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Datasheet for ABIN956145  
**TNNC1 ELISA Kit**

### Overview

Quantity: 96 tests

Target: TNNC1

Reactivity: Mouse

Method Type: Sandwich ELISA

Application: ELISA

### Product Details

Sample Type: Serum

Analytical Method: Quantitative

Detection Method: Colorimetric

Components: Anti-cTnI-coated microtiter wells, 96 wells  
Mouse cTnI Calibrator (lyophilized), reconstitute with 0.40 mL H<sub>2</sub>O  
cTnI Calibrator Diluent, 12 mL  
cTnI HRP Conjugate, 11 mL  
Wash Solution (20X), 50 mL  
TMB Reagent (HRP substrate solution), 11 mL  
Stop Solution (1N HCl), 11 mL.

Material not included: Pipettes: P-10, P-200 & P-1000 or equivalent  
Disposable pipette tips  
Distilled or de-ionized water  
Vortex mixer  
Absorbent paper

## Product Details

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Graph paper or appropriate PC graphing software  
Polypropylene microcentrifuge tubes (1.5 mL)  
Microtiter plate reader capable of reading 0 to 4 OD at 450 nm.

## Target Details

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Target:	TNNC1
Alternative Name:	Cardiac Troponin-1 ( <a href="#">TNNC1 Products</a> )
Background:	<p>Troponin is the inhibitory or contractile regulating protein complex of striated muscle. It is located periodically along the thin filament of the muscle and consists of three distinct proteins: troponin I, troponin C, and troponin T. The troponin I subunit exists in three isoforms, two in fast-twitch and slow-twitch skeletal muscle fibers, and one in cardiac muscle. At the sequence level cardiac troponin-I (cTnI) is significantly different from the skeletal isoforms and antibodies can be prepared that specifically recognize cTnI. The unique isoform and tissue specificity of cTnI are the basis for its use as a marker of cardiac muscle damage.</p>

## Application Details

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Plate:	Pre-coated
Protocol:	<p>The high sensitivity Mouse Cardiac Troponin-I ELISA recognizes an epitope on mouse cTnI that is relatively resistant to proteolysis in mouse serum, thereby improving detection capability. The assay uses two different affinity purified antibodies. One is used for solid phase immobilization (on the microtiter wells). The second is conjugated to horseradish peroxidase (HRP). The serum sample is allowed to react simultaneously with the two antibodies, resulting in cTnI being sandwiched between the solid phase and HRP-conjugated antibodies. After one hour incubation at room temperature on a plate shaker, the wells are washed to remove unbound HRP-conjugated antibodies. A solution of TMB (Tetramethylbenzidine), an HRP substrate, is then added and incubated for 20 minutes, resulting in the development of a blue color. The color development is stopped with the addition of 1N HCl changing the color to yellow. The concentration of cTnI is proportional to the absorbance at 450 nm.</p>
Sample Preparation:	<p>Serum should be prepared as quickly as possible after blood collection and stored at 4°C. All samples should be similarly processed (i.e., storage times and temperatures should be the same). If serum samples cannot be assayed immediately they should be frozen at -70°C and thawed only once prior to use.</p>
Assay Procedure:	<ol style="list-style-type: none"><li>1. Secure the desired number of coated wells in the holder.</li></ol>

2. Dispense 100  $\mu\text{L}$  of cTnI HRP Conjugate into each well.
3. Dispense 100  $\mu\text{L}$  of calibrators and samples into the appropriate wells.
4. Thoroughly mix for 10-15 seconds. It is very important to mix completely.
5. Incubate at room temperature (18-25°C) on a plate shaker (150 rpm) for one hour.
6. Remove the incubation mixture either with a plate washer or by flicking plate contents into a bio-waste container.
7. Wash and empty the microtiter wells 5 times with 1X wash solution. This may be performed using either a plate washer (400  $\mu\text{L}$ /well) or with a squirt bottle. The entire wash procedure should be performed as quickly as possible
8. Strike the wells sharply onto adsorbent paper or paper towels to remove all residual droplets.
9. Dispense 100  $\mu\text{L}$  of TMB Reagent solution into each well. Gently mix for 5 seconds.
10. Incubate on a plate shaker (150 rpm) at room temperature for 20 minutes.
11. Stop the reaction by adding 100  $\mu\text{L}$  of Stop Solution to each well.
12. Gently mix until all the blue color changes to yellow.
13. Read absorbance at 450 nm with a microtiter plate reader within 15 minutes. Please note: Due to plate reader differences, the high calibrator absorbance values may be out of range occasionally. If this occurs, absorbance values may be determined at 405 nm instead.
14. If absorbance values exceed the high calibrator, the samples should be appropriately diluted with sample diluent and re-determined. Samples with absorbance values below those of the lowest calibrator should be assigned a zero troponin-I value.

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### Calculation of Results:

1. Calculate the mean absorbance values ( $A_{450}$ ) for each set of reference calibrators and samples.
2. Construct a calibration curve by plotting the mean absorbance obtained from each reference calibrator against its concentration in ng/mL on graph paper, with absorbance values on the vertical or Y-axis and concentration on the horizontal or X-axis.
3. Using the mean absorbance value for each sample, determine the corresponding concentration of cTnI in ng/mL from the calibration curve.
4. If available, graphing software may be used to analyze the data. Depending on the range of the calibration curve used, we find that good fits of the data may be obtained with linear regression analysis or using a two-site binding model. Alternatively, calibration curves may be generated using a point-to-point fit.

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### Restrictions:

For Research Use only

## Handling

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Storage: 4 °C/-20 °C

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## Handling

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Storage Comment: The lyophilized reference calibrator should be stored at -20°C for optimum stability. The remainder of the kit should be stored refrigerated at 4°C. Keep the microtiter plate in a sealed bag with desiccant to minimize exposure to damp air. The expiration date of the kit is indicated on the box label.

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Expiry Date: The expiry date is stated on the label.