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Datasheet for ABIN837930 EGFR ELISA Kit

Validation

Image



Overview

| Quantity: | 96 tests |
|--------------------------|----------------|
| Target: | EGFR |
| Reactivity: | Human |
| Method Type: | Sandwich ELISA |
| Detection Range: | 0.312-20 ng/mL |
| Minimum Detection Limit: | 0.312 ng/mL |
| Application: | ELISA |

Product Details

| Purpose: | This assay employs the quantitative sandwich enzyme immunoassay technique. |
|-----------------------------|--|
| Sample Type: | Serum, Plasma, Tissue Homogenate |
| Analytical Method: | Quantitative |
| Detection Method: | Colorimetric |
| Specificity: | This assay has high sensitivity and excellent specificity for detection of Human EGFR. |
| Cross-Reactivity (Details): | Limited by current skills and knowledge, it is impossible for us to complete the cross-reactivity detection between the target antigen and all analogues for other species. Therefore, cross reaction may still exist. |
| Sensitivity: | 0.019 ng/mL |
| Components: | Assay plate (12 × 8 coated Microwells) Standard (freeze dried) |

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- Biotin-antibody (100 × concentrate)
- HRP-avidin (100 × concentrate)
- Biotin-antibody Diluent
- HRP-avidin Diluent
- Sample Diluent
- Wash Buffer (25 × concentrate)
- TMB Substrate
- Stop Solution
- Adhesive Strip (for 96 wells)
- Instruction manual

Target Details

| Target: | EGFR |
|-------------|--|
| Abstract: | EGFR Products |
| Background: | Synonyms: ERBB, ERBB1, HER1, PIG61, mENA, avian erythroblastic leukemia viral (v-erb-b) oncogene homolog cell growth inhibiting protein 40 cell proliferation-inducing protein 61 epidermal growth factor recept |
| HGNC: | 3236 |
| UniProt: | P13387 |
| Pathways: | NF-kappaB Signaling, RTK Signaling, Fc-epsilon Receptor Signaling Pathway, EGFR Signaling Pathway, Neurotrophin Signaling Pathway, Stem Cell Maintenance, Hepatitis C, Positive Regulation of Response to DNA Damage Stimulus, Interaction of EGFR with phospholipase C- gamma, Thromboxane A2 Receptor Signaling, EGFR Downregulation, S100 Proteins |

Application Details

| Application Notes: | The supplier is only responsible for the kit itself, but not for the samples consumed during the assay. The user should calculate the possible amount of the samples used in the whole test. Please reserve sufficient samples in advance. Samples to be used within 5 days may be stored at 2-8°C, otherwise samples must be stored at -20°C (≤ 1 month) or -80°C (≤ 2 months) to avoid loss of bioactivity and contamination. Grossly hemolyzed samples are not suitable for use in this assay. If the samples are not indicated in the manual, a preliminary experiment to determine the validity of the kit is necessary. Please predict the concentration before assaying. If values for these are not within the range of the standard curve, users must determine the optimal sample dilutions for their particular experiments. |
|--------------------|---|
|--------------------|---|

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| | Tissue or cell extraction samples prepared by chemical lysis buffer may cause unexpected ELISA results due to the impacts of certain chemicals. Owing to the possibility of mismatching between antigens from another resource and antibodies used in this supplier's kits (e.g., antibody targets conformational epitope rather than linear epitope), some native or recombinant proteins from other manufacturers may not be recognized by this supplier's products. Influenced by factors including cell viability, cell number and cell sampling time, samples from cell culture supernatant may not be recognized by the kit. Fresh samples without long time storage are recommended for the test. Otherwise, protein degradation and denaturalization may occur in those samples and finally lead to wrong results. |
|----------------|---|
| Comment: | Detection wavelength: 450 nm |
| | Information on standard material: |
| | Depending on the antigen to be detected, standards can be either native or recombinant |
| | protein. The recombinant proteins are being expressed in CHO cells in most cases. Please |
| | inquire for more information. The formulation of auxiliary material in the standard is considered |
| | proprietary information, however it does not contain any poisonous substance. Proclin 300 |
| | (1:3000) is used as preservative. |
| | Information on reagents: |
| | In most cases the stop solution provided is 1 N H2SO4. The formulation of wash solution is |
| | proprietary information. None of the components contain (sodium) azide, thimerosal, 2- |
| | mercaptoethanol (2-ME) or any other poisonous materials. For the sandwich method kits, the |
| | sample diluent, antibody diluent, enzyme diluent and standard all contain BSA. |
| | Information on antibodies: |
| | The antibodies provided in different kits vary in regards to clonality and host. Some antibodies |
| | are affinity purified, some are Protein A |
| Sample Volume: | 100 µL |
| Assay Time: | 1 - 4.5 h |
| Plate: | Pre-coated |
| Protocol: | Antibody specific for EGFR has been pre-coated onto a microplate. Standards and samples are |
| | pipetted into the wells and any EGFR present is bound by the immobilized antibody. After |
| | removing any unbound substances, a biotin-conjugated antibody specific for EGFR is added to |
| | the wells. After washing, avidin conjugated Horseradish Peroxidase (HRP) is added to the wells. |

