

Datasheet for ABIN956294

FABP2 ELISA Kit

[Go to Product page](#)

Overview

Quantity:	96 tests
Target:	FABP2
Reactivity:	Mouse
Method Type:	Competition ELISA
Application:	ELISA

Product Details

Purpose:	This FABP2 ELISA kit is intended for laboratory research use only and not for use in diagnostic or therapeutic procedures.
Analytical Method:	Quantitative
Detection Method:	Colorimetric
Specificity:	1. The sensitivity in this assay is 0.1 ng/mL. 2. This assay has high sensitivity and excellent specificity for detection of FABP2. No significant cross- reactivity or interference between FABP2 and analogues was observed. Note: Limited by current skills and knowledge, it is impossible for us to complete the cross-reactivity detection between FABP2 and all the analogues, therefore, cross reaction may still exist in some cases.
Characteristics:	The coated well immunoenzymatic assay for the quantitative measurement of FABP2 utilizes a polyclonal anti-FABP2 antibody and a FABP2-HRP conjugate. The assay sample and buffer are incubated together with FABP2-HRP conjugate in pre-coated plate for one hour. After the incubation period, the wells are decanted and washed five times. The wells are then incubated with a substrate for HRP enzyme. The product of the enzyme-substrate reaction forms a blue colored complex. Finally, a stop solution is added to stop the reaction, which will then turn the

Product Details

solution yellow. The intensity of color is measured spectrophotometrically at 450 nm in a microplate reader. The intensity of the color is inversely proportional to the FABP2 concentration since FABP2 from samples and FABP2-HRP conjugate compete for the anti-FABP2 antibody binding site. Since the number of sites is limited, as more sites are occupied by FABP2 from the sample, fewer sites are left to bind FABP2-HRP conjugate. Calibrators of known FABP2 concentrations are run concurrently with the samples being assayed and a calibration curve is plotted relating the intensity of the color (O.D.) to the concentration of FABP2. The FABP2 concentration is interpolated from this calibration curve.

Components:

- Microtiter Plate: 96 wells
- Calibrator 1 (0 ng/mL) 1 vial Calibrator 2 (0.5 ng/mL) 1 vial Calibrator 3 (1 ng/mL) 1 vial
Calibrator 4 (2.5 ng/mL) 1 vial Calibrator 5 (5 ng/mL) 1 vial Calibrator 6 (10 ng/mL) 1 vial
- Enzyme Conjugate (1 x 6 mL)
- Substrate A (1 x 6 mL)
- Substrate B (1 x 6 mL)
- Stop Solution (1 x 6 mL)
- Wash Buffer (100X concentrate) (1 x 6 mL)
- Lysis Buffer Solution (1 x 6 mL)
- Note: The lysis buffer solution is used only when the sample is cell culture fluid & body fluid & tissue homogenate, if the sample is serum or blood plasma, then the lysis buffer solution is a superfluous reagent.

Material not included:

1. Microplate reader capable of measuring absorbance at 450 nm.
2. Pipettes and pipette tips.
3. 100 mL and 1 liter graduated cylinders.
4. Calibrated adjustable precision pipettes, preferably with disposable plastic tips. (A manifold multi- channel pipette is desirable for large assays.)
5. 37°C incubator.
6. Absorbent paper.
7. Distilled or de-ionized water
8. Data analysis and graphing software. Graph paper: linear (Cartesian), log-log or semi-log, or log-logit as desired.
9. Tubes to prepare calibrator or sample dilutions.

Target Details

Target:

FABP2

Target Details

Alternative Name: FABP2 ([FABP2 Products](#))

Application Details

Plate:	Pre-coated
Reagent Preparation:	Bring all kit components and samples to room temperature (18-25°C) before use. Dispense 10 µL of lysis buffer solution into 100 µL specimens, mix and stand for one hour (The proportion of lysis buffer and specimens shall be no less than 1:10). (NOTE: This step is required when the sample is cell culture fluid & body fluid & tissue homogenate, if the sample is serum or blood plasma, then this step should be skipped.) Wash Solution Dilute 10 mL of Wash Solution concentrate (100X) with 990 mL of de-ionized or distilled water to prepare 1,000 mL of Wash Solution (1X).
Assay Procedure:	It is recommended that all Calibrators and Samples be added in duplicate to the Microtiter Plate. 1. Secure the desired number of coated wells in the holder then add 100 µL of Calibrators or Samples to the appropriate well of the antibody pre-coated Microtiter Plate. 2. Add 50 µL of Conjugate to each well. Mix well. Mixing well in this step is important. Cover and incubate the plate for 1 hour at 37°C. 3. Wash the Microtiter Plate using one of the specified methods indicated below: 4. Manual Washing: Remove incubation mixture by aspirating contents of the plate into a sink or proper waste container. Fill in each well completely with diluted wash solution, and then aspirate contents of the plate into a sink or proper waste container. Repeat this procedure five times for a total of five washes. After washing, invert plate, and blot dry by hitting plate onto absorbent paper or paper towels until no moisture appears. Note: Hold the sides of the plate frame firmly when washing the plate to assure that all strips remain securely in frame. Complete removal of liquid at each step is essential to good performance. 5. Automated Washing: Wash plate five times with diluted wash solution (350-400 µL/well/wash) using an auto washer. After washing, dry the plate as above. It is recommended that the washer be set for a soaking time of 10 seconds and shaking time of 5 seconds between each wash. 6. Add 50 µL Substrate A and 50 µL Substrate B to each well, subsequently. Cover and incubate for 15 minutes at 20-25°C. (Avoid sunlight). 7. Add 50 µL of stop solution to each well. Mix well. 8. Read the optical density (O.D.) at 450 nm using a microtiter plate reader immediately.
Calculation of Results:	1. This calibration curve is used to determine the amount of an unknown sample. Construct a

Application Details

- calibration curve by plotting the average O.D. (450 nm) for each calibrator on the vertical (Y) axis against the concentration on the horizontal (X) axis, and draw a best fit curve through the points on the graph.
2. First, calculate the mean O.D. value for each calibrator and sample. All O.D. values are subtracted by the mean value of the blank control before result interpretation. Construct the calibration curve using graph paper or statistical software.
3. To determine the amount in each sample, first locate the O.D. value on the Y-axis and extend a horizontal line to the calibration curve. At the point of intersection, draw a vertical line to the X-axis and read the corresponding concentration.
4. Any variation in operator, pipetting and washing technique, incubation time or temperature, and kit age can cause variation in result. Each user should obtain their own calibration curve.

Restrictions: For Research Use only

Handling

Storage: 4 °C

Storage Comment: All reagents provided are stored at 4°C. Refer to the expiration date on the label.

Expiry Date: The expiry date is stated on the label.