubation.
- 5. Following incubation, aspirate the contents of the wells.

3. Pipette 100 µL of sample (in duplicate) into pre designated wells.

- 6. Completely fill each well with appropriately diluted Wash Solution and aspirate. Repeat three times, for a total of four washes. If washing manually: completely fill wells with wash buffer, invert the plate then pour/shake out the contents in a waste container. Follow this by sharply striking the wells on absorbent paper to remove residual buffer. Repeat 3 times for a total of four washes.
- 7. Pipette 100 μ L of appropriately diluted Detection Antibody to each well. Incubate while shaking on a microplate shaker at 400 rpm at room temperature for twenty (20 \pm 2) minutes. Keep plate covered in the dark and level during incubation.
- 8. Wash and blot the wells as described in Steps 5/6.
- 9. Pipette 100 μ L of appropriately diluted HRP-streptavidin to each well. Incubate while shaking on a microplate shaker at 400rpm at room temperature for twenty (20 \pm 2) minutes. Keep plate covered in the dark and level during incubation.
- 10. Wash and blot the wells as described in Steps 5/6.
- 11. Pipette 100 µL of TMB Substrate Solution into each well.
- 12. Incubate in the dark while shaking on a microplate shaker at 400rpm at room temperature for precisely ten (10) minutes. Keep plate covered in the dark and level during incubation.
- 13. After ten minutes, add 100 µL of Stop Solution to each well.
- 14. Determine the absorbance (450 nm) of the contents of each well within 30 minutes. Calibrate the plate reader to manufacturer's specifications.

Restrictions:

For Research Use only

Handling

Storage:

4°C

Storage Comment:

1. For unopened kit: All reagents should be stored according to the labels on the vials. All reagents should be stored at 4 °C for long term storage. Keep away from heat or direct

sunlight.

2. For opened kits: the remaining reagents must be stored according to the above storage conditions. In addition, please return the unused wells to the foil pouch containing the desiccant and seal the foil pouch with the zipper.

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Expiry Date:

6 months

Images

3.000 2.500 2.000 1.500 0.500 0.100 1.000 1.000 1.000 10.000 100.000 1000.000 Concentrations

ELISA

Image 1. Standard Curve Graph