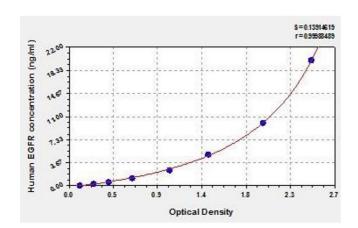
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| | Following a wash to remove any unbound avidin-enzyme reagent, a substrate solution is added |
|----------------------|--|
| | to the wells and color develops in proportion to the amount of EGFR bound in the initial step. |
| | The color development is stopped and the intensity of the color is measured. |
| Reagent Preparation: | Biotin-antibody (1×) - Centrifuge the vial before opening. Biotin-antibody requires a 100-fold dilution. The suggested dilution is 10µL of Biotin-antibody + 990µL of Biotin-antibody Diluent. HRP-avidin (1×) - Centrifuge the vial before opening. HRP-avidin requires a 100-fold dilution. The suggested dilution is 10µL of HRP-avidin + 990µL of HRP-avidin Diluent. Wash Buffer (1×) - If crystals have formed in the concentrate, warm up to room temperature and mix gently until the crystals have completely dissolved. Dilute 20mL of Wash Buffer Concentrate (25×) into deionized or distilled water to prepare 500mL of Wash Buffer (1×). Standard - Centrifuge the standard vial at 6000-10000rpm for 30s. Reconstitute the Standard with 1ml of Sample Diluent. Do not substitute other diluents. This reconstitution produces a stock solution of 200pg/mL. Mix the standard to ensure complete reconstitution and allow the standard to sit for a minimum of 15 minutes with gentle agitation prior to making dilutions. Pipette 250µL of Sample Diluent into each tube. Use the stock solution to produce a 2-fold dilution series. Mix each tube thoroughly before the next transfer. The undiluted Standard serves as the high standard (200pg/mL). Sample Diluent serves as the zero standard (0ng/mL). |
| | Note: Kindly use graduated containers to prepare the reagent. Please don't prepare the reagent directly in the Diluent vials provided in the kit. Bring all reagents to room temperature (18-25°C) before use for 30 min. Prepare fresh standard for each assay. Use within 4 hours and discard after use. Making serial dilution in the wells directly is not permitted. Please carefully reconstitute Standards according to the instruction. Avoid foaming and mix gently until the crystals have completely dissolved. To minimize imprecision caused by pipetting, use small volumes and ensure that pipettors are calibrated. It is recommended to suck more than 10µL when pipetting. It is recommended to use distilled water to prepare reagents and samples. Using contaminated water or container for reagent preparation will influence detection result. |
| Assay Precision: | Intra-assay precision (precision within an assay): Three samples of known concentration were tested twenty times on one plate to assess precision. Inter-assay precision (precision between assays): Three samples of known concentration were tested in twenty assays to assess precision. Intra-assay: CV% less than 8% |

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| Application Details | | |
|---------------------|---|--|
| Restrictions: | For Research Use only | |
| Handling | | |
| Precaution of Use: | The Stop Solution provided with this kit is an acid solution. Wear eye, hand, face and clothing protection when using this material. | |
| Handling Advice: | The kit should not be used beyond the expiration date on the kit label. Do not mix or substitute reagents with those from other lots or sources. If samples generate values higher than the highest standard, dilute the samples with Sample Diluent and repeat the assay. Any variation in Sample Diluent, operator, pipetting technique, washing technique, incubation time/temperature and kit age can cause variation in binding. This assay is designed to eliminate interference by soluble receptors, binding proteins and other factors present in biological samples. Until all factors have been tested in the Immunoassay, the possibility of interference cannot be excluded. | |
| Storage: | 4 °C/-20 °C | |
| Storage Comment: | nent: For unopened kit: All the reagents should be kept according to the labels on vials. | |
| Expiry Date: | 6 months | |

Images



ELISA

Image 1. Typical standard curve

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| | Successfully validated (ELISA (ELISA)) |
|---------------------------|---|
| | by Alamo Laboratories Inc |
| | Report Number: 029597 |
| | Date: Feb 09 2014 |
| | |
| 10: 829597 DATE: 02/89/14 | |
| ot Number: | S24095908 |
| Method validated: | ELISA (ELISA) |
| Positive Control: | Human individual postmenopausal female serum |
| Negative Control: | Goat serum |
| Notes: | Signal was detected in the positive control sample and not in the negative control sample. |
| Primary Antibody: | - Antigen: Human epidermal growth factor receptor, EGFR - Catalog Number: ABIN837930 - |
| | Supplier: Cusabio - Supplier catalog number: CSB-E12124h - Lot Number: S24095908 |
| Controls: | Positive control: Serum from normal adult human (specimen known to contain the target protein). |
| | Negative control: Serum from normal goat (specimens known to not contain the target protein). |
| | Standard curve: Serial two-fold dilutions from 20 ng/ml [20, 10, 5, 2.5, 1.25, 0.625, 0.312, 0] |
| | were generated from the standard provided in the kit using standard/sample diluent buffer. |
| | • Spike control: Standard diluted in standard/PBS diluent buffer [1.25 and 0]. |
| Protocol: | All reagents in the ELISA kit were brought up to room temperature (RT) before use. |
| | 100 µL of standard or sample were added to wells in ELISA plate pre-coated with capture antibody. All samples and standards were assayed in triplicate. |
| | The plate was covered with sealer (provided in kit) and incubated for 2 hours at 37°C. |
| | Unbound material was aspirated but the wells were NOT Washed. |
| | • 100 µL of Biotin-Antibody (diluted 1:100 in "Biotin-Antibody Diluent") was added to each well. |
| | Plate was covered with sealer (provided in kit) and incubated for 1 hour at 37°C. Unbound |
| | Biotin-Antibody was removed from each well and plate was washed three times with 350 μ L |
| | of wash buffer (provided in the kit). After the last wash the plate was inverted against clean |
| | absorbent paper to remove any remaining liquid. |
| | • 100 μ L of HRP-Avidin Conjugate (diluted 1:100 in "HRP-Avidin Diluent") was added to each |
| | well. Plate was covered with sealer (provided in kit) and incubated for 1 hour at 37°C. |
| | |
| | |
| | |
| | Unbound HRP-Avidin was removed by washing five times with 350 μL of wash by (provided in the kit). After the last wash the plate was inverted and blotted against absorbent paper to remove any remaining liquid. 90 μL of TMB substrate was added to wells and the plate was covered with a new plate was covered with a new plate was added to wells and the plate was covered with a new plate was added to wells and the plate was covered with a new plate was covered with a new plate was plate was added to wells and the plate was covered with a new plate was plat |

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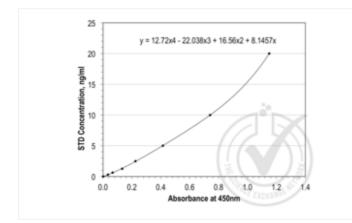
- After 30 min, when an apparent gradient appeared in the standard wells, the reaction was terminated by adding 50 µL of Stop Solution to each well.
- The optical density (OD value) of each well was read using a microplate reader set to 450 nm.
- The triplicate readings for each sample were averaged and the average zero standard optical density subtracted to yield 'corrected absorbance at 450 nm'. A standard curve was generated by plotting the mean OD value for each standard on the X-axis against the concentration on the Y-axis using Excel. Standard curve was generated by regression analysis with four-parameter logistic.
- An equation (y = 12.72x4 22.038x3 + 16.56x2 + 8.1457x) was derived from the standard curve and used to calculate Epidermal Growth Factor Receptor concentrations in samples based on their Average Absorbance values.

Experimental Notes:

Тур

None

Images for Validation report #029597



os at 450

1.308 1.371 1.375 1.30

0.0

lYr (+ve C

0.07

nglesi

1.4

0.0

EGFR conc in undiluted Serum, woman73Yr (+ve Control): 26.57 EGFR conc in Goat Serum (-ve Control) : - 0.01 ng/ml

Serum, Goat EGFR conc in 1:10 Diluted Se 5

7.18

'Out-of-range of STD

| Reading | Corrected ODesture | SD | Calculated conc ngimi | Validation image no. 2 for Epidermal Growth Factor |
|-------------------------|-------------------------|-------|-----------------------------|---|
| 1.224 | 1.150 | 0.023 | 20.00 10.00 | Receptor (EGFR) ELISA Kit (ABIN837930) |
| 0.488 0.298 0.206 | 0.414 0.224 0.132 | 0.009 | 5.02 2.44 1.32 | Table 1: ELISA. Epidermal Growth Factor Receptor is |

Table 1: ELISA. Epidermal Growth Factor Receptor is present in human serum and undetectable in goat serum. Spike controls indicate no interference in absorbance readings from the diluent used to prepare standards and sera samples. Absorbance readings (OD450 nm) are shown for standard curve, spike controls and unknown samples. Value for Avg Reading is derived from the average reading of three samples. Avg Reading for "0" amount of Standard



Figure 1: Graph of corrected OD450 nm plotted for standard curve samples.

was subtracted from all Avg Readings to yield "Corrected OD450 nm "values for Standards, spike controls and unknown samples. Standard deviation is included for all samples. Standard curve was generated by regression analysis with four-parameter logistic. An equation (y = 12.72x4 - 22.038x3 + 16.56x2 + 8.1457x) was derived from the standard curve and used to calculate Epidermal Growth Factor Receptor concentrations shown in Table 1